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New method for determining total calcium content in tissue applied to skeletal muscle with and without calsequestrin

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We describe a new method for determining the concentration of total Ca in whole skeletal muscle samples ($[Ca_T]_{WM}$ in units of mmoles/kg wet weight) using the Ca-dependent UV absorbance spectra of the Ca chelator BAPTA (1,2-bis(2-aminophenoxy)ethane-*N,N,N',N'*-tetraacetic acid). Muscle tissue was homogenized in a solution containing 0.15 mM BAPTA and 0.5% sodium dodecyl sulfate (to permeabilize membranes and denature proteins) and then centrifuged. The solution volume was adjusted so that BAPTA captured essentially all of the Ca. $[Ca_T]_{WM}$ was obtained with Beer's law from the absorbance change produced by adding 1 mM EGTA to capture Ca from BAPTA. Results from mouse, rat, and frog muscles were reasonably consistent with results obtained using other methods for estimating total [Ca] in whole muscles and in single muscle fibers. Results with external Ca removed before determining $[Ca_T]_{WM}$ indicate that most of the Ca was intracellular, indicative of a lack of bound Ca in the extracellular space. In both fast-twitch (extensor digitorum longus, EDL) and slow-twitch (soleus) muscles from mice, $[Ca_T]_{WM}$ increased approximately linearly with decreasing muscle weight, increasing approximately twofold with a twofold decrease in muscle weight. This suggests that the Ca concentration of smaller muscles might be increased relative to that in larger muscles, thereby increasing the specific force to compensate for the smaller mass. Knocking out the high capacity Ca-binding protein calsequestrin (CSQ) did not significantly reduce $[Ca_T]_{WM}$ in mouse EDL or soleus muscle. However, in EDL muscles lacking CSQ, muscle weights were significantly lower than in wild-type (WT) muscles and the values of $[Ca_T]_{WM}$ were, on average, about half the expected WT values, taking into account the above $[Ca_T]_{WM}$ versus muscle weight relationship. Because greater reductions in $[Ca_T]_{WM}$ would be predicted in both muscle types, we hypothesize that there is a substantial increase in Ca bound to other sites in the CSQ knockout muscles.

INTRODUCTION

This Methods and Approaches article describes a new method for determining the concentration of total calcium in a tissue sample. The technique involves releasing Ca into a solution containing the Ca chelator 1,2-bis(2-aminophenoxy)ethane-*N,N,N',N'*-tetraacetic acid (BAPTA), present at a concentration severalfold greater than that needed to capture all of the released Ca. The amount of Ca bound by BAPTA is determined by the Ca-dependent change in BAPTA's UV absorbance spectrum.

One motivation for this study was to determine the physiological concentration of total Ca in whole skeletal

muscles including fast- and slow-twitch muscles from mice and rats and twitch muscles from frog. Because experiments aimed at understanding excitation–contraction coupling and its regulation and other questions involving Ca homeostasis are often performed with isolated single muscle cells or portions of such cells, such as cut fibers or skinned fibers, one reason for these measurements was to establish to what extent the range of total Ca concentrations previously published for some of these isolated preparations (e.g., Pape et al., 1995, and Fryer and Stephenson, 1996) agree with the physiological range in whole muscle reported here. As shown in the Discussion, the results with whole twitch muscles from

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Abbreviations used in this paper: AAS, atomic absorption spectroscopy; BAPTA, 1,2-bis(2-aminophenoxy)ethane-*N,N,N',N'*-tetraacetic acid; CSQ, calsequestrin; EDL, extensor digitorum longus; KO, knockout; SERCA, sarcoplasmic/endoplasmic reticulum Ca^{2+} -ATPase; TCA, trichloroacetic acid.

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frog in this study are in close agreement with the total amount of releasable Ca from the SR in cut fibers under nominally physiological conditions. However, in contrast to an earlier report that less than half of the total Ca in whole muscle from frog is intracellular, indicating a very large amount of bound Ca in the extracellular space (Kirby et al., 1975), results in this study indicate a relatively small extracellular Ca component.

A second motivation for this study relates to the current understanding of the role of calsequestrin (CSQ) as a high capacity Ca-binding protein in skeletal muscle. Starting with the discovery of CSQ by MacLennan and Wong (1971), almost all researchers have suggested that this protein acts as a major buffer of total Ca in the SR. In support of this view, based on experiments with Ca indicators introduced into voltage-clamped cut fibers from frog, Pape et al. (2007) reported that almost all of the Ca in the SR is in the bound form in resting muscle; similarly, almost all of the Ca released by a voltage-clamp step comes from the bound pool (Fénelon et al., 2012). Further, the Ca-binding curve relating bound to free Ca in the SR is largely consistent with a single, cooperative Ca-binding component. This curve was interpreted as indirect evidence that a single Ca-binding entity, presumably CSQ, is responsible for 80% or more of the Ca binding in the SR (Fénelon et al., 2012). This possibility is consistent with earlier, more direct evidence indicating that the maximal Ca-binding capacity of the SR in skinned mammalian muscle fibers is accounted for by CSQ (Murphy et al., 2009). These findings—combined with the knowledge that CSQ is localized near the SR Ca²⁺ release channels in the luminal SR, tethered there by triadin and junctin (Zhang et al., 1997)—strongly suggest that CSQ plays an important role in SR Ca²⁺ release. In apparent contradiction to this idea, the amplitude of the twitch response is about the same or slightly greater in both fast- and slow-twitch muscle with CSQ knocked out (Paolini et al., 2007, 2011).

Another question arising from studies of muscles with the skeletal isoform of CSQ (type 1 CSQ, CSQ1) knocked out is whether or not the loss of CSQ results in the expected large reduction in the Ca content in muscle. One indirect approach for assessing total releasable Ca in permeabilized single extensor digitorum longus (EDL) fibers indicated an ~70% decrease in readily releasable Ca in CSQ1-knockout (KO) fibers compared with control fibers (Fig. 8, B and E, of Paolini et al., 2007; discussed further in the Discussion). In contrast, results for EDL muscle in Table 1 of Sztretzye et al. (2011) indicated only a modest (~25%) decrease in the total SR Ca in fibers from CSQ1-KO versus WT mice. A main motivation for developing the new technique described in this Methods and Approaches article was to try to help resolve this controversy.

We found that knocking out CSQ had little effect on the concentration of total Ca in whole muscles, providing

a full or partial explanation for the lack of effect of CSQ KO on the twitch response. Decreases in negative feedback mechanisms controlling SR Ca²⁺ release, in particular Ca inactivation, would also help explain the mostly unaltered twitch response (in a manner well explained in Discussion section in Posterino and Lamb, 2003). Even with the apparently contradictory result with the twitch response, CSQ is still expected to play an important role on SR Ca²⁺ release as evidenced, in part, by faster fatigue development with CSQ knocked out (Paolini et al., 2011). In regards to the Ca-storage function of CSQ and its loss with CSQ KO, we hypothesize that this function is replaced by the up-regulation of one or more other Ca-binding proteins and/or other Ca-binding sites in the SR.

One unexpected new finding in this study was the observation of significant increases in the concentration of total Ca content with decreasing muscle weight in both fast- and slow-twitch muscles from mice. This result suggests the presence of a physiological mechanism that increases the concentration of Ca in the SR in response to increased specific work demands, thereby increasing the specific force that the muscle can generate with a twitch.

The technique described here should be useful for determining total Ca levels in almost any tissue.

A preliminary report of some of these results was presented to the Biophysical Society (Kake-Guena et al., 2014).

MATERIALS AND METHODS

Absorbance measurements (250–340 nm) were made at room temperature (22–23°C) with a spectrophotometer (Ultraspec 2000; Pharmacia). The quartz cuvette generally used had 1-cm path length, a height of 2.5 cm, and a chamber width of 3 mm, allowing a somewhat small sample volume, typically 0.45 ml. Unless indicated, experiments were performed in a solution containing 120 mM KCl and 2 mM HEPES, pH 8.0, and a nominal concentration of 0.15 mM BAPTA. The value of pH 8.0 was chosen to give a negligible value for the protonated form of BAPTA (see Fig. S1 and Sections 1 and 2 of the [supplemental text](#)). This solution is referred to as the “usual measurement solution” or the “measurement solution.” The only other solution used for the measurements was one that contained 2 mM [Mg] in addition to the constituents of the usual measurement solution (see last section of Results). The term “final solution” refers to the measurement solution with SDS and muscle added and processed as described below; muscle-sample absorbance values were obtained with this final solution.

All absorbance values shown in this Methods and Approaches article were corrected for the blank, i.e., by taking the difference between the sample absorbance and that measured with distilled water in the same cuvette. (There was no detectable difference between the absorbance values with just water and those with a solution that was the same as the usual measurement solution but without BAPTA.)

Composition of solutions other than the measurement solutions

Muscle samples were generally placed in a physiological or modified physiological solution for some period of time (usually just

briefly) before being blotted dry and weighed (see below). The mammalian Ringer's solution used for the mammalian muscle samples contained 146 mM NaCl, 5 mM KCl, 2 mM CaCl₂, 1 mM MgCl₂, and 10 mM HEPES titrated with NaOH to pH 7.4.

The composition of the normal Ringer's solution used for frog muscle is given in Table 1 of Irving et al. (1987); this solution contained 1.8 mM Ca. The 0 Ca Ringer's solution for the frog muscle contained (mM): 107 NaCl, 2.5 KCl, 2.15 Na₂HPO₄, 0.85 NaH₂PO₄, and 10 MgCl₂, pH 7.1.

Properties of Ca binding to BAPTA

The experiment described in this section gives properties of the lot of BAPTA used in this study, done in part to confirm that these properties agree with published values. Fig. 1 plots BAPTA-related absorbance at 292 nm versus different concentrations of total Ca (denoted "[Ca_T]" in addition to the usual constituents of the usual measurement solution. The different values of [Ca_T] were obtained by adding Ca from another solution containing the usual measurement solution with a known amount of total Ca (to give 100 mM CaCl₂) added from a known CaCl₂ standard. The red (solid) curve in Fig. 1 shows the least-squares best fit to the data of predicted absorbance values assuming a one-to-one binding reaction between Ca and BAPTA. The adjustable parameters for this fit were the K_d , the concentration of total BAPTA ($[B_T]$; total defined as the sum of Ca-free plus Ca-bound forms of BAPTA), and the absorbance values with no Ca present (A_{zero}) and with infinite Ca (A_{inf}), i.e., with all of the BAPTA in the Ca-bound form. An intermediate step of the fitting process was to determine values of

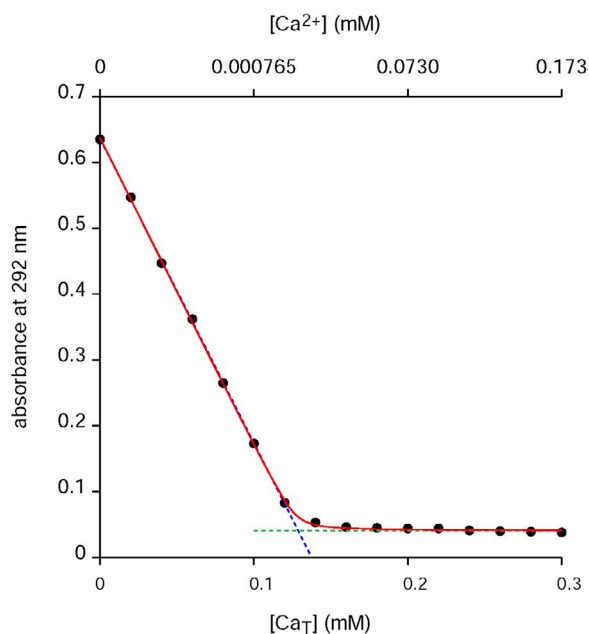


Figure 1. BAPTA absorbance at 292 nm versus $[Ca_T]$. See the description of this figure in the text for details of the experimental results and how the measured absorbance values were obtained. The text also describes how the solid red curve was obtained based on a 1:1 binding relationship for Ca to BAPTA. The best-fit value of A_{zero} associated with the red curve was 0.6375. The best-fit value of A_{inf} associated with the red curve was 0.0407, plotted as the horizontal hashed line (in green). The sloped hashed line shown (in blue) gives the least-squares best fit to the first six points. The slope and y intercept for this line are -4.625 mM^{-1} and 0.6362, respectively. The experiment ID was 821131. Experiment IDs are provided as an aid to the authors should access to the data and associated material be required.

$[Ca^{2+}]$ from the known values of $[Ca_T]$ and the adjustable parameters $[B_T]$ and K_d ; the nonlinear, horizontal axis at the top of Fig. 1 shows the values of $[Ca^{2+}]$ corresponding to those of $[Ca_T]$ shown on the bottom axis. The good fit of the red solid curve to the data in Fig. 1 is consistent with the conclusion of Tsien (1980) of a simple 1:1 stoichiometry, which he observed for the full tested ranges of BAPTA concentrations (0.01–1 mM) and concentrations of free Ca²⁺ (up to 1 mM). As $[B_T]$ was $<0.15 \text{ mM}$ for all of the experiments reported here, the simple 1:1 stoichiometry should apply to all of the results reported in this Methods and Approaches article, assuming the addition of tissue does not affect this Ca-BAPTA reaction. The extinction coefficients at 292 nm of the Ca-free and Ca-bound forms of BAPTA, ϵ_B and ϵ_{CaB} , were obtained with Beer's law from the best-fit values of A_{zero} and A_{inf} , respectively, and $[B_T]$. The best-fit values of K_d , $[B_T]$, ϵ_B , and ϵ_{CaB} were, respectively, 0.22 μM , 0.127 mM, 5,009 $\text{M}^{-1} \text{ cm}^{-1}$, and 320 $\text{M}^{-1} \text{ cm}^{-1}$. This value of K_d , which applies to the ionic strength of this study (0.12 M), is reasonably close to the value of 0.18 μM estimated from data in Harrison and Bers (1987). Because the nominal concentration of total BAPTA was 0.15 mM, the value of 0.127 mM for $[B_T]$ indicates a purity of 85% ($100 \times 0.127 \div 0.15$). This lack of 100% purity is probably caused by the presence of water, as Harrison and Bers (1987) reported a purity of 79% and determined that the impurity was caused by the presence of water. Our estimated values of 320 and 5,009 $\text{M}^{-1} \text{ cm}^{-1}$ for ϵ_{CaB} and ϵ_B , respectively, are in very good agreement with the values of 339 and 5,052 $\text{M}^{-1} \text{ cm}^{-1}$ at 292 nm estimated from Fig. 1 of Tsien (1980). Of particular interest for this Methods and Approaches article is the difference in extinction coefficients between the Ca-bound and Ca-free forms at 292 nm, denoted " $\Delta\epsilon$," given by the value $-4,689 \text{ M}^{-1} \text{ cm}^{-1}$ ($-4,689 = 320 - 5,009$). In conclusion, the properties of BAPTA estimated in this study agree with published values, and the reaction of Ca with BAPTA follows a simple 1:1 stoichiometry, at least in the absence of tissue.

