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*Captive breeding does not alter brain volume in a marsupial over a few generations*

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3

4 **Captive breeding does not alter brain volume in a marsupial over a few**  
5 **generations**

6

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24

25 **Abstract**

26 Captive breeding followed by re-introduction to the wild is a common component of  
27 conservation management plans for various taxa. Unfortunately, captive breeding can  
28 result in morphological changes, including brain size decrease. Brain size reduction  
29 has been associated with behavioural changes in domestic animals and such changes  
30 may negatively influence re-introduction success of captive bred animals. Many  
31 marsupials are currently bred in captivity for re-introduction yet the impacts of  
32 captive breeding on brain size have never been studied in this taxa. We investigated  
33 the impacts of a few generations (2-7) of captive breeding on brain volume in the  
34 stripe-faced dunnart (*Sminthopsis macroura*), and found that captive breeding in a  
35 relatively enriched environment did not cause any changes in brain volume.  
36 Nonetheless, we advocate that great care be taken to provide suitable husbandry  
37 conditions and to minimize the number of captive generations if marsupial re-  
38 introduction programs are to be successful.

39

40 **Keywords:** Australia; re-introduction; domestication; stripe-faced dunnart;

41 *Sminthopsis macroura*

42

43 **Introduction**

44 Captive breeding and re-introduction to the wild are major conservation and  
45 management techniques for a variety of threatened species (Blamford et al., 1996).  
46 Unfortunately, morphological and behavioural changes that may negatively impact on  
47 reintroduction success have been associated with captive breeding in various taxa  
48 (Kraaijeveld-Smit et al., 2006; Lewis and Thomas, 2001; Moore and Battley, 2006).  
49 In domestic species, a significant decrease in brain size (8-34%) compared with their  
50 wild ancestors is widespread and is associated with profound changes in behaviour  
51 (Kruska, 2005; Stahnke, 1987). Captive breeding has been suggested to correspond to  
52 the early stages of domestication and has resulted in a brain size reduction of a similar  
53 magnitude in a number of species (e.g. Guay and Iwaniuk, 2008; Röhrs and Ebinger,  
54 1998). This raises the possibility that long-term captive breeding could result in  
55 domestication and a loss of wild traits.

56

57 Most current captive breeding programs try to minimize the number of generations in  
58 captivity to decrease the risk of adaptation to captivity and domestication (McPhee,  
59 2003). Although brain size reduction has been reported over only a few generations  
60 in captivity (Runzheimer, 1969), it is not clear how many generations of captive  
61 breeding will result in a significant brain size reduction in different species.

62

63 Many marsupials are listed as either endangered or critically endangered in Australia  
64 and captive breeding has been identified as a major strategy in the conservation and  
65 management of some of these species (e.g. Wilson et al., 2003). It is thus very  
66 important to determine the effects of captive breeding on the marsupial brain.

67

68 Here we investigate the impacts of short term captive breeding (up to 7 generations)  
69 on brain size in a small dasyurid marsupial, the stripe-faced dunnart (*Sminthopsis*  
70 *macroura*), to determine the suitability of captive breeding as a source of animals for  
71 marsupial reintroductions.

72

### 73 **Methods**

#### 74 *Stripe-faced dunnart*

75 The stripe-faced dunnart is a small dasyurid marsupial that is found in the semi-arid  
76 and arid zones of central and northern Australia (Morton, 1995). Although little is  
77 known about the stripe-faced dunnart in the wild, it has been successfully maintained  
78 in long-term captive breeding colonies and has been studied extensively in captivity  
79 (Au et al., 2010; Menkhorst et al., 2007; Selwood and Woolley, 1991).