The sloped dashed line shown in blue in Fig. 1 gives the least-squares best-fit line to the first six points, and the horizontal dashed line in green reflects the best-fit value of A_{inf} . As expected from the underlying relationships (not depicted), the value of $[Ca_T]$ at the intersection of these two lines is very close to the best-fit value of $[B_T]$. It is clear that the relationship between BAPTA absorbance and $[Ca_T]$ is essentially linear over the range of $[Ca_T]$ values from zero to $\sim 0.11 \text{ mM}$, which is $\sim 87\%$ of the best-fit (measurement-derived) value of 0.127 mM for $[B_T]$ given above. With the exception of the intentional addition of higher saturating levels of $[Ca_T]$, the total concentration of Ca from all sources (calcium from the muscle, the background solution, and the known amount from a Ca standard) was generally $<75\%$ of the measured concentration of BAPTA in the muscle experiments, so that the relationship between absorbance and total Ca concentration should be approximately linear for wavelengths such as 292 nm once background absorbance from muscle components is subtracted. Although the relationships derived below for determining $[Ca_T]$ in a tissue sample do not assume a linear relationship between $[Ca_T]$ and BAPTA absorbance for this range of $[Ca_T]$ values, the final results presented for $[Ca_T]$ do neglect any contribution of $[Ca^{2+}]$ to $[Ca_T]$. This is justified because the calculated values of $[Ca^{2+}]$ were no more than 0.4% of $[Ca_T]$ over this range of $[Ca_T]$ values, this maximum value occurring when $[Ca_T]$ was 62% of $[B_T]$ (from results associated with Fig. 1).

Processing of muscle samples

A whole muscle (from tendon to tendon) was removed from the animal. Unless indicated, the sample was placed in the mammalian Ringer's solution (for mice or rat) or normal Ringer's solution (for frog) for a short period (generally $<10 \text{ min}$), and then gently blotted dry and weighed. Because most experiments did not require living tissue, muscle samples were often weighed and

then frozen before further processing. The solution used in the further processing was either the usual measurement solution or the measurement solution with Mg added. For both mice and rats, soleus (slow-twitch) and EDL (fast-twitch) muscles were used. For frog, the muscles used were semitendinosus (all twitch fibers) and ileofibularis, with the latter composed of almost all twitch fibers with a very small fraction of slow fibers. Care was taken to take as little tendon as possible without damaging the muscle.

Much or all of the processing of the samples described below was performed with the samples in the tubes used for centrifugation, using weight to keep track of the total volume of measurement solution added. In the case of frog and rat muscles (ranges of muscle weights: 60–120 mg and 120–300 mg, respectively), the high density polyethylene centrifuge tubes (Sarstedt) used had volumes of 15 and 50 ml, respectively. In the case of mouse muscles (4–13 mg), 1.5-ml Eppendorf tubes (made from “homo-polymer”; MCT-150-C; Axygen) were used. As described below, background levels of Ca were determined in the same manner as described here, except with no muscle present. Early studies with polycarbonate centrifuge tubes gave background Ca levels that differed from tube-to-tube, consistent with Ca leaching from the tubes at different rates. If Ca leaching did occur with the centrifuge tubes described above used for this study, these rates were small and, more importantly, there was no detectable tube-to-tube variability so that any contribution of Ca leaching to background Ca would have been correctly estimated and subtracted.

After weighing, a muscle was homogenized in a centrifuge tube with either an electrical homogenizer (PowerGen 125 grinder with a 10×95 -mm saw tooth stator; Thermo Fischer Scientific) or a manual tissue grinder (Kontes Dual Tissue Grinder with PTFE pestle of appropriate size for the centrifuge tube; Thermo Fischer Scientific). For a muscle sample from a frog, rat, or mouse, homogenization was typically performed with, respectively, 1, 1, or 0.2 ml of measurement solution added to the muscle. In the case of frog muscle, the muscle was ground with the electric homogenizer running for 1 min at 14,000 rpm and 20 s or more at 8,000 rpm. For a rat or mouse muscle, the muscle was ground manually for 3–5 or 1–2 min, respectively. In every case, the aim was to completely homogenize the non-tendon part (i.e., fleshy part) of the muscle. After homogenization, additional measurement solution was added to give a final volume of 10 or 14 ml for frog (for ileofibularis or semitendinosus muscles, respectively), 40 ml for rat, and 0.5–1.2 ml for mouse muscle samples. For the most common muscle weights from mice, 7–13 mg, the final volume was generally 1.2 ml. For mouse muscles weighing <7 mg, the final solution volume was 0.5 or 0.6 ml, in which case the A_{∞} and the optional A_S aliquots were not done. As described below for this case, a very close estimate of the concentration of total Ca in the muscle sample could still be obtained, with an error generally <2%. Much of the measurement solution added to reach the final volume was used to rinse the material off of the grinding instrument into the centrifuge tube. The detergent SDS was added to give a concentration of 0.5% (wt/vol) in the final solution. In the case of the electric grinder, the SDS was added after the grinding to avoid excessive foaming. For the manual homogenization, 0.5% SDS was present throughout. In addition to dissolving the surface and T-system membranes, SDS greatly reduced the intrinsic absorbance associated with proteins, presumably by its ability to denature the proteins so that they were more readily removed with the next step, centrifugation. The volume of the final solution including the added SDS (denoted “ V_{solution} ” below) was determined by weight (subtracting off the previously measured weight of the centrifuge tube), assuming a density of 1 g/ml for the mostly aqueous solution at this point.

To remove proteins and other insoluble muscle components that would produce unwanted absorbance or light scattering, samples were centrifuged at 14,000 rpm for 45 min at 6°C. The

supernatant was then transferred to another tube for later use in the different aliquots described below.

It is noted that it was not considered necessary to correct V_{solution} for the part of the processed sample in which BAPTA might be excluded, namely the solid part of the muscle in the pellet after centrifugation. This excluded volume is negligible as indicated by the following calculation. Considering typical values of a mouse muscle sample weighing 10 mg and 1.2 ml for V_{solution} and assuming that ~80% of a muscle is water (Baylor et al., 1983), the pellet would contain ~2 mg of solids, giving an excluded volume of ~0.002 ml, which is <0.2% of V_{solution} .

The method assumes that BAPTA was able to complex all of the Ca, leaving none bound to muscle components and none in the pellet except for that present in any aqueous part of the pellet. Although no attempt was made to measure Ca in the pellet, calculations discussed later indicate that BAPTA should have been able to compete Ca off all muscle components with the possible exception of Ca/Mg-binding sites on parvalbumin, troponin, and myosin. Experiments to be discussed—with 2 mM Mg to mostly eliminate Ca binding to such sites—indicate that BAPTA was able to compete Ca off these sites probably because of decreases in the affinities of these proteins for Ca binding caused by the denaturing effect of SDS on proteins.

Absorbance measurements used to determine the amount of total Ca in a tissue sample

Four absorbance measurements, A_M , A_0 , A_S , and A_{∞} , were generally made with the supernatant solution after the centrifugation step described above. A_M was determined with the supernatant alone, i.e., with no further processing. For the A_0 measurement, EGTA was added to an aliquot of supernatant solution (usually added to the quartz cuvette with the same aliquot used for the A_M measurement) to capture the Ca from BAPTA. With the nominal concentration of 1 mM EGTA used for A_0 , calculations indicate that EGTA should complex >99.5% of the total Ca present (second to last section of the Appendix). For the A_S measurement, a known amount of Ca from a calcium standard was added to another aliquot of the supernatant solution (i.e., the A_M solution). A_{∞} was obtained by adding excess Ca (to give a concentration of 1 mM Ca) to another aliquot of the supernatant or to the A_S aliquot so that essentially all of the BAPTA present was in the Ca-bound form.

Assumptions and equations used for interpreting absorbance data

This section gives the equations used to analyze the absorbance data, equations based on the following assumptions:

Assumption 1. BAPTA binds essentially all of the Ca initially present in the muscle sample and any background Ca present in the solution, which includes possible Ca leached from the centrifuge tubes. We denote the corresponding concentrations of CaBAPTA from these sources as “[CaB]_M” and “[CaB]_{backgrounds},” respectively. As justified above, the level of free [Ca] is negligible compared with the Ca bound to BAPTA. In addition, with its high affinity and relative large concentration (~0.13 mM), BAPTA should be able to capture Ca from essentially all of the large-capacity, Ca-binding proteins in the muscle, including CSQ, sarcoplasmic/endoplasmic reticulum Ca²⁺-ATPase (SERCA), and the Ca-regulatory sites on troponin (see Table 2 in the first section of the Appendix and its associated text). A possible exception to this was Ca bound to the Ca/Mg-binding sites on parvalbumin and the high affinity Ca/Mg-binding sites on troponin and myosin. As shown in the last section of Results, experiments with added Mg indicate that Ca was not lost by this route, so that BAPTA did apparently capture almost all of the Ca initially present in the muscle samples.

Assumption 2. Binding of ions other than Ca to BAPTA is negligible because of the binding properties of BAPTA and the maximum expected concentration of other ions (e.g., Mg^{2+}) present in the muscle sample. This assumption is justified for protons at the value of 8 for the pH for the measurement solution (Section 1 of the supplemental text) and for Mg (last section of the Appendix, “The contribution of Mg from muscle samples is negligible”). The contribution of other possible ions should also be negligible.

Assumption 3. The reaction of Ca with BAPTA has a simple 1:1 stoichiometry in the concentration range of interest for BAPTA, i.e., 0.04–0.15 mM. (The justification for this assumption is given above.)

Assumption 4. The intrinsic absorbance of the sample (denoted “ A_{INTR} ”; see below)—defined as non-BAPTA-related absorbance—is not affected by changes in the concentration of free Ca^{2+} or by the addition of EGTA. It will be seen that A_{INTR} becomes quite pronounced at shorter wavelengths and that almost all of it is caused by muscle components remaining in the solution after the centrifugation.

Assumption 5. BAPTA and CaBAPTA do not bind to the intrinsic muscle components or any other components in the solution (specifically the pH buffer, HEPES), or such binding, if it does occur, does not affect the results.

With these assumptions, the equation for A_M is given by

$$A_M = A_{INTR} + \left\{ \begin{array}{l} \varepsilon_{CaB} ([CaB]_M + [CaB]_{background}) + \\ \varepsilon_B ([B_T] - [CaB]_M - [CaB]_{background}) \end{array} \right\} \cdot l, \quad (1)$$

where the contributions of Ca-free and Ca-bound BAPTA are given by the Beer–Lambert law, where l is the path length in the cuvette (always 1 cm), and ε_{CaB} and ε_B are the extinction coefficients of the Ca-bound and Ca-free forms of BAPTA, respectively. Likewise, with all of the BAPTA in the Ca-free form,

$$A_0 = A_{INTR} + \varepsilon_B [B_T] \cdot l, \quad (2)$$

With all of the BAPTA in the Ca-bound form

$$A_\infty = A_{INTR} + \varepsilon_{CaB} [B_T] \cdot l. \quad (3)$$

The absorbance value of the aliquot with the added standard is given by

$$A_S = A_{INTR} + \left\{ \begin{array}{l} \varepsilon_{CaB} ([CaB]_M + [CaB]_{background} + [CaB]_S) + \\ \varepsilon_B ([B_T] - [CaB]_M - [CaB]_{background} - [CaB]_S) \end{array} \right\} \cdot l, \quad (4)$$

where $[CaB]_S$ denotes the increase in the concentration of total Ca caused by the addition of Ca from the standard. With A_{INTR} assumed to be the same for all four absorbance values (Assumption 4), one obtains the following relationships:

$$\frac{A_M - A_0}{\Delta\varepsilon \cdot l} = [CaB]_M + [CaB]_{background}, \quad (5)$$

$$\frac{A_\infty - A_0}{\Delta\varepsilon \cdot l} = [B_T], \quad (6)$$

and

$$\frac{A_S - A_M}{\Delta\varepsilon \cdot l} = [CaB]_S, \quad (7)$$

where $\Delta\varepsilon \equiv \varepsilon_{CaB} - \varepsilon_B$. It is noted that $A_S - A_M$ in combination with the amount of Ca added from the standard can be used as a check on the product $\Delta\varepsilon \cdot l$. Therefore, the method does not require the use of the standard measurement. In particular, the essential value $[B_T]$ can be determined from Eq. 6 or from the following relationship obtained by combining Eqs. 6 and 7:

$$[B_T] = \frac{A_\infty - A_0}{A_S - A_M} [CaB]_S. \quad (8)$$

It is useful to express binding in terms of the fraction of BAPTA bound with Ca, denoted “ f ,” and given by

$$f \equiv \frac{[CaB]_M + [CaB]_{background}}{B_T}. \quad (9)$$

The value of f can be considered as the sum of two parts depending on the source of Ca, i.e., muscle (M) or background, given by the following relationships:

$$f = f_M + f_{background}, \quad (10)$$

where

$$f_M \equiv \frac{[CaB]_M}{[B_T]} \quad (11)$$

and

$$f_{background} \equiv \frac{[CaB]_{background}}{[B_T]}. \quad (12)$$

From Eqs. 5, 6, and 9, f is determined from the absorbance values by

$$f = \frac{A_M - A_0}{A_\infty - A_0}. \quad (13)$$

The value of $f_{background}$ was determined in the same way as f , except that muscle was not present, using the relationship

$$f_{background} = \frac{S_M - S_0}{S_\infty - S_0}, \quad (14)$$

where S (which designates solution only) replaces A to denote absorbance values with no muscle present. Because the background Ca could come from one or more of several sources—including the KCl used for the measurement solution, the detergent (SDS), and possibly the HEPES buffer and from Ca leaching from the centrifuge tubes—the absorbance measurements without muscle were obtained from solutions treated the same way as the muscle samples, including the addition of SDS followed by centrifugation. From Eqs. 10, 13, and 14,

$$f_M = f - f_{background} = \frac{A_M - A_0}{A_\infty - A_0} - \frac{S_M - S_0}{S_\infty - S_0}. \quad (15)$$

The value of $[CaB]_M$ was obtained from the relationship

$$[CaB]_M = f_M \cdot [B_T], \quad (16a)$$

where $[B_T]$ can be determined by Eq. 6 or by Eq. 8, as indicated above. Substituting Eqs. 6 and 15 into Eq. 16a gives:

$$[CaB]_M = \left[(A_M - A_0) - (S_M - S_0) \left(\frac{A_x - A_0}{S_x - S_0} \right) \right] \cdot \left(\frac{1}{\Delta\epsilon \cdot l} \right). \quad (16b)$$

As indicated later, with Eq. 18 in Results, the ratio $(A_x - A_0)/(S_x - S_0)$ gives the ratio of $[B_T]$ in the solution with the muscle sample to that without the muscle sample. As shown in Results shortly after Eq. 18, the average value for this ratio was 0.961, which is close to 1, indicating that there was almost no loss of BAPTA caused by interaction with muscle components. The main things to note from this are: (a) the $A_M - A_0$ and $S_M - S_0$ are the most important measurements, (b) A_S and S_S are not needed, and (c) A_x and S_x are almost not important at all. In fact, as seen next, a very good approximation of $[CaB]_M$ can be made without A_x and S_x .

If one assumes a value of 1 for $(A_x - A_0)/(S_x - S_0)$, a very good approximation of $[CaB]_M$ is given by

$$[CaB]_M \cong \left[(A_M - A_0) - (S_M - S_0) \right] \cdot \left(\frac{1}{\Delta\epsilon \cdot l} \right). \quad (16c)$$

This approximation was used for some measurements with small muscle samples where enough supernatant was not available to make the A_S and A_x measurements (see above). The degree to which this approximation agrees with Eq. 16b can be assessed by dividing Eq. 16c by Eq. 16b, giving:

$$\frac{[CaB]_{M \text{ from eqn 16c}}}{[CaB]_{M \text{ from eqn 16b}}} = \frac{1 - \frac{S_M - S_0}{A_M - A_0}}{1 - \frac{S_M - S_0}{A_M - A_0} \cdot \frac{A_x - A_0}{S_x - S_0}}. \quad (16d)$$

Because the ratio $(A_x - A_0)/(S_x - S_0)$ was generally <1 , the approximation generally results in an underestimation. As an example of the error, with a value of 0.3 for the ratio $(S_M - S_0)/(A_M - A_0)$, which is near the maximum measured, and with the value above of 0.961 for the ratio $(A_x - A_0)/(S_x - S_0)$, Eq. 16d gives a value of 0.984 $((1 - 0.3) \div (1 - (0.3 \times 0.961)))$, i.e., only a 1.6% error. Because the error is almost negligible, no attempt was made to correct for it, for example by assuming a value of 0.961 instead of 1 for the ratio $(A_x - A_0)/(S_x - S_0)$.

The main value of interest is the concentration of total Ca from the muscle in units of mmoles/kg (denoted " $[Ca_T]_{WM}$ ", where the subscript "WM" refers to whole muscle):

$$[Ca_T]_{WM} = \frac{V_{\text{solution}}}{W_{\text{muscle}}} [CaB]_M, \quad (17)$$

where V_{solution} is the volume of the solution and muscle combined before the centrifugation step (see description of V_{solution} above), and W_{muscle} is the wet weight of the muscle in grams (muscles blotted dry and weighed).

Correcting for small differences in BAPTA concentration in the different samples

Unless indicated, all of the absorbance values (except A_M) were corrected for small differences in BAPTA concentration, so that all of the absorbance values refer to the concentration of BAPTA in the A_M solution. As an example, for the results from muscles from mice, A_M was measured first with 0.45 ml in a cuvette followed by A_0 obtained with 9 μ l of 50 mM EGTA added to the same solution to give a final concentration of 1 mM EGTA. The A_0 data were therefore scaled by 1.02 $(1.02 = (0.45 \text{ ml} + 0.009 \text{ ml}) \div 0.45 \text{ ml})$,

so that the corrected A_0 value refers to the concentration of BAPTA that was present in A_M . When comparing absorbance measurements with muscle samples present to those without muscle samples present (Eqs. 13 and 14), a similar correction was made for the fact that the BAPTA was diluted by the volume of the muscle added. For example, with a mouse muscle weighing 8 mg added to a volume 1.2 ml of measurement solution, the volume of the muscle added is assumed to be 0.00755 ml (0.008 g divided by the assumed density of muscle, 1.06 g/ml; Mendez and Keys, 1960). To refer to the concentration of BAPTA present in the A_M measurement, the absorbance values with no muscle present (S values) were scaled by 0.994 $(0.994 = 1.2 \text{ ml} \div (1.2 \text{ ml} + 0.00755 \text{ ml}))$.

A model protocol with practical suggestions for adapting it to other tissues

The first part of this section summarizes the protocol currently used in our laboratory for a 7–13-mg mouse muscle sample (referred to below as the \sim 10-mg tissue sample). This protocol could serve as a model for adapting the BAPTA method to other size tissue samples, a process that would likely require scaling some of the variables involved if tissue samples are outside of the 7–13-mg range. Some practical suggestions are given for this process and for some of the steps in the method.

After extraction and weighing of the \sim 10-mg tissue sample, the tissue is put into a 1.5-ml Eppendorf tube. The sample is optionally frozen for later processing, with the latter all done on the same day to minimize possible slow leaching of Ca into the sample once aqueous solution has been added. For the same reason, the measurement solution with 2 mM Mg is made on the same day, and 0.5% SDS is added. About 0.2 ml of this solution is added to the Eppendorf tube, and the tissue is manually homogenized. After homogenization, more of the measurement solution with Mg and SDS is added to give a volume of \sim 1.2 ml, the actual volume (V_{solution}) determined by weight as described above. After a period of \sim 30 min to assure complete redistribution of Ca to BAPTA (no tests were performed to establish this waiting period, and it is unknown if any time beyond a few minutes is actually required), the sample is centrifuged and the supernatant is removed and placed in another Eppendorf tube. For the A_M measurement, 0.45 ml of this supernatant is introduced into the quartz cuvette (see above). For the A_0 measurement, 9 μ l of a 50-mM EGTA solution is added to the cuvette to give 1 mM EGTA. For the A_S measurement, the quartz cuvette is then thoroughly rinsed and dried, and another 0.45 ml of the supernatant is added followed by the addition of 9 μ l of a 1.5-mM $CaCl_2$ standard solution (made by multiple dilutions from a 0.1-M $CaCl_2$ standard) to give an additional 0.03 mM of Ca ($[CaB]_S$). (For reasons described later, glass Pasteur pipettes should not be used for mixing the A_S aliquot, as these introduce Ca. Our current method for mixing is to cover the cuvette with a small square of Parafilm, grasp the cuvette between fingers on the top and bottom of the cuvette, and turn it over several times. The finger pressure on the Parafilm helps seal the cuvette.) For the A_x measurement, 9 μ l of a 50-mM $CaCl_2$ standard solution is added to the cuvette with the A_S aliquot to give an additional 1 mM of Ca. Glass Pasteur pipettes can be used for mixing the A_0 and A_x aliquots because the Ca contamination would not be a problem for these aliquots.

Although absorbance measurements made over a range of wavelengths have the advantage of confirming well-behaved spectral properties (see Figs. 3–5), measurements are only needed at one wavelength. A wavelength of 292 nm would be a good choice because: (a) information about $\Delta\epsilon$ is provided in this Methods and Approaches article for this wavelength, (b) a local peak in the difference spectra occurs near this wavelength, and (c) there should be relatively little intrinsic absorbance at this wavelength (Figs. 2–5).

The main variables when considering modifying the protocol and/or adapting it to different tissue masses are the concentration

of BAPTA in the measurement solution, the volume of measurement solution added (a value very close to V_{solution}), and the size of the cuvette. (The size of the centrifuge tubes used would depend on V_{solution} .) With a 1-cm path length for the quartz cuvette, the nominal BAPTA concentration of 0.15 mM should provide reasonably high maximum absorbance values without being too high. If smaller tissue samples are required, one option would be to decrease the cuvette volume by using a quartz cuvette with a shorter path length, in which case the BAPTA concentration can be proportionally increased. Otherwise, the BAPTA concentration can be considered as a fixed variable. The only other main variable is V_{solution} . As a first estimate, V_{solution} could be chosen to give 1.2 ml per 10 mg of tissue, with V_{solution} adjusted for each tissue sample or kept constant over a range of perhaps $\pm 30\%$ as done above (we use V_{solution} values of ~ 1.2 ml for samples ranging from 7 to 13 mg). The aim would be for f_M values to be somewhere in the range of 0.1 to 0.5. The low end of the range is to provide resolution above $f_{\text{background}}$, which was typically 0.06 or less. The high end is to keep the absorbance measurements in the linear range of BAPTA absorbance versus $[\text{Ca}_T]$, as seen with Fig. 1.

Statistical significance

The difference between two sets of results was considered statistically significant if P was < 0.05 with the Student's two-tailed t test. For least-squares best-fit lines, the slopes were considered significantly different from zero with p -values < 0.05 (determined with GraphPad software package).

Online supplemental material

Section 1 of the supplemental text gives the relationships used for assessing Ca^{2+} , Mg^{2+} , and proton binding to BAPTA and EGTA. Section 2 shows the absorbance spectra of BAPTA at different pH values. In addition to yielding information about how protons interact with BAPTA, the results led to our choice of pH 8.0 for the measurement solution. Section 3 gives the relationship used for assessing stray light in the spectrophotometer and the relationship used to correct for this stray light component. Section 4 gives evidence of Ca leaching from some of the labware used in the study and results indicating that this leaching does not affect $[\text{Ca}_T]_{\text{WM}}$ values reported in this study. Section 5 gives results for the one set of mice for which dried muscle weights were determined in addition to the usual measurements. Mouse weights were also recorded with this set of mice. Unless indicated, these results were not included in the main text. Results in Section 5a indicate that the inverse relationships between $[\text{Ca}_T]_{\text{WM}}$ and wet muscle weight in Fig. 6 are not caused by differences in the fractional water content. Results in Section 5b indicate that smaller muscles are associated with fractionally greater mouse body weights. Results in Section 5c indicate

that an apparent non-reproducibility in $[\text{Ca}_T]_{\text{WM}}$ results was almost certainly caused by contamination by aluminum ions as a result of the fact that the muscles were dried on aluminum foil. Section 6 describes criteria used to reject the outlier points in this study and possible explanations for these outlier points. (Only 2 out of the almost 200 data points in this study were rejected, neither of which was in the main set of results.) Section 7 gives predicted values for $[\text{Ca}_T]_{\text{WM}}$ with CSQ KO in EDL and soleus muscles if the only effect of the KO was to remove Ca bound to CSQ. The online supplemental material is available at <http://www.jgp.org/cgi/content/full/jgp.201411250/DC1>.

RESULTS

The Materials and methods section describes how the concentration of total Ca in a muscle sample can be determined from the four absorbance measurements with the following solutions: (a) the processed muscle sample itself (A_M), the same A_M solution with (b) EGTA added so that essentially all of the Ca is displaced from BAPTA (A_0), (c) a known amount of Ca from a Ca standard added (A_S), and (d) enough Ca added so that essentially all the BAPTA is in the Ca-bound form (A_∞). Although this determination only requires information at a single wavelength, it is of interest to make measurements over a range of wavelengths to confirm that assumptions made for the determination of Ca content are valid. For example, problems with the spectra could be indicative of various possibilities including a Ca/BAPTA stoichiometry other than 1:1 (invalidating Assumption 3 in Materials and methods) and/or a substantial interaction of BAPTA with muscle components (invalidating Assumption 5), and/or Ca dependence of the absorbance of residual muscle components, components not removed by the centrifugation procedure (invalidating Assumption 4). These issues are addressed in the Figs. 2–5 by assessing UV absorbance spectra measured between 250 and 340 nm.

Test of Assumption 4 that A_{intr} is constant and EGTA lacks absorbance

Fig. 2 A shows the four absorbance spectra (A_M , A_0 , A_S , and A_∞) with an EDL muscle from a mouse processed

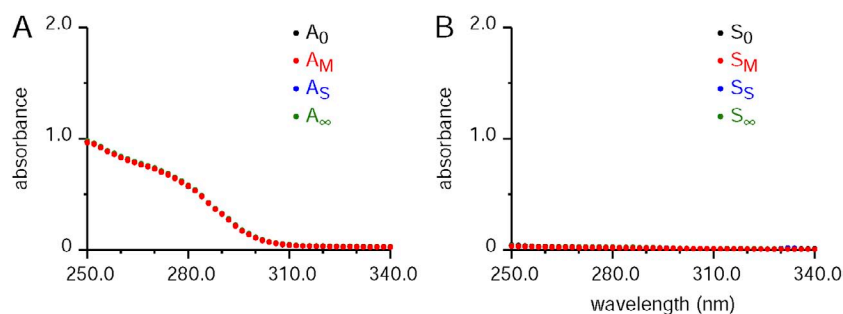


Figure 2. Absorbance spectra with no BAPTA present: justification of the assumption of constant “intrinsic” absorbance, A_{intr} . (A) Plots of the absorbance spectra for the A_0 , A_M , A_S , and A_∞ aliquots for an 8.9-mg EDL muscle from a mouse in 1.2 ml of final solution volume (including homogenized muscle) processed in the usual way, except that BAPTA was not present in the usual measurement solution. In this and B, only one set of values is visible, as the other points are covered by the last spectrum plotted (the A_M values in red for A). (B) The same format as A, with the results processed in the same way (including the addition of SDS and centrifugation), except that muscle was not present. The experiment IDs for A and B were, respectively, 310131_E2no-Ball and 310131_LnoBall.

and measured in the usual way, except that no BAPTA was present. The purpose of this experiment was to test Assumption 4 that the intrinsic absorbance (A_{intr})—arising mainly from “intrinsic” muscle components still present after the centrifugation step—was unchanged in the four conditions. Only one spectrum is clearly visible because all four spectra were almost identical. The results from this and similar experiments with muscle from other species (frog and rat) indicate that the intrinsic absorbance does not depend on the presence of EGTA used for A_0 or the presence of saturating Ca used for A_∞ , thereby verifying Assumption 4. It is noted that this result does not rule out the possibility that BAPTA interacts somehow with the remnant muscle components, a possibility that is evaluated below.

In Fig. 2, B is identical to A, except that no muscle sample was present. There was a small, nonzero absorbance at shorter wavelengths of unknown origin, but overall the absorbance values are essentially negligible compared with those attributable to muscle in A. Because the S_0 spectrum was not different from the others, the results indicate that EGTA does not exhibit absorbance over the full spectral range measured. Our unpublished results suggest that the EGTA does have significant absorbance between 250 and 280 nm when pH is close to 7, which is one reason for carrying out the experiments at pH 8.