80

#### 81 *Skeletal measurements*

82 We measured 79 dunnart specimens, 43 wild and 36 captive bred. Only sexed  
83 specimens with unfractured skulls were considered. For each specimen, we measured  
84 endocranial volume using size 12 lead shot (Guay and Iwaniuk, 2008; Iwaniuk, 2001).  
85 The skull was filled with lead shot through the foramen magnum. While filling, the  
86 skull was repeatedly tapped to ensure good compaction of the shot. Once the cavity  
87 was filled, the shot were decanted and weighed to the nearest 0.01 g using a digital  
88 scale. Measurement error was estimated to be below 1% by repeated measurement (5  
89 times) for a subset of the skulls (n = 38). This is similar to the error reported by  
90 others (Iwaniuk, 2001; Marino, 1999). To transform lead shot mass into endocranial  
91 volume, we established a calibration curve by measuring the mass of various volumes

92 of shot using a graduated syringe (volume [ml] = 0.1559 X lead shot mass [g]). We  
93 also measured skull length (to the nearest 0.1mm using dial callipers).  
94  
95 Captive specimens used in this study had been lodged with Museum Victoria and  
96 were derived from a captive colony maintained by Dr. L. Selwood at La Trobe  
97 University from 1985 to 2000 (Selwood and Cui, 2006). Animals were kept as  
98 described by Woolley (1982) and were provided enrichment via running wheels and  
99 play balls in the cages and inclusion of live food (insects) in the diet. For breeding,  
100 all animals received a similar treatment irrespective of temperaments and efforts were  
101 made to pair females with unrelated or distantly related males. The dunnarts  
102 measured died between 1985 and 1992 and had been bred in captivity for 2 to 7  
103 generations.

104

#### 105 *Statistical analysis*

106 We performed two types of analyses to compare brain volume between captive and  
107 wild specimens. 1) We used analysis of variance (ANOVA) to evaluate the effect of  
108 captivity and sex on absolute brain volume and 2) we used analysis of covariance  
109 (ANCOVA) to evaluate the impact of captivity and sex on brain volume relative to  
110 body mass. The latter is necessary to control for potential changes in body size in  
111 captivity. As brain size scales allometrically with body mass (Harvey, 1988), we  
112 used body mass for our analyses of relative brain volume. Not all specimens had  
113 attached body mass data and thus we repeated the analysis using skull length as a  
114 proxy for body size. Body mass, skull length and brain volume were  $\log_{10}$   
115 transformed before analysis. All statistical analyses were performed using PASW  
116 Statistic 18 (SPSS Inc.).

117

## 118 **Results**

119 The average brain volume ( $\pm$  SE) of male and female dunnarts was 0.370ml ( $\pm$  0.007)  
120 and 0.355ml ( $\pm$  0.005) respectively. There was no differences in absolute brain  
121 volume between wild and captive specimens ( $F_{1, 75} = 0.61$ ,  $P = 0.436$ ) or between the  
122 sexes ( $F_{1, 75} = 2.55$ ,  $P = 0.114$ ). Brain volume was not correlated with body mass, but  
123 was highly correlated with skull length (Table 1). The lack of correlation between  
124 brain volume and body mass is not unexpected because that correlation is stronger at  
125 higher taxonomic levels and is often not significant intraspecifically (Martin and  
126 Harvey, 1985; Pagel and Harvey, 1989). There were no effects of captive breeding on  
127 brain volume relative to body mass or skull length, but female dunnarts had smaller  
128 brains relative to their mass than males (Table 1).

129

## 130 **Discussion**

131 Our measurements of stripe-faced dunnart brain volume are similar to those reported  
132 by Ashwell (2008). We found no difference in either absolute or relative brain  
133 volume between wild dunnarts and dunnarts that had been bred in captivity for a small  
134 (2-7) number of generations. In contrast, studies in various taxa discovered a 5-16%  
135 brain size reduction in captive bred individuals (Guay and Iwaniuk, 2008; Röhrs and  
136 Ebinger, 1998; Runzheimer, 1969). Thus, we expected stripe-faced dunnarts that  
137 have been bred in captivity to have smaller brains compared to wild specimens.  
138 Although we did not detect any changes in overall brain volume in captive-bred  
139 dunnarts, we cannot discount the possibility that various parts of the brain may have  
140 been affected by captivity without causing changes in size of the whole brain (e.g.

141 Bennett, 1976). Alternatively, 7 generations of captive breeding may be insufficient  
142 to cause brain size reduction in dunnarts.

143

144 Any reduction in brain size and correlated behavioural changes could have important  
145 effects on captive bred marsupial reintroduction since, among species, smaller brain  
146 size has been associated with lower colonization success in new habitats (Sol et al.,  
147 2008). If marsupials show similar traits, brain size reduction could potentially explain  
148 poor reintroduction success of captive-bred marsupials (Short et al., 1992).