The shapes of the uncorrected difference spectra with BAPTA present do not match over the full spectral range Fig. 3 shows typical spectra with BAPTA present. A and B were obtained with an EDL mouse muscle sample, and C and D were obtained in the same way but without muscle. The spectra of A_M , A_0 , A_S , and A_∞ are shown in A. Fig. 3 B plots the following difference spectra: $A_0 - A_\infty$ (black circles), $A_M - A_S$ (blue circles), and $A_0 - A_M$ (red circles). (Note that for clarity, absorbance differences in equations and associated text in Materials and methods were given by the absorbance with the greater level of Ca present minus that with the lower amount of Ca, whereas the inverse was used here and elsewhere in Results to give positive values. For example, $A_M - A_0$ is used in Materials and methods, whereas $A_0 - A_M$ is used in Results. This small inconvenience is because the Ca-free form of BAPTA has a higher absorbance than the Ca-bound form.) In Fig. 3 B, the $A_0 - A_\infty$ difference spectrum between 282 and 340 nm (black circles) has been least-squares best fitted with a scaled version of the $A_0 - A_M$ difference spectrum; this scaled spectrum is shown as the superimposed solid red curve. The hashed curve (between 250 and 282 nm) shows the same scaling factor applied to the rest of the $A_0 - A_M$ spectrum, i.e., from the data not included in the least-squares best fit. (This convention of solid and hashed lines, respectively, for the fitted and nonfitted parts applies to similar such fitted spectra shown in the rest of this article.)

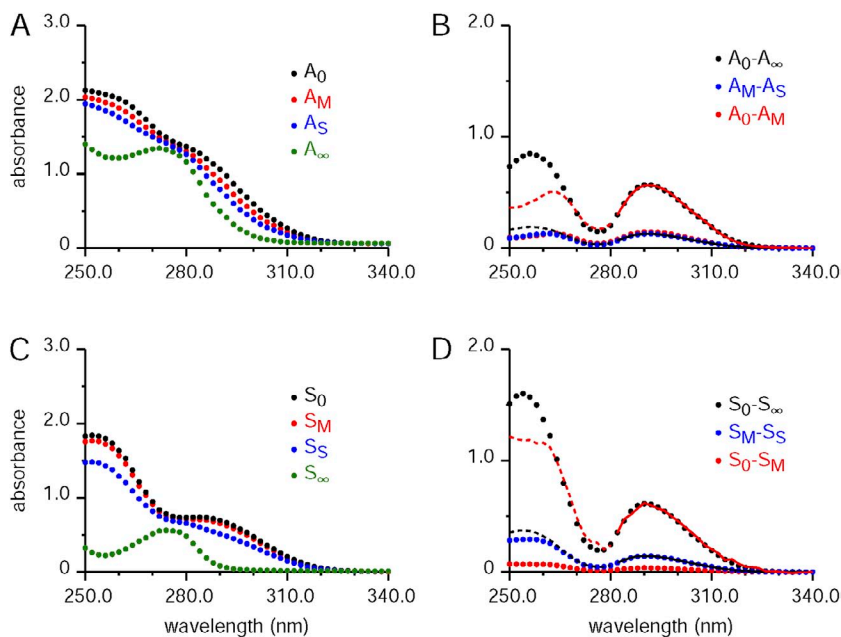


Figure 3. Absorbance spectra for a muscle sample and associated spectra without muscle for determining background Ca. The format of panels A and C is the same as Fig. 2 (A and B, respectively), except that BAPTA was present at the usual, nominal concentration of 0.15 mM. A and B were obtained with a 9.8-mg EDL muscle from a mouse in 1.36 ml of final solution volume. The Ca standard aliquot (A_S) was obtained in the usual way of adding 9 μ l of a solution containing 1.5 mM CaCl_2 made from a 1-M CaCl_2 standard to 0.45 ml of the supernatant (or A_M aliquot), giving a concentration of added standard of 0.03 mM (concentration referred to volume before adding standard). (A) As indicated, the highest to the lowest spectra for the four aliquots are in the order A_0 , A_M , A_S , and A_∞ ; this is the order because the absorbance of the Ca-free form of BAPTA is higher than that of the Ca-bound form. (B) Shows differences of spectra in A. As indicated in the text describing this figure, the solid red curve shows the least-squares best fit of the $A_0 - A_M$ difference spectra from 282 to 340 nm to the

corresponding $A_0 - A_\infty$ data. The best-fit scaling factor for the ratio $(A_0 - A_\infty)/(A_0 - A_M)$ was 3.895. For the corresponding fit of the $A_0 - A_\infty$ data to the $A_M - A_S$ difference spectrum (solid black curve), the best-fit scaling factor for the ratio $(A_M - A_S)/(A_0 - A_\infty)$ was 0.2244. The hashed lines show the part of the scaled spectra from the wavelength range not used for the fit (i.e., wavelengths below 282 nm). (C and D) Obtained in the same way as A and B, except that muscle was not present. For D, the best-fit ratios $(S_0 - S_\infty)/(S_0 - S_M)$ and $(S_M - S_S)/(S_0 - S_\infty)$ were, respectively, 17.00 and 0.2339. The experiment ID for A and B was 012214_E6, and that for C and D was 012214_LnoMg.

Although the scaled $A_0 - A_M$ difference spectrum is close to that of $A_0 - A_\infty$ of the spectral region included in the fit, it is obviously a poor fit for much of the spectral region below 282 nm. A similar poor fit is seen with the corresponding difference spectra without muscle present, i.e., the fit of $S_0 - S_M$ to $S_0 - S_\infty$ in Fig. 3 D.

The difference spectra match if corrected for stray light component

Fig. 4 shows the same absorbance spectra from Fig. 3 corrected for stray light using the procedure described in Section 3 of the supplemental text. As in Fig. 3 B, the solid red curve shows the least-squares best fit of the $A_0 - A_M$ spectrum to the $A_0 - A_\infty$ spectrum over the wavelength range of 282 to 340 nm. In contrast to Fig. 3, the $A_0 - A_M$ difference spectrum matches well with that of $A_0 - A_\infty$ over the nonfitted region of the spectrum (250–282 nm). The solid and dashed black curves give the corresponding fit of $A_0 - A_\infty$ to the $A_M - A_S$ difference spectrum. These black curves are difficult to see because, again, the fit is excellent for the full wavelength range. Likewise, the corresponding fits without muscle present in Fig. 4 D are excellent for the full measured wavelength range. Similar good matches were obtained in other experiments with uncorrected spectra if the effects of stray light were greatly decreased by decreasing the magnitude of the absorbance signals. The lower absorbance values were achieved by either diluting the muscle and BAPTA concentrations by a factor of ~ 3 or by using quartz cuvettes with shorter path lengths (1 or 3 mm) compared with that used for all of the data given in this article (path length of 1 cm).

Similarly good fits were observed over the full wavelength range for all of the difference spectra used for

the results in this article, with the exception of some small variation in the extrapolated fits near the peak at 254 nm. This variation is attributable to small variations in the stray-light component that plays a much larger role at the higher absorbance values near 254 nm. Whether the stray light component is mostly removed by reducing the overall absorbance or by the correction procedure, the good matches of the difference spectra over the measured full wavelength range indicate that the BAPTA spectra are not distorted as might occur if BAPTA were significantly bound to muscle components.

An example of the determination of the concentration of total Ca in a muscle sample

This section provides a specific example of how the concentration of total Ca in a mouse EDL muscle sample was obtained from the absorbance data with the BAPTA method, Fig. 4 (B and D) in this case. According to Eq. 13, the fraction of BAPTA complexed with Ca (f) is given by $(A_M - A_0)/(A_\infty - A_0)$ (which equals $(A_0 - A_M)/(A_0 - A_\infty)$), determined here by the reciprocal of the best-fit scaling factor relating the $A_0 - A_M$ difference spectrum to that of $A_0 - A_\infty$ in Fig. 4 B. The value of f in this case was 0.263. Correspondingly, the best fit of $S_0 - S_M$ to $S_0 - S_\infty$ in Fig. 4 D gives a value for $f_{background}$ (see Eq. 14) of 0.060. As seen in Eq. 15, the difference between f and $f_{background}$ gives f_M , the fraction of BAPTA with Ca that originated from the muscle sample, which has a value of 0.203 in this case.

The next step was to determine $[B_T]$ from Eq. 6 using the values of 1 cm for l and $-4,689 \text{ M}^{-1} \text{ cm}^{-1}$ for $\Delta\epsilon$ at 292 nm, as given in Materials and methods. The value of 0.6008 for $A_0 - A_\infty$ at this wavelength is from the point indicated by the top arrow in Fig. 4 B. The value

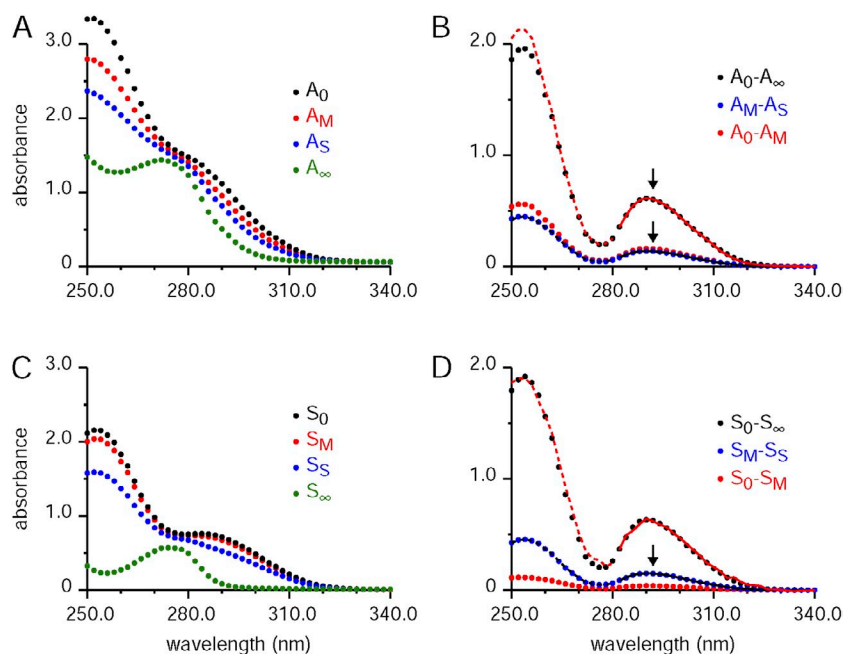


Figure 4. Absorbance spectra corrected for stray light used to determine $[Ca_T]_{WM}$ with the BAPTA method. (A–D) The absorbance spectra in this figure are from the same experiment shown in Fig. 3, except that they are corrected for the stray light component. Details of this correction are given in the text associated with this figure and Section 3 of the supplemental text. The format of the figure is the same as that used for Fig. 3. For B, the best-fit ratios $(A_0 - A_\infty)/(A_0 - A_M)$ and $(A_M - A_S)/(A_0 - A_\infty)$ were, respectively, 3.809 and 0.2275. For D, the best-fit ratios $(S_0 - S_\infty)/(S_0 - S_M)$ and $(S_M - S_S)/(S_0 - S_\infty)$ were, respectively, 16.73 and 0.2368.

of $[B_T]$ in this case is 0.128 mM ($-0.6008 \div (-4.689 \text{ mM}^{-1} \text{ cm}^{-1} \times 1 \text{ cm})$). The concentration of Ca bound to BAPTA, which originated from the muscle sample, $[CaB]_M$, is given by the product of f_M and $[B_T]$ (see Eq. 16a); $[CaB]_M$ in this case is 0.0260 mM ($0.203 \times 0.128 \text{ mM}$).

The estimated concentration of total Ca in the original muscle sample ($[Ca_T]_{WM}$; concentration referred to the whole-muscle mass) is given by Eq. 17. For this equation, the weight of the EDL muscle sample in this experiment, W_{muscle} , was 0.0098 g (9.8 mg), and the final volume of the solution with the homogenized muscle, V_{solution} , was 1.36 ml, giving a value for $[Ca_T]_{WM}$ of 3.61 mmoles/kg ($3.61 \text{ mmoles/kg} = 1.36 \text{ ml} \times 0.0260 \text{ mM} \div 0.0098 \text{ g}$).

Difference spectra match with and without muscle

Fig. 5 plots difference spectra with (symbols) and without (curves) muscle present; these are the same as four of the difference spectra in Fig. 4 (B and D). The very close matches of the forms of the difference spectra indicate that BAPTA absorbance was not altered by possible binding to muscle components. With the exception of the problematic spectral region near the higher peak at 254 nm caused by stray light (see above), similar close matches were found for all of the experiments in this study. As described near the end of Materials and methods, the difference spectra without muscle were scaled by 0.995 to take into account the dilution of the BAPTA concentration by the muscle sample so that all absorbance values refer to the concentration of BAPTA in the A_M solution. Because no other scaling factor was applied, the very close match of the magnitude of the $A_0 - A_\infty$ difference spectrum to that of $S_0 - S_\infty$ indicates that the presence of muscle components did not significantly change the estimate of the concentration of total BAPTA, $[B_T]$, using Eq. 6 above and the analogous equation for solution-only measurements. Likewise, the very

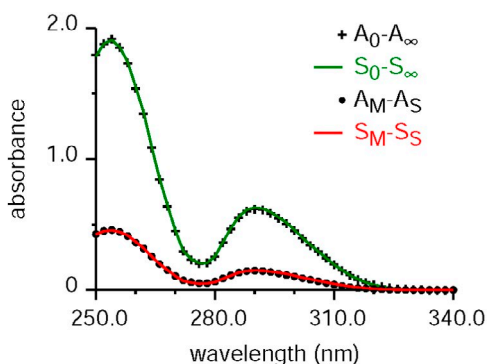


Figure 5. $[B_T]$ and the response of BAPTA to an added amount of known Ca were very similar with and without muscle present. The difference spectra plotted as symbols ($A_0 - A_\infty$ and $A_M - A_S$) were obtained with muscle present and are the same as those shown in Fig. 4 B. The difference spectra plotted as lines ($S_0 - S_\infty$ and $S_M - S_S$) were obtained without muscle present and are the same as those shown in Fig. 4 D.

close match of the magnitude of the $A_M - A_S$ difference spectrum to that of $S_M - S_S$ indicates that the presence of muscle components did not significantly change the response to the same amount of Ca added, $[CaB]_S$, as seen by Eq. 7.

The next two sections give additional details including quantification of the degree to which the magnitudes of $A_0 - A_\infty$ match $S_0 - S_\infty$ and those of $A_M - A_S$ match $S_M - S_S$ in all of the experiments in this study by comparing their values at 292 nm.

The presence of muscle has almost no effect on the measured value of $[B_T]$

From Eq. 6 and the analogous equation for solution-only measurements, the ratio of the estimated concentration of total BAPTA ($[B_T]$) with muscle present to that without muscle, denoted “ γ ,” is given by

$$\gamma = \frac{A_0 - A_\infty}{S_0 - S_\infty}. \quad (18)$$

(Values of $S_0 - S_\infty$ used with this ratio were reduced to account for the reduction of BAPTA concentration caused by the addition of muscle to the measurement solution, as described in “Correcting for small differences in BAPTA concentration...” in Materials and methods.) A value of 1 for γ would indicate that $[B_T]$ is not affected by either of the following two cases: (1) loss of BAPTA caused by binding to muscle components in the pellet after centrifugation (which would result in a decrease in $[B_T]$ and a value of $\gamma < 1$), or (2) a combination of BAPTA binding to muscle components in the supernatant and changes in BAPTA’s absorbance properties (which would also likely give a value of $\gamma < 1$). For the 148 mouse muscle samples reported in this article, the average value of γ was 0.961 (SEM = 0.002). Although significantly < 1 ($P < 0.0001$), the results indicate only a loss of 3.9% on average of the reported BAPTA concentration. A loss of BAPTA caused by case 1 (loss in the pellet) would not present a problem for the method of detecting Ca in a tissue sample, as it depends on the value of $[B_T]$ present in the supernatant (see Eqs. 6 and 16a). However, binding of BAPTA to muscle components in the supernatant (case 2) could present a problem. For example, the 3.9% loss could be caused by a much larger percentage of Ca-free BAPTA bound to muscle components if this binding were accompanied by a partial loss of its ability to absorb light (e.g., the 3.9% loss could be caused by the binding of 39% of BAPTA to muscle components with a 10% decrease in absorptivity).

The presence of muscle has almost no effect on the responsiveness of BAPTA to Ca

This section considers the change in BAPTA absorbance ($A_M - A_S$ or $S_M - S_S$) upon the addition of a presumably known amount of Ca. The important question addressed

is whether the presence of muscle affects this response and, thereby, the ability of the method to accurately detect the concentration of Ca in a muscle sample. The measured value of $[CaB]_S$ is given by Eq. 7, reproduced here in slightly different forms:

$$[CaB]_S = \frac{A_S - A_M}{\Delta\epsilon \cdot l} = -\frac{A_M - A_S}{\Delta\epsilon \cdot l}, \quad (19a)$$

or, in the absence of muscle,

$$[CaB]_S = \frac{S_S - S_M}{\Delta\epsilon \cdot l} = -\frac{S_M - S_S}{\Delta\epsilon \cdot l}. \quad (19b)$$

As seen in Section 4 of the supplemental text, values of $[CaB]_S$ were always greater, both with and without muscle present, than the expected amounts, a problem attributable to Ca leaching from the labware used for the standard measurements. For the purposes of comparing whether muscle affects the response to Ca, the Ca-leaching problems can be ignored because the errors associated with the Ca leaching should have been the same with and without muscle present. For this purpose, it is helpful to define a parameter, denoted “ ρ ,” that gives the ratio of the measured value of $[CaB]_S$ with muscle present to that measured with no muscle present, obtained by taking the ratio of Eqs. 19a and 19b and given by the relationship

$$\rho = \frac{A_S - A_M}{S_S - S_M} = \frac{A_M - A_S}{S_M - S_S}. \quad (20)$$

A value of 1 for ρ would indicate that muscle does not affect the responsiveness of BAPTA to Ca. It is noted that the values of $S_M - S_S$ should not be corrected for the small difference in BAPTA concentration in the solution compared with the muscle sample, as done with Fig. 5, as the change in absorbance upon the addition of the standard does not depend on $[B_T]$ (see Eq. 7). For the example in Figs. 4 and 5, the values of $A_M - A_S$ and $S_M - S_S$ were, respectively, 0.1371 and 0.1458 (from the points labeled with the bottom arrow in Fig. 4 B and the arrow in Fig. 4 D, respectively), giving a value for ρ of 0.940.

All of the data in this Methods and Approaches article were obtained in 11 sets of 8–20 samples with muscle present and an associated single sample with solution only; all samples in each set were processed and measured on the same day. The average value of ρ , including experiments with CSQ knocked out and their controls, was 0.967 (SEM = 0.004; $n = 164$). Although significantly <1 , the difference is small (only 3.3%), indicating that possible binding of BAPTA to muscle components is small or, if a significant amount of such binding does occur, it has only a minor effect on the ability of BAPTA to detect Ca.

Values of $[Ca_T]_{WM}$ in mice increase with decreasing muscle weight

All four panels in Fig. 6 plot the concentration of total Ca in whole muscles ($[Ca_T]_{WM}$; determined with Eq. 17)

versus muscle weight. All of the results were with control muscles from 3–6-mo-old mice. A and B show results with EDL muscle samples, and C and D show results with soleus muscle samples. All of the mice in A and C and most of the mice in B and D were C57BL/6 mice, whereas some of the mice in B and D were from hybrid mixes, as indicated in the figure legend. The results in A and C are from the largest set of control mice done with a single set of mice obtained in the same shipment and studied together over a 2-d period. In each of the panels, the line is the least-squares best fit to the $[Ca_T]_{WM}$ versus muscle weight results in that panel. The slopes of all of these lines were significantly different from zero with the exception of that in C, which was almost significantly different ($P = 0.0539$; this and the other p-values are given in the figure legend). The results in Fig. 6 indicate that $[Ca_T]_{WM}$ increases with decreasing muscle weight in both EDL and soleus muscles.

These inverse relationships suggest the existence of a physiological mechanism that up-regulates the cellular Ca load to somehow compensate for the lower muscle weights. In particular, if the lower muscle weights were required to move a heavier load (a greater weight of mouse) with respect to the size of the muscle, this could activate a signaling pathway that stimulates Ca entry into the cell, thereby increasing Ca storage in the SR to somehow increase the specific force generated by the muscle. We assessed the first part of this hypothesis, namely whether the smaller muscles were associated with a relatively greater load, using a parameter (denoted “ R ”) given by the ratio of the weight of the mouse from which a muscle was taken to the weight of the muscle. As shown in Fig. S5 (E and F), R versus muscle weight values are well described by a linear, inverse relationship, i.e., confirming that R does, in fact, increase with decreasing muscle weight.

To show how $[Ca_T]_{WM}$ varies with R , the horizontal scales at the tops of the panels in Fig. 6 show R values corresponding to the muscle weights shown on the abscissa, generated using the equations for the least-squares best-fit lines of the R versus muscle weight data in E (EDL) or F (soleus) in Fig. S5. (As indicated in Section 5b of the supplemental text, the linear relations used for the R scales are expected to apply to the results in Fig. 6, even though mouse weights, and therefore R values, were not determined for most of the data in Fig. 6.) As seen in all of the panels in Fig. 6, $[Ca_T]_{WM}$ increases with increasing R values, results consistent with the idea that increasing the relative workload of a muscle increases Ca loading of the SR.

Knocking out CSQ reduced muscle weight and $[Ca_T]_{WM}$ in EDL but not soleus muscle

Fig. 7 plots measurements of $[Ca_T]_{WM}$ versus muscle weight from EDL (A) and soleus (B) muscle samples with either just the skeletal-muscle form of CSQ (CSQ1)

knocked out (red circles) or with both the skeletal and cardiac forms of CSQ (CSQ1 and CSQ2) knocked out (B, blue triangles). The associated control results measured at the same time are plotted as open triangles. The control results and the lines in A and B are the same as those shown in Fig. 6 (B and D, respectively). The lines indicate the expected average value of $[Ca_T]_{WM}$ for that particular muscle weight for control muscle samples. For both EDL and soleus muscle samples, there were no significant differences in the average values of $[Ca_T]_{WM}$ for both the single and double CSQ KO.

It is evident from the results for EDL muscle samples in Fig. 7 A that the CSQ-KO muscles tended to weigh less than the control muscle, and that the values of $[Ca_T]_{WM}$ for 11 of the 12 KO muscles were significantly below the line for the control muscles. In the following

assessment, the point indicated by the arrow in Fig. 7 A is not included; it is one of the two outlier points mentioned in Section 6 of the supplemental text that were rejected with the criterion that its value is more than three standard deviations above the mean of the other points in its group. The degree to which $[Ca_T]_{WM}$ is lower than the control muscles was assessed by taking the ratio, for each point, of the measured value of $[Ca_T]_{WM}$ and the expected control value for the muscle weight of the point, given by the value of the line. The average (SEM) values for this ratio for the EDL muscle results in Fig. 7 A are 0.559 (0.047; $n = 6$), 0.686 (0.043; $n = 5$), and 0.616 (0.037; $n = 11$) for, respectively, the CSQ1-KO muscles, double-KO muscles, and the combined data from both groups. All three averages are significantly < 1 . The mean for the CSQ1-KO muscles is not significantly less than that for the double-KO muscles,

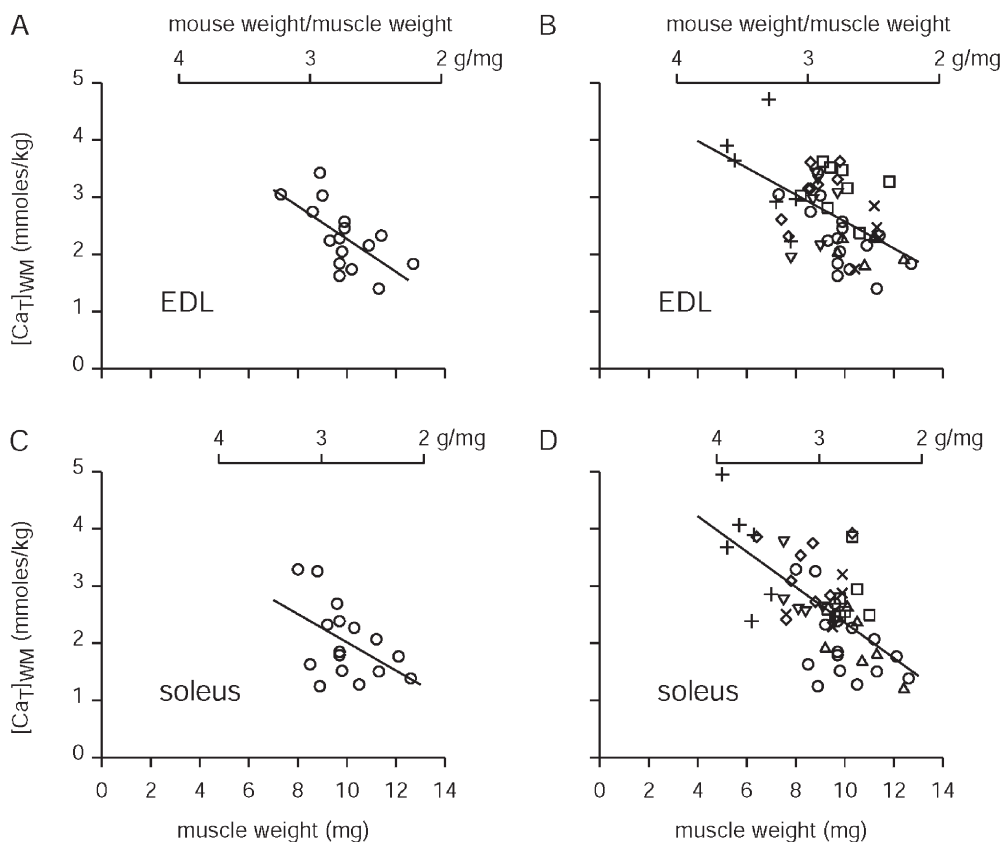


Figure 6. Inverse relationship between $[Ca_T]_{WM}$ and muscle weight in muscles from mice. (A–D) The open circles in A and C plot $[Ca_T]_{WM}$ versus muscle weight for EDL and soleus muscles, respectively, from a set of 4–6-mo-old C57BL/6 mice. B and D plot these same results in A and C, respectively, along with several other sets of muscles. Each set of muscles in a set were processed together on the same day or, in one case, a 2-d period. As for the set plotted with open circles from A and C, the sets plotted with open squares and open diamonds were also obtained with 4–6-mo-old C57BL/6 mice. All of the other four sets were genetically modified to allow for selective deletion of normally expressed genes. Because the genes function normally and because the mice were crossbred on background C57BL/6 or C57BL/6J mice, results for these other four sets would not be expected to differ from those

with the C57BL/6 mice. For the two sets plotted with + and × symbols, the mice ($RANK^{floxed/floxed}$) were genetically modified to allow for deletion of the gene for the receptor activator of nuclear factor κB (RANK), an enzyme involved in regulating bone remodeling. These mice also underwent mock surgical experiments involving exposing the sciatic nerve without cutting it, and then closing the wound 2 wk before the measurements here. Because this would not be expected to alter $[Ca_T]_{WM}$, it seems reasonable to include these sets. For the two sets plotted with open triangles and open-inverted triangles, the mice were modified to allow for deletion of CSQ (results given in Fig. 7). The $RANK^{floxed/floxed}$ mice were 3–4 mo old, somewhat younger than the range of 4 to 6 mo for the mice in the other sets. The lines in each panel were obtained with least-squares best fits to the data. The best-fit slopes in units of millimoles/kilogram per milligram of muscle weight (y intercepts in millimoles/kilogram; p-values) for the lines in A–D are, respectively, -0.2866 (5.13; $P = 0.0072$), -0.2347 (4.92; $P < 0.0001$), -0.2489 (4.48; $P = 0.0539$), and -0.3102 (5.46; $P < 0.0001$). The average values (and n values) of $[Ca_T]_{WM}$ in A–D are, respectively, 2.30 (16), 2.71 (54), 2.02 (16), and 2.62 (54). For EDL muscles in A and B, the horizontal scales on top were calculated with the linear least-squares best-fit line of R versus muscle weight in Fig. S5 E given by $R = -0.187 \times \text{muscle weight} + 4.59$. The corresponding relationship for soleus muscles in C and D is given by $R = -0.239 \times \text{muscle weight} + 5.14$.

although the difference is close to significant ($P = 0.082$). In summary, taking into account the effect of muscle weight on $[Ca_T]_{WM}$ for the combined results, we conclude, in EDL muscle, that knocking out CSQ does decrease $[Ca_T]_{WM}$ by 38% on average compared with WT muscles. This decrease is significantly less than the >80% reduction expected as a result of the loss of Ca bound to CSQ (see Discussion).

$[Ca_T]_{WM}$ values in skeletal muscles from rats

Values for $[Ca_T]_{WM}$ were determined for whole skeletal muscle samples from rats and frogs, processed in essentially the same way as those from mice. For EDL muscle samples from Sprague-Dawley rats, the average value of $[Ca_T]_{WM}$ was 1.46 mmoles/kg (SEM = 0.09 mmoles/kg; $n = 8$). For soleus muscles from the same four rats, the average value of $[Ca_T]_{WM}$ was 1.26 mmoles/kg (SEM = 0.10 mmoles/kg; $n = 8$). Weights of the EDL muscles ranged from 0.135 to 0.296 g and averaged 0.199 g (SEM = 0.023 g). Weights of the soleus muscle ranged from 0.128 to 0.257 g and averaged 0.183 g (SEM = 0.016 g). Although there was no apparent dependence of $[Ca_T]_{WM}$ on muscle weight for the rat muscles, the sample size was too small to firmly establish a lack of effect. The values of $[Ca_T]_{WM}$ in rats given here are about half of those for mice (the average values for EDL and soleus muscles from mice given in the legend of Fig. 6 were 2.71 and 2.62 mmoles/kg, respectively).