149

150 Various strategies, including decreasing the number of generations in captivity  
151 (McPhee, 2003), and equalisation of family size (Allendorf, 1993), have been  
152 proposed to mitigate artificial selection in captivity. Providing a captive environment  
153 as similar as possible to the natural habitat has also been advocated (Frankham, 2008).  
154 Often in a zoo setting, this takes the form of environmental and behavioural  
155 enrichment (Newberry, 1995).

156

157 Overall, our results demonstrate that, in the case of the stripe-faced dunnart, captive  
158 breeding for a small number of generations does not cause brain size reduction. This  
159 suggests that captive breeding for reintroduction of marsupial mammals over a small  
160 number of generations may be appropriate and may not cause any significant  
161 reduction of overall brain size. We suggest that, through various processes including  
162 environmental enrichment and low number of captive generation, efforts must be  
163 made to ensure that captive breeding does not result in selection for adaptation to  
164 captivity as this may reduce the success of breeding colonies and reintroduction  
165 programs in marsupials (Williams and Hoffman, 2009).

166

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175 **References**

- 176 Allendorf FW. 1993. Delay of adaptation to captive breeding by equalizing family  
177 size. *Conservation Biology* 7:416-419.
- 178 Ashwell KWS. 2008. Encephalization of Australian and New Guinean marsupials.  
179 *Brain Behaviour and Evolution* 71:181-199.
- 180 Au PCK, Nation A, Parrott M, Selwood L. 2010. Induced ovulation mimics the time-  
181 table of natural development in the stripe-faced dunnart, *Sminthopsis*  
182 *macroura*, and results in the birth of fertile young. *Reproduction* 139:419-425.
- 183 Bennett EL. 1976. Cerebral effects of differential experience and training. In:  
184 Rosenzweig MR, Bennett EL, editors. *Neural Mechanisms of Learning and*  
185 *Memory*. Cambridge, USA: The MIT Press. p 279-287.
- 186 Blamford A, Mace GM, Leader-Williams N. 1996. Designing the Ark: Setting  
187 priorities for captive breeding. *Conservation Biology* 10:719-727.
- 188 Frankham R. 2008. Genetic adaptation to captivity in species conservation programs.  
189 *Molecular Ecology* 17:325-333.
- 190 Guay PJ, Iwaniuk AN. 2008. Captive breeding reduces brain size in waterfowl  
191 (Anseriformes). *Condor* 110:276-284.
- 192 Harvey PA. 1988. Allometric analysis and brain size. In: Jerison HJ, Jerison I, editors.  
193 *Intelligence and Evolutionary Biology*. Berlin, Germany: Springer-Verlag. p  
194 199-210.
- 195 Iwaniuk AN. 2001. Interspecific variation in sexual dimorphism in brain size in  
196 Nearctic ground squirrels (*Spermophilus spp.*). *Canadian Journal of Zoology*  
197 79(5):759-765.
- 198 Kraaijeveld-Smit FJL, Griffiths RA, Moore RD, Beebee TJC. 2006. Captive breeding  
199 and the fitness of reintroduced species: A test of the responses to predators in a  
200 threatened amphibian. *Journal of Applied Ecology* 43:360-365.
- 201 Kruska DCT. 2005. On the evolutionary significance of encephalization in some  
202 eutherian mammals: Effects of adaptive radiation, domestication, and  
203 feralization. *Brain, Behavior and Evolution* 65(2):73-108.