$[Ca_T]_{WM}$ values in twitch skeletal muscles from frogs and assessment of whether or not there is a significant extracellular Ca component

Values for $[Ca_T]_{WM}$ were obtained for semitendinosus and ileofibularis skeletal muscles from frog. The semitendinosus muscles are composed of twitch fibers. For ileofibularis, almost all of the fibers are twitch fibers, with only a small fraction of slow fibers found mixed with twitch fibers in the “tonus bundle” (Peachey and

Huxley, 1962), which is a small part of the whole muscle. The average values of $[Ca_T]_{WM}$ were about the same for the two groups, so the results for the two muscles are combined here. Three conditions were evaluated that differed by exposure to external solution of the whole muscles after they were removed. The first condition involved the usual procedure of an ~5-min exposure of the muscle to normal Ringer’s solution, followed by gentle blotting dry and weighing of the muscle. Because of the brief exposure time and the nominally physiological level of Ca in the normal Ringer’s solution, this condition should not have affected $[Ca_T]_{WM}$. For the second and third conditions, the muscles were exposed for periods of 1 h in, respectively, the 0-Ca Ringer’s and normal Ringer’s solution. For conditions 1, 2, and 3, the average (SEM) values in mmoles/kg for $[Ca_T]_{WM}$ were 1.88 (0.11; $n = 16$), 1.95 (0.25; $n = 8$), and 2.26 (0.27; $n = 7$). (An outlier point with a value of 5.25 mmoles/kg was excluded for condition 3. This was one of the two outlier points excluded in this study for the reasons given in Section 6 of the supplemental text.) There was no statistically significant difference between any of the three conditions. The average values for conditions 1 and 2 of 1.88 and 1.95 mmoles/kg were almost identical, indicating that there was not a significant extracellular Ca component in frog muscle. The slightly higher average value with exposure to the 0-Ca Ringer’s is likely a result of random variation. As the two sets of muscles were processed on different days, this could have been caused by random errors affecting one or both of the background levels of Ca for the two sets. The similarity of the results for conditions 2 and 3 (which were from opposite leg muscles from the same frogs) provides additional evidence that there was not a large amount of Ca in the extracellular space. As discussed further in the Discussion, this contrasts with an earlier study with ^{45}Ca , which reported that more than half of the total Ca content of whole muscle from frog is extracellular, a result

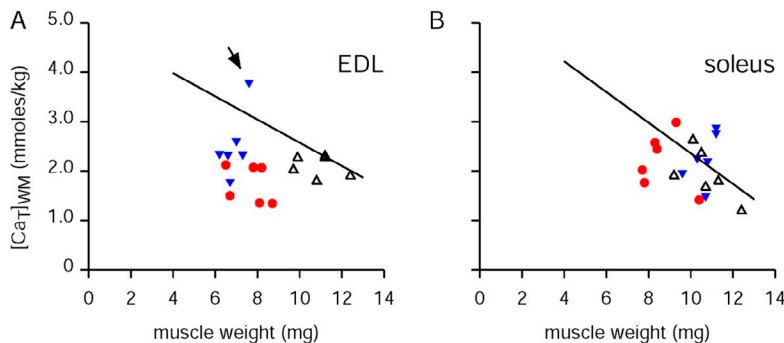


Figure 7. Effect of knocking out CSQ on $[Ca_T]_{WM}$. The solid lines in A and B for EDL and soleus muscles, respectively, are the same best-fit lines obtained for all of the control results shown in Fig. 6 (B and D, respectively). The red circles were obtained with muscles from mice with the skeletal muscle isoform of CSQ (CSQ1) knocked out, and the blue triangles were with both the skeletal and cardiac isoforms (CSQ1 and CSQ2) knocked out. The open triangles were from the control muscles processed at the same time; these were also plotted in Fig. 6. For EDL muscles ($n = 6$ for each case), the average (SEM) values of $[Ca_T]_{WM}$ in millimolars were 2.12 (0.09), 1.73 (0.16), and 2.54 (0.29) for, respectively, the controls, the CSQ1 KOs, and the double KOs. For soleus muscles ($n = 6$ for each case), the corresponding average (SEM) values were 1.95 (0.22), 2.24 (0.25), and 2.31 (0.22).

attributed to the binding of Ca to extracellular components (Kirby et al., 1975).

The presence of a relatively small amount of extracellular Ca indicates that all or most of extracellular Ca was present in the free form, an assumption used in the Discussion for assessing the intracellular component. Although the average value for $[Ca_T]_{WM}$ of 2.26 mmoles/kg with condition 3 (1 h in normal Ringer's solution) was not significantly greater than the average value of 1.88 mmoles/kg with condition 1 ($P = 0.131$), it was close to being significantly greater. (Inclusion of the outlier point above would result in the difference being significant.) Assuming it could be shown with more data points that the 1-h exposure to normal Ringer's solution with condition 3 does tend to increase $[Ca_T]_{WM}$, one possible explanation for this is that the concentration of Ca in the normal Ringer's solution (1.8 mM) is somewhat greater than the physiological extracellular fluid present in the freshly extracted muscle. In support of this possibility, the value of 1.8 mM generally used for frog muscle could be too high, as it is apparently based on limited measurements made almost 80 years ago and before. (Its apparent origin was the calculation of 1.8 mM by Fenn [1936] for a Ringer's solution that contains 0.2% [wt/vol] of $CaCl_2$. The origin of this percentage is unclear, as it is much greater than the 0.012% [wt/vol] reported by Ringer [1883] from his five parts of $CaCl_2$ solution in 100 cm^3 of saline and other components; the $CaCl_2$ solution contained 1 part of $CaCl_2$ in 390 parts water. The 1.8-mM value does, however, agree with Fenn's measured value of 2 mM for the concentration of Ca in plasma from frog.)

Summary of $[Ca_T]$ values from different species

For the purpose of a convenient reference, Table 1 gives average values for $[Ca_T]_{WM}$ determined in this study and the concentration (mM) of total intracellular Ca referred to the myoplasmic water volume for all of the muscle types and species evaluated in this study. The latter values are provided because they should be close to the concentration of total releasable Ca from the SR referred to the myoplasmic volume usually determined with a myoplasmic indicator. Considering total Ca in just the extracellular (EC) and intracellular (IC) compartments, the following mass balance for Ca should apply:

$$W_M [Ca_T]_{WM} = W_M \cdot 0.124 \cdot [Ca_T]_{EC} + 0.58 \cdot W_M [Ca_T]_{IC_ref_myo}, \quad (21)$$

where W_M is the weight of the muscle (in kg) and $[Ca_T]_{EC}$ is an assumed concentration (mM) of total Ca in the extracellular space. The factor 0.124 gives liters of extracellular water volume per kilogram of whole skeletal muscle and was determined for rat skeletal muscle (Cieslar et al., 1998). The factor 0.58 has units of liters

of myoplasmic water per kilogram of whole muscle (Baylor et al., 1983). Both of these factors are assumed to apply for all of the muscle types given in Table 1. Eq. 21 can be rearranged to give

$$[Ca_T]_{IC_ref_myo} = 1.72 \cdot [Ca_T]_{WM} - 0.214 \cdot [Ca_T]_{EC}. \quad (22)$$

Values for $[Ca_T]_{EC}$ of 2 and 1.8 mM, corresponding to the concentrations of Ca generally used, respectively, in mammalian and frog Ringer's solutions, were assumed for mammalian and frog muscles in Table 1. It is noted that the values for $[Ca_T]_{IC_ref_myo}$ in Table 1 would overestimate total releasable Ca in the SR by the amount of nonreleasable Ca in the SR (if such a component exists) and by resting Ca outside the SR. Estimates of this latter component are given in the Discussion.

Evaluation of the possibility that Ca was "lost" to parvalbumin by adding Mg to the measurement solution

The myoplasmic protein parvalbumin is involved in rapid relaxation in twitch muscle from frogs and fast-twitch muscle from mammals (e.g., Heizmann et al., 1982; Baylor et al., 1983) and is either absent or at nearly undetectable levels in slow-twitch muscles (see, e.g., Heizmann et al., 1982). Each of two metal-binding sites on parvalbumin (formed by E-F hand motifs) binds Ca with very high affinity and Mg with moderate affinity. As shown in Table 2 and associated text in the first part of the Appendix, if parvalbumin maintained its high affinity for Ca after exposure to SDS (which denatures proteins), BAPTA would not have been able to capture Ca from parvalbumin when using our usual measurement solution, i.e., the measurement solution without Mg. If this was the case, the results with the BAPTA method reported thus far would have underestimated Ca by $\sim 25\%$

TABLE 1
Average (SEM) values of $[Ca_T]$

	1	2	3	4
Muscle type	$[Ca_T]_{WM}$	$[Ca_T]$ referred to myoplasm	n	
	mmoles/kg wet weight	mmoles/liter of myoplasmic volume		
1 Mouse EDL	2.71 (0.09)	4.23 (0.14)	54	
2 Mouse soleus	2.62 (0.11)	4.08 (0.17)	54	
3 Rat EDL	1.46 (0.09)	2.08 (0.13)	8	
4 Rat soleus	1.26 (0.10)	1.74 (0.14)	8	
5 Frog ileofibularis and semitendinosus	1.88 (0.11)	2.85 (0.17)	10	

$[Ca_T]$ values for all of the muscle types used in this study. For column 2, the values for mouse EDL and soleus muscles were from the data plotted in Fig. 6 (B and D, respectively), and the values for rat and frog muscles were, respectively, from the third and second preceding sections giving results for these muscles. The values for frog twitch muscle were obtained with condition 1, i.e. with muscles blotted dry soon after extraction. See text associated with this table for the determination of values in column 3.

because of “loss” of Ca bound to parvalbumin, either in the pellet after centrifugation or with any parvalbumin still in the supernatant (see Tables 2 and 3 in the Appendix). However, results in Cox et al. (1979; their Table 1) showed that treatment of parvalbumin with 1% SDS resulted in loss of parvalbumin’s metal-sensitive fluorescence. This strongly suggests that the denaturing of parvalbumin by SDS prevents metal binding to parvalbumin. Because the concentration of SDS was similar in this study (0.5 vs. 1%), it is likely that metal binding to parvalbumin was also eliminated or at least parvalbumin’s affinity was reduced enough so that BAPTA was able to capture Ca from parvalbumin.

To assess whether or not Ca was lost to parvalbumin, Mg was added to the usual measurement solution to lower parvalbumin’s apparent affinity for Ca (see Eq. A2 in the Appendix). Results are compared below with no added Mg in the measurement solution (the usual solution used for all of the results thus far) and with Mg added so that the modified measurement solution contained 2 mM [Mg] in addition to its usual constituents. Before presenting these results, it is useful to first consider simulations of the experiments with the BAPTA method using published values for the Ca- and Mg-binding constants of BAPTA, EGTA, and parvalbumin, performed with the following three sets of conditions (see Table 3 in the Appendix and associated text): condition 1: with a Mg-free measurement solution and with no binding of Ca or Mg to parvalbumin; condition 2: with a Mg-free measurement solution and now including metal binding to parvalbumin; and condition 3: with 2 mM Mg and also including metal binding to parvalbumin. As seen from the last row of Table 3 for conditions 1–3, the BAPTA method would have reported 99.6, 75.2, and 94.8%, respectively, of the total Ca present (ignoring the relatively small amounts of Ca that might be bound to other sites as seen in Table 2). Based on these simulations, if parvalbumin’s metal-binding properties were not affected by SDS, the reported values of $[Ca_T]_{WM}$ with 2 mM Mg present (condition 3) would be greater on average than those without Mg present (condition 2) by a predicted 26.1% ($100 \times (94.8 - 75.2) \div 75.2$). If parvalbumin’s ability to bind metals was lost because of the SDS treatment, the addition of 2 mM Mg is predicted to have a negligible effect, decreasing the percentage of total Ca reported by BAPTA from 99.6% (condition 1) to 99.4% (results for this condition are not depicted).

For muscles from eight mice, a value of $[Ca_T]_{WM}$ for EDL muscle with 2 mM Mg in the measurement solution was obtained from the muscle from one leg of a mouse (left or right), and the value with no Mg present (the usual measurement solution) was obtained from the muscle taken from the opposite leg. The average value of 3.194 mmoles/kg (SEM = 0.275 mmoles/kg) for $[Ca_T]_{WM}$ with 2 mM Mg present was almost the same

as that without Mg present, 3.124 mmoles/kg (SEM = 0.180 mmoles/kg). These results support the idea that denaturation of parvalbumin by the SDS treatment occurred, thereby allowing BAPTA to capture Ca that might otherwise have been lost as a result of binding to parvalbumin. However, this conclusion is not certain because of unexpected results for the soleus muscles from the same eight mice. Soleus muscle contains little, if any, parvalbumin (Heizmann et al., 1982), so that one would not expect a difference in $[Ca_T]_{WM}$ with Mg added. Contrary to this expectation for the soleus muscles, the average of 2.418 mmoles/kg (SEM = 0.120 mmoles/kg) with 2 mM Mg present was significantly lower ($P = 0.015$; paired *t* test) than the average value of 3.268 mmoles/kg (SEM = 0.204 mmoles/kg) without Mg present in the measurement solution. We have no explanation for this difference other than random error or the possibility of some unexplained effect of Mg. Because of this unresolved problem, we can only tentatively conclude that the BAPTA method was able to record essentially all (within a few percent) of the Ca present in the muscle, a conclusion based on the lack of any effect of the added Mg on the reported $[Ca_T]_{WM}$ values in EDL muscle and the results from Cox et al. (1979) cited above.

DISCUSSION

Much of this Methods and Approaches article was devoted to verifying the reliability of a new method for measuring the total Ca content of a tissue sample based on BAPTA absorbance. The Discussion begins with an explanation of the need to measure background Ca, much of which may come from Ca leaching from labware. Results reviewed afterward indicate that the method does provide a reliable measure of total muscle Ca if the background level of Ca and the additional problem of stray light are properly addressed.

The problem of Ca leaching from labware

Results in Fig. S4 and Section 4 of the supplemental text show that Ca leaches slowly (over many days) from glass scintillation vials (used to store the 1.5-mM $CaCl_2$ standard solution) and very rapidly from the Pasteur pipettes used to mix all of the aliquots. The very rapid leaching from Pasteur pipettes is presumably caused by readily releasable Ca present on the surface of the glass. As explained with Fig. S4, because this Ca leaching only affected the Ca-standard aliquots (A_5 and S_5), this did not impact the results reported in this article, as the Ca-standard measurements were not needed and not used for the determination of total Ca.

It has long been recognized by some researchers that leaching of divalent ions (Ca^{2+} , Mg^{2+} , Pb^{2+}) from some types of glass used for patch-clamp electrodes can affect ionic currents, and that chelating agents can minimize

such effects (Cota and Armstrong, 1988; Rojas and Zuazaga, 1988). We were unable to find any references addressing the problem of Ca leaching from other forms of labware, although anecdotal evidence suggests that at least some researchers are aware of the possibility. Given this background, we are not sure to what extent the problem of Ca leaching revealed here is a new finding. In any case, the results suggest that such Ca leaching could produce unexplained experimental results under some conditions if it is not recognized and addressed. The BAPTA method would be useful for assessing Ca leaching from labware, as done with Fig. S4.

For the purposes of this study, Ca leaching would not be a problem as long as the amount of Ca leached was the same for all of the labware. To highlight this point, it is noted that some initial, confounding results were later attributed to a variable degree of Ca leaching from polycarbonate centrifuge tubes initially used for the larger muscles from frogs and rats. These results are not reported here, and other types of centrifuge tubes were subsequently used that gave constant background Ca levels indicating either no leaching or about the same degree of Ca leaching from each tube. The reason a constant amount of leached Ca would not be a problem is that it would be included in the background Ca level, which is subtracted off. Therefore, in addition to making measurements in parallel without tissue present, application of the BAPTA method (and most other methods for assessing total Ca) should include confirmation of constant background Ca levels with the labware used.

The BAPTA method can reliably estimate the total Ca content of a muscle sample

In regards to the reliability of the BAPTA method, one major concern was the possibility that muscle components could somehow affect the results. One problem would be if remnant muscle components in the supernatant after centrifugation or substances in the solution (such as the pH buffer used or any remaining SDS) had a Ca-dependent absorbance in the UV. This possibility was discounted by results in Fig. 2 that showed no difference in the “intrinsic” absorbance of the four aliquots (A_M , A_0 , A_S , and A_z ; measured without BAPTA present). This finding justified Assumption 4 in Materials and methods (so that A_{intr} was the same in all four aliquots).

Another key assumption, based on the relatively large concentration of BAPTA and BAPTA’s high affinity for Ca, is that BAPTA captures essentially all of the Ca initially present in the muscle (Assumption 1 in Materials and methods). Calculations in Tables 2 and 3 indicate that BAPTA should, in fact, capture essentially all of the Ca on the major Ca-binding proteins in muscle, with the possible exception of Ca bound to the Ca/Mg sites on parvalbumin, troponin, and myosin. Based on experiments in the last section of Results with 2 mM Mg to greatly reduce possible Ca binding to these sites, we

tentatively concluded that Ca was not lost to these sites. As mentioned, the most likely explanation for this is that SDS denatured parvalbumin and perhaps also troponin and myosin, thereby lowering their abilities to bind Ca, allowing BAPTA to capture all or most of the Ca that might otherwise have remained bound to the Ca/Mg sites. Even though the lack of added Mg in our basic measurement solution did not appear to have affected the results in this article, a Mg concentration of 2 mM or more in the measurement solution is recommended for future applications of the BAPTA method.

Results showed that $[B_T]$ with muscle present was, on average, $\sim 96\%$ of the value with no muscle present, indicating that very little BAPTA was lost in the pellet formed in the centrifugation step and/or by some interaction with soluble muscle components still present in the supernatant. More importantly, based on comparisons of BAPTA’s response to the addition of a Ca standard in the presence and absence of muscle, the ability of BAPTA to detect Ca was not greatly affected by the presence of muscle, with the results indicating a possible underestimate of $[Ca_T]_{\text{WM}}$ of only 3.3% on average. This result largely justifies Assumption 5 in Materials and methods that binding of Ca-free or Ca-bound BAPTA to muscle components does not occur or, if it does, that it has little effect on BAPTA’s response to total Ca.

This section covered three of the five assumptions for the BAPTA method given in Materials and methods. As indicated in Materials and methods, the other two assumptions should also be valid. We conclude that the five key assumptions are all valid and that the BAPTA method should provide a reliable measure of the total Ca content of a tissue sample.

Different components of total Ca in whole muscle

For discussions below, it is helpful to consider the following components of the concentration of total Ca in whole muscle: extracellular ($[Ca_T]_{\text{EC}}$), intracellular outside of the SR ($[Ca_T]_{\text{NonSR}}$), and intracellular inside the SR ($[Ca_T]_{\text{SR}}$). With these definitions,

$$[Ca_T]_{\text{WM}} = [Ca_T]_{\text{EC}} + [Ca_T]_{\text{NonSR}} + [Ca_T]_{\text{SR}}. \quad (23a)$$

It is useful to consider the different Ca components in fractional terms, obtained by dividing both sides of Eq. 23a by $[Ca_T]_{\text{WM}}$, giving

$$1.0 = f_{\text{EC}} + f_{\text{NonSR}} + f_{\text{SR}}. \quad (23b)$$

$$\text{EDL muscle: } 1.0 = 0.11 + 0.12 + 0.77 \quad (23c)$$

$$\text{soleus muscle: } 1.0 = 0.11 + 0.02 + 0.87 \quad (23d)$$

Eqs. 23c and 23d give estimates for the fractions of $[Ca_T]_{WM}$ associated with the different components in EDL and soleus muscles, respectively. The value of 0.11 for f_{EC} is from the value of 0.124 liters of extracellular water volume per kilogram of whole skeletal muscle determined for rat skeletal muscle (Cieslar et al., 1998), the concentration of 2 mM for Ca in the extracellular space (set by the concentration in the mammalian Ringer's solution), and the average value of $[Ca_T]_{WM}$ for the EDL muscles from C57BL/6 mice in Fig. 6 A of 2.30 mmoles/kg ($0.11 = 0.124 \times 2 \text{ mM} \div 2.30$). The value of 0.12 for f_{NonSR} in Eq. 23c is from the sum of values of 0.224 and 0.046 mmoles/kg of whole muscle for the estimated concentrations of Ca bound to the Ca/Mg sites on parvalbumin and troponin in the resting state (see "Estimation of Ca bound in the resting state..." in the Appendix; $0.12 = (0.224 + 0.046) \div 2.30$). Because soleus muscle has little, if any, parvalbumin (Heizmann et al., 1982), the value of 0.02 for f_{NonSR} in Eq. 23d is that estimated for troponin only ($0.02 = 0.046 \div 2.30$). As indicated in the second section of the Appendix, parvalbumin and the Ca/Mg sites on troponin are the only myoplasmic sites with a nonnegligible resting Ca level compared with the total Ca content of muscle. As also indicated, the resting Ca content of mitochondria should also be negligible. It is expected that other non-SR components of Ca are also negligible. The values of 0.77 and 0.87 for f_{SR} in Eqs. 23c and 23d, respectively, were calculated to balance the equations. We assume that the values in Eq. 23c also apply to twitch muscle from frog discussed in the next section.

The intracellular Ca content with the BAPTA method agrees with the SR Ca content in frog cut twitch fibers
As mentioned in the Introduction, one reason for obtaining information about the Ca content of whole tissue is to assess whether or not the Ca contents in isolated single-cell preparations under nominally physiological conditions are, in fact, physiological. As shown in this section, the values of Ca content in whole muscle reported in this study for frog muscle agree with the total amount of releasable Ca from the SR in cut twitch fibers from frog measured with the EGTA/phenol red method (same as $[Ca_T]_{SR}$ in Eq. 23a above; concentrations referred to myoplasmic water volume). Similar to the BAPTA method introduced in this study, with the EGTA/phenol red method, a relatively large concentration of a Ca buffer (EGTA in this case) is present, capturing almost all (>97%) of the released Ca. Cut fibers were equilibrated for a period of ~60 min with an internal solution (i.e., end-pool solution) that contained 20 mM EGTA and 1.76 mM of total Ca at pH 7, giving an estimated concentration of 36 nM for free Ca^{2+} in the end pools. Based on the active pH signal in response to a "fully depleting" train of action potentials, Pape et al. (1995; their Table 2) obtained an average value for $[Ca_T]_{SR}$

of 2.685 mM (SEM = 0.252 mM; $n = 12$; concentration referred to myoplasmic water volume) under current-clamp conditions (normal Ringer's external solution and K internal solution). The average value under voltage-clamp conditions of 2.544 mM (SEM = 0.070 mM; $n = 11$) was very similar to that obtained under current-clamp conditions. Two estimates for $[Ca_T]_{SR}$ from the results in this study are given next.

The first estimate of $[Ca_T]_{SR}$ is based on the average value of 1.96 mmoles/kg (SEM = 0.25 mmoles/kg; $n = 8$) for $[Ca_T]_{WM}$ for muscles exposed to the 0-Ca Ringer's solution for 1 h (condition 2 in "[Ca_T]_{WM} values in twitch skeletal muscles from frogs..." in Results) and assumes that all of the extracellular Ca diffused out of the muscle during the 1-h period and that none of the intracellular Ca was lost. From Eq. 23b above with $[Ca_T]_{EC} = 0$, the estimated fraction of $[Ca_T]_{WM}$ coming from the SR is 0.87 ($0.87 = 0.77 \div (0.012 + 0.77)$). With this fraction along with the value for $[Ca_T]_{WM}$ and the factor of 0.58 kg (or liter) of myoplasmic water per kilogram of whole muscle (Baylor et al., 1983), the first estimate of $[Ca_T]_{SR}$ is 2.94 mM (SEM = 0.38 mM; $n = 8$; the values 2.94 and 0.38 mM are given by 1.96 and 0.25 mmoles/kg, respectively, multiplied by 0.87 and divided by 0.58). This average value is not significantly different from the average value of 2.685 mM for $[Ca_T]_{SR}$ in cut fibers above ($P = 0.47$).

The second estimate of $[Ca_T]_{SR}$ is obtained from the average value of 1.88 mmoles/kg (SEM = 0.11 mmoles/kg; $n = 16$) for $[Ca_T]_{WM}$ for muscles quickly rinsed in normal Ringer's solution and blotted dry just after extraction, thereby minimizing any diffusional loss or gain from the extracellular space (condition 1 in "[Ca_T]_{WM} values in twitch skeletal muscles from frogs..." in Results). This second estimate is 2.50 mM ($2.50 = 1.88 \times 0.77 \div 0.58$), which is very close to the average value of 2.685 mM for $[Ca_T]_{SR}$ in cut fibers. Because the second estimate did not require an assumption concerning possible loss of Ca from the extracellular compartment, it should give the best estimate of $[Ca_T]_{SR}$ under physiological conditions. In summary, the nominally physiological values of $[Ca_T]_{SR}$ for cut fibers from frog twitch muscle in Pape et al. (1995) are, in fact, close to the physiological level estimated here.

Comparison with the SR Ca contents determined in mammalian skinned fibers

From Results, the average values of $[Ca_T]_{WM}$ from EDL and soleus muscles from rat (SEM; all units in mmoles/kg of muscle; $n = 8$ for each case) were 1.46 (0.09) and 1.26 (0.10), respectively. For comparison with results below, these values were multiplied by the assumed density of muscle (1.06 kg per liter) to give units in millimolar that referred to whole-muscle volume and then multiplied by the fractions of 0.77 and 0.87 for EDL and soleus muscles, respectively, from Eqs. 23b and 23c,

respectively. The resulting estimates of $[Ca_T]_{SR}$ that referred to whole-muscle volume are 1.12 (0.07) and 1.10 (0.09) mM (units of mmoles of Ca/L of muscle). These values are compared here with $[Ca_T]_{SR}$ values estimated in skinned fibers from EDL and soleus muscles from rat (Fryer and Stephenson, 1996). The somewhat involved approach consisted of mechanically skinning the fibers under oil, followed by a brief (~1-min) “equilibrating period” in a solution containing BAPTA (in part to prevent saturation of the Ca-regulatory sites on troponin) and Ca with a Ca/BAPTA ratio giving a nominally physiological concentration of free Ca^{2+} of 71 nM (pCa = 7.15). The total Ca content was then determined by the integrated tension response after permeabilization of the SR membrane with triton, calibrated afterward with tension responses with different Ca/BAPTA ratios. The average values of total Ca from the SR that referred to the whole fiber volume for EDL (fast-twitch) and soleus (slow-twitch) fibers in Fryer and Stephenson (1996) were 1.32 mM (SEM = 0.02 mM) and 1.35 mM (SEM = 0.08 mM), respectively (values of n were not given, although they were apparently in the range of 20 to 65). The average value of 1.32 mM for EDL muscle was significantly greater than that of 1.12 mM estimated above from the value of $[Ca_T]_{WM}$ in this study, whereas the value of 1.35 mM for soleus muscle was not significantly greater than that of 1.10 mM from this study (regardless of the value of n assumed for the skinned-fiber results, assumed to be 20 or 65). Despite the significant difference for the EDL muscles, the average values are fairly close. Also, because whole-muscle volume contains interstitial space and whole fiber volume does not, the values of 1.12 and 1.10 mM for $[Ca_T]_{SR}$ from this study should be scaled up somewhat for the comparison with those estimated with skinned fibers. We conclude that the values with the two methods are in agreement. This agreement indicates that the total Ca content obtained in both fast-twitch and slow-twitch skinned fibers from rat by Fryer and Stephenson (1996) were very close to the physiological level, in support of the requirement of a minimal loss or gain of Ca during the equilibrating period of their protocol described above.

Using essentially the same approach for estimating $[Ca_T]_{SR}$ in skinned fibers from human skeletal muscle, Lambole et al. (2013b) obtained average values of 0.85 and 0.76 mM (these referred to whole-fiber volume) for fast-twitch (type II) and slow-type (type I) fibers, respectively, both obtained from vastus lateralis muscle biopsies. These values are similar, although somewhat less than the average values above for rat muscle.

Agreement with earlier methods for measuring Ca content of mammalian whole muscle

The most common method previously used for determining total Ca content of whole muscle involved some type of treatment of the muscle sample, followed by

measurement of total Ca with atomic absorption spectroscopy (AAS). The most common, and also most recent, pretreatment protocol was somewhat similar to that used in this study, involving homogenization of the sample and then exposure to trichloroacetic acid (TCA) to precipitate proteins, followed by centrifugation to remove muscle components. With this approach, Gissel and Clausen (1999) reported an average value of 1.21 mmoles/kg (SEM = 0.04 mmoles/kg; $n = 10$) for $[Ca_T]_{WM}$ in rat soleus muscle. This average value is not significantly different from the corresponding average value obtained for rat soleus muscle in this study of 1.26 mmoles/kg (SEM = 0.01 mmoles/kg; $n = 8$). With the same approach with TCA and AAS, Everts et al. (1993) reported an average value of 1.46 mmole/kg (SEM = 0.04; $n = 3$) for rat EDL muscle, which is the same value obtained in this study, 1.46 mmoles/kg (SEM = 0.09 mmoles/kg; $n = 8$). It is noted that the problem of possible loss of Ca to parvalbumin addressed in the last section of Results would also apply to the pretreatment protocol with TCA. The agreement of the total Ca contents in rat EDL muscle with the TCA-AAS approach with the results from this study suggests that the denaturing effect of TCA also reduced parvalbumin’s apparent affinity for Ca.