- 204 Lewis OT, Thomas CD. 2001. Adaptation to captivity in the butterfly *Pieris brassicae*  
 205 (L.) and the implications for ex situ conservation. *Journal of Insect*  
 206 *Conservation* 5:55-63.
- 207 Marino L. 1999. Brain growth in the harbor porpoise and Pacific white-sided dolphin.  
 208 *Journal of Mammalogy* 80(4):1353-1360.
- 209 Martin RD, Harvey PH. 1985. Brain size allometry: Ontogeny and phylogeny. In:  
 210 Jungers WL, editor. *Size and Scaling in Primate Biology*. New York, USA:  
 211 Plenum Press. p 147-173.
- 212 McPhee ME. 2003. Generations in captivity increases behavioral variance:  
 213 Considerations for captive breeding and reintroduction programs. *Biological*  
 214 *Conservation* 115:71-77.
- 215 Menkhorst E, Ezard N, Selwood L. 2007. Induction of ovulation and natural oestrous  
 216 cycling in the stripe-faced dunnart, *Sminthopsis macroura*. *Reproduction*  
 217 133:495-502.
- 218 Moore SJ, Battley PF. 2006. Differences in the digestive organ morphology of captive  
 219 and wild Brown Teal *Anas chlorotis* and implications for releases. *Bird*  
 220 *Conservation International* 16(3):253-264.
- 221 Morton SR. 1995. Stripe-faced dunnart *Sminthopsis macroura*. In: Strahan R, editor.  
 222 *The Mammals of Australia*. Sydney, Australia: Reed Books. p 148-149.
- 223 Newberry RC. 1995. Environmental enrichment - increasing the biological relevance  
 224 of captive environments. *Applied Animal Behaviour Science* 44:229-243.
- 225 Pagel MD, Harvey PH. 1989. Taxonomic differences in the scaling of brain on body  
 226 weight among mammals. *Science* 244:1589-1593.
- 227 Röhrs M, Ebinger P. 1998. Przewalski horses from zoological gardens: Are they  
 228 domesticated horses? *Berliner und Münchener Tierärztliche Wochenschrift*  
 229 111(7-8):273-280.
- 230 Runzheimer J. 1969. Quantitative Untersuchungen an der 5.  
 231 Gefangenschaftsgeneration von *Clethrionomys glareolus* (Schreber, 1780).  
 232 *Zeitschrift für Säugetierkunde* 34:9-37.
- 233 Selwood L, Cui S. 2006. Establishing long-term colonies of marsupial to provide  
 234 models for studying developmental mechanisms and their application to  
 235 fertility control. *Australian Journal of Zoology* 54:197-209.
- 236 Selwood L, Woolley PA. 1991. A timetable of embryonic development, and ovarian  
 237 and uterine changes during pregnancy, in the stripe-faced dunnart, *Sminthopsis*  
 238 *macroura* (Marsupialia: Dasyuridae). *Journal of Reproduction and Fertility*  
 239 91:213-227.
- 240 Short J, Bradshaw SD, Giles J, Prince RIT, Wilson GR. 1992. Reintroduction of  
 241 macropods (Marsupialia: Macropodoidea) in Australia - A review. *Biological*  
 242 *Conservation* 62:189-204.
- 243 Sol D, Bacher S, Reader SM, Lefebvre L. 2008. Brain size predicts the success of  
 244 mammals species introduced into novel environments. *American Naturalist*  
 245 172:S63-S71.
- 246 Stahnke A. 1987. Behavioral-comparison of wild and domestic cavies. *Zeitschrift für*  
 247 *Säugetierkunde* 52(5):294-307.
- 248 Williams SE, Hoffman EA. 2009. Minimizing genetic adaptation in captive breeding  
 249 programs: A review. *Biological Conservation* 142:2388-2400.
- 250 Wilson BA, Dickman CR, Fletcher TP. 2003. Dasyurid dilemmas: Problems and  
 251 solutions for conserving Australia's small carnivorous marsupials. In: Jones

252 M, Dickman C, Archer M, editors. Predators with Pouches. Collingwood,  
253 Australia: CSIRO Publishing. p 407-421.  
254 Woolley PA. 1982. The laboratory maintenance of dasyurid marsupials. In: Evans  
255 DD, editor. Management of Australian Mammals in Captivity. Melbourne,  
256 Australia: Zoological Board of Australia. p 13-21.  
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**Table 1.** Results of the ANCOVA analysis of the effects of sex and captivity on brain volume in stripe-faced dunnarts (*Sminthopsis macroura*). Presented are the *F*-ratio and the *P*-value in parenthesis. Values in bold are significant at the  $P < 0.05$  level.

	Covariates	
	Body Mass (g)	Skull length (mm)
df	1, 29	1, 71
Captivity	0.28 (0.602)	2.14 (0.148)
Sex	<b>5.85 (0.022)</b>	1.08 (0.302)
Captivity x Sex	0.46 (0.504)	0.35 (0.556)
Covariate	0.55 (0.463)	<b>73.87 (&lt;0.001)</b>

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266