There are only a few reports of total Ca content in whole muscles from mice. Our best estimates for this data, given in bar graphs for tibialis anterior in Yoshida et al. (2006) and gluteus and quadriceps muscle in Glesby et al. (1988), are 1.68, 1.68, and 1.77 mmoles/kg. These values are within the range but on the low side of values with EDL and soleus muscles in this study; the average values for $[Ca_T]_{WM}$ for EDL and soleus muscles given in Table 1 are 2.71 and 2.62 mmoles/kg, respectively.

Comparison of Ca content in frog whole-muscle samples with results from earlier methods

Kirby et al. (1975) estimated the concentration of total Ca in whole toe muscles from frog (also *Rana pipiens*) using a ^{45}Ca washout method and AAS. Their values for $[Ca_T]_{WM}$ with the two methods were, respectively, 1.96 and 2.00 mmoles/kg, values not significantly different from the average value in this study of 1.88 mmoles/kg for $[Ca_T]_{WM}$ for twitch muscle (the ileofibularis and semitendinosus muscles, which are leg muscles). In contrast to the results in this study, Kirby et al. (1975; see their Table 2) reported that only 0.83 of the 1.96 mmoles/kg Ca obtained with the ^{45}Ca method was intracellular, with the remaining 1.13 mmoles/kg coming from the extracellular compartment, i.e., only 42% was intracellular ($42\% = 100 \times 0.83 \text{ mM} \div 1.96 \text{ mM}$). They concluded that there must be a very large amount of bound Ca in the extracellular space, perhaps to proteins such as collagen and/or to the surface and T-system membranes. We have no plausible explanation for the fact that our results indicate a much larger amount of intracellular Ca (~2.3-fold) and no bound pool in the extracellular

space. It seems unlikely that the difference is caused by the different muscles used, leg versus toe muscles.

When CSQ is knocked out, muscles compensate for the loss of Ca content associated with CSQ by increasing Ca at other sites

To help interpret the results with CSQ knocked out, Section 7 of the supplemental text gives predicted values of $[Ca_T]_{WM}$ with CSQ KO assuming that the only effect of the KO was to remove the Ca component bound to CSQ. With this assumption, CSQ KO is predicted to reduce $[Ca_T]_{WM}$ to 0.256 and 0.160, respectively, of the values for EDL and soleus muscles from WT mice. Because $[Ca_T]_{WM}$ with EDL muscles from mice with CSQ knocked out in EDL muscles was 62% on average of the value expected for control muscle (combined single- and double-KO results taking into account muscle weight; the 62% value is given in text associated with Fig. 7), the average value of $[Ca_T]_{WM}$ with CSQ KO muscles was 2.4-fold greater than predicted ($2.4 = 0.62 \div 0.256$). For soleus muscles, CSQ KO did not affect $[Ca_T]_{WM}$, so the values of $[Ca_T]_{WM}$ with the CSQ KO muscles were 6.3-fold greater than predicted ($6.3 = 1.0 \div 0.160$). Possible reasons are given below for why the measured value of $[Ca_T]_{WM}$ was so much greater than the predicted values.

As noted in the Introduction, apparently contradictory results have been reported as to whether knocking out CSQ results in a large reduction in total releasable Ca in the SR (which should be equivalent to $[Ca_T]_{SR}$ above). With the assumption that there was no change in the extracellular and intracellular non-SR Ca components, the modest decrease in $[Ca_T]_{WM}$ for CSQ1-KO EDL muscles relative to controls in this study is in agreement with the modest (25%) decrease in readily releasable total Ca in patch-clamped single EDL fibers from CSQ1-KO versus WT mice reported in Sztretye et al. (2011; their Table 1). This modest decrease contrasts with the $\sim 70\%$ reduction in readily releasable Ca estimated in fibers from CSQ1-KO versus WT EDL muscle from mice (Paolini et al., 2007; their Fig. 8, B and E). One way to distinguish between the possibilities would be if one or the other method gave a more dependable estimate of $[Ca_T]_{SR}$. However, although both methods are indirect and can be challenged, they both appear, at first glance, capable of determining the degree to which $[Ca_T]_{SR}$ in KO muscle fibers might vary from that in WT fibers. For the following three reasons, we believe the most likely possibility is that the loss of Ca associated with the removal of CSQ was compensated for by an increase in SR Ca, probably Ca binding to some other protein in the SR. (1) It appears unlikely that the compensatory increase would have occurred outside the SR. A sufficiently large increase in Ca in the extracellular space seems unlikely, as $[Ca^{2+}]$ in the extracellular space was set by the experimental conditions. Also, because both approaches involved isolated single fibers, a difference in $[Ca_T]_{EC}$ would have required

the expression of a previously unknown high capacity Ca-binding protein in the T-system when CSQ is knocked out (a possibility, although a seemingly remote one). In regards to the intracellular, non-SR possibility, parvalbumin is the only known candidate with both the required high capacity and high affinity Ca-binding sites (see second section of Appendix). One argument against an increase in parvalbumin concentration is that it would not explain the lack of a decrease in $[Ca_T]_{WM}$ in soleus muscle because parvalbumin is normally not present in soleus muscle from mice. Also, in EDL muscle, an increase in parvalbumin concentration would have tended to speed up relaxation (the primary function of parvalbumin), whereas relaxation was slower in fibers with CSQ knocked out compared with WT fibers (Paolini et al., 2007), although the somewhat slower relaxation could be caused by other factors. (2) At least in regards to the possibility of a large increase in Ca binding to a myoplasmic site, there would be no apparent physiological advantage, whereas restoring the CSQ's function of storing Ca in the SR would be advantageous. (3) And lastly, at least four Ca-binding protein components in the SR have already been identified that may serve to replace some of the Ca-storing function lost with CSQ KO (the four protein components including sarcoplasmic reticulum are reviewed on p. 415 of Fénelon et al., 2012; to our knowledge, the other two are still unnamed).

Although arguments other than the difference in the two methods above favor the possibility of a compensatory increase in $[Ca_T]_{SR}$ as opposed to an increase in non-SR Ca, it seems worthwhile to discuss the methods used to understand why they may have given different results. Sztretye et al. (2011) estimated SR Ca content from the voltage-activated myoplasmic free Ca^{2+} transient with total Ca released derived from this transient and assumptions about the Ca-buffering properties of myoplasm including added Ca buffer. Importantly, much of the released Ca was captured by the relatively large concentrations of added Ca buffer in the myoplasm, either EGTA or BAPTA, reducing some of the uncertainty in the method and also preventing early termination of SR Ca^{2+} release caused by Ca inactivation so that a large fraction of the resting SR Ca content could be released. The approach of Paolini et al. (2007) was similar to the skinned-fiber approach described above (Fryer and Stephenson, 1996) in that the total Ca content was assumed to be proportional to the integrated tension signal in response to the maximal release of SR Ca (by caffeine in the case of Paolini et al., 2007). One potential problem with the approach of Paolini et al. (2007), however, was that excess Ca buffer was not added so that the tension response would tend to be saturated and, therefore, nonlinear with respect to total Ca released. Another important difference was that released Ca^{2+} would have been able to diffuse away from the fiber with the conditions used in Paolini et al. (2007), unlike

the approach with skinned fibers, as they were immersed in oil. Because results from several laboratories indicate, or at least suggest, that SR Ca^{2+} release is completed significantly earlier in fibers from muscles with CSQ knocked out compared with those from WT muscles (Paolini et al., 2007, 2011; Canato et al., 2010; Sztretye et al., 2011), the tension response should also be shorter if the released Ca is able to diffuse away from the fiber resulting in reduction in the integrated tension response. Any of these differences with the experimental approach of Paolini et al. (2007) may have resulted in a greater reduction in the apparent SR Ca content than what actually occurred with CSQ KO.

Given past results indicating that most of the Ca in muscle is bound to CSQ in WT muscle (see above), and because knocking out CSQ must have eliminated this large component, there must have been a large compensatory increase in Ca at some other site or sites.

As argued above, the SR is the most likely location for this compensatory increase. Paolini et al. (2007) and Protasi et al. (2009) reported several ultrastructural changes in EDL muscles with CSQ knocked out, including the presence of multiple T-tubular elements stacked between somewhat flattened junctional SR regions. Although an increase in SR volume was not observed, the increase in SR membrane area might result in increased Ca-binding sites to SR membrane or any associated protein. In addition, given the changes in the ultrastructure observed, it seems reasonable to suppose that other adaptations may have occurred to maintain normal SR Ca^{2+} release, including the up-regulation of one or more of the four previously identified Ca-binding proteins in the SR (see above) or some, as yet unidentified, Ca-binding protein. Because the concentration of free Ca^{2+} in the SR was apparently not significantly altered by CSQ KO (Canato et al., 2010; Sztretye et al., 2011), a compensatory increase in Ca bound to an alternate Ca-binding protein in the SR would almost certainly have had to involve up-regulation of the concentration and/or the Ca^{2+} -binding affinity of this alternate protein.

The possibility of a physiological mechanism that increases muscle Ca content with increased specific force requirements

As seen in Fig. 6, $[\text{Ca}_T]_{\text{WM}}$ increases substantially as muscle weight decreases in both fast-twitch and slow-twitch skeletal muscle from mice. As seen from the best-fit lines to the data, the Ca contents nearly doubled in both cases with a twofold decrease in muscle weight from 12 to 6 mg, close to the full range of whole-muscle weights for both types of muscle. The least-squares best-fit line in Fig. 6 B for EDL muscle gave values for $[\text{Ca}_T]_{\text{WM}}$ at 12 and 6 mg of 2.104 and 3.513 mmoles/kg, respectively, a 1.67-fold increase. For the same muscle weights for soleus muscle, the values of $[\text{Ca}_T]_{\text{WM}}$ were 1.739 and

3.600 mmoles/kg, respectively, a 2.07-fold increase. The $[\text{Ca}_T]_{\text{WM}}$ in Fig. 6 were also plotted as a function of R (the ratio of mouse weight to muscle weight), a parameter likely reflecting the relative workload or specific force associated with the muscle's contribution to mouse movements. For the decrease in muscle weights above of 12 to 6 mg, the value of R was expected to increase from 2.35 to 3.47 g/mg for EDL muscles and 2.27 to 3.71 in soleus muscle (values determined from linear relationships for R vs. muscle weight given in legend of Fig. 6). For EDL muscles, this means that the 1.67-fold increase in $[\text{Ca}_T]_{\text{WM}}$ was associated with a 1.48-fold estimated increase in R with the decrease in muscle weight from 12 to 6 mg. For soleus muscle, the 2.07-fold increase in $[\text{Ca}_T]_{\text{WM}}$ was associated with a 1.63-fold estimated increase in R . To our knowledge, the inverse relationship between $[\text{Ca}_T]_{\text{WM}}$ and muscle weight is a new finding.

Given the increase in $[\text{Ca}_T]_{\text{WM}}$ with increasing R values, it seems reasonable to hypothesize the existence of a physiological mechanism that somehow increases the Ca load in smaller muscles to maintain the specific force or some other contractile property required by the muscle. Moreover, because an individual muscle fiber would not know the size of the whole muscle, it seems likely that such a mechanism or mechanisms would be controlled by the force demands on individual fibers in the whole muscle.

There could be several explanations for the increase in $[\text{Ca}_T]_{\text{WM}}$ with decreasing muscle weight and increasing R . Although the results thus far do not rule out increases in Ca in non-SR locations, such as the extracellular space and/or mitochondria, the location of the increased Ca is most likely the SR. Such an increase would have to be caused by an increase in the volume fraction of SR in the muscle and/or an increase in the resting concentration of total Ca in the SR ($[\text{Ca}_T]_{\text{SR,R}}$). An increase in $[\text{Ca}_T]_{\text{SR,R}}$ could be caused by an increase in Ca bound to CSQ (and perhaps other Ca-binding proteins in the SR) either by increasing the concentration of CSQ and/or increasing the fraction of binding sites occupied by Ca by increasing the resting concentration of free Ca in the SR ($[\text{Ca}^{2+}]_{\text{SR,R}}$). (Because free Ca^{2+} is only a small fraction of total Ca in the SR, it is unlikely that the large increase in total SR Ca with decreasing muscle weight would be attributable to any significant extent to an increase in the pool of free Ca^{2+} .) The latter case of increasing Ca bound to CSQ by increasing $[\text{Ca}^{2+}]_{\text{SR,R}}$ probably does not occur with soleus muscle, as Fryer and Stephenson (1996) reported that bound Ca in the SR of soleus muscle is at a saturated level. (This was not the case for the fast-twitch muscles that had $[\text{Ca}_T]_{\text{SR,R}}$ values of about one third the saturating level.) Therefore, the large increase in $[\text{Ca}_T]_{\text{WM}}$ with decreasing muscle weight for soleus muscle in this study is presumably caused by increased fractional SR volume and/or an increased

concentration of CSQ and/or some other Ca-binding protein in the SR.

Could an increase in $[Ca_T]_{SR,R}$ result in an increase in the specific force generated by a muscle?

Because measurements combined with modeling results indicate that almost all of the Ca-regulatory sites on troponin are occupied by Ca in response to a single action potential in both fast-twitch and slow-twitch mammalian fibers (Baylor and Hollingworth, 2007; Hollingworth et al., 2012), one question is whether or not an increase in $[Ca_T]_{SR,R}$ could actually result in an increase in specific force generated by a muscle. One argument against the idea that the tension response to a single action potential (twitch response) is already maximal is that the peak of the twitch response is generally significantly less than the tetanic response (e.g., Caputo and Bolaños, 1994). The explanation for this is that maximum tension is only reached when elastic elements in the muscle (mainly the contractile proteins) are adequately stretched, which would usually take more than a single action potential (e.g., Celichowski and Grottel, 1993). In addition, an increase in the twitch response was actually observed after at least one type of increase in $[Ca_T]_{SR,R}$, namely an increase in $[Ca^{2+}]_{SR,R}$ with no change in the concentration of CSQ. Posterino and Lamb (2003) found that Ca overloading of skinned fibers from fast-twitch (rat) or twitch (toad) skeletal muscles resulted in a faster rise time and greater amplitude (by 50%; from their Table 1) of the twitch response. These effects and a much slower rate of relaxation were not caused by differences in the amount of Ca^{2+} released, which remained approximately constant, but rather by a greatly reduced rate of reuptake of Ca^{2+} into the SR by SERCA, presumably caused by a higher value of $[Ca^{2+}]_{SR,R}$. As far as we are aware, there are no reports of the effect on tension of an increase in the concentration of CSQ or any other Ca-binding protein without changes in $[Ca^{2+}]_{SR,R}$, so it is unclear whether such increases would also enhance the twitch response. Nevertheless, the results with Ca-overloading results indicate that the twitch can be significantly increased and prolonged with at least one type of increase in $[Ca_T]_{SR,R}$. As a result, it is possible, as hypothesized above, that a physiological mechanism exists that increases $[Ca_T]_{SR,R}$ to generate a greater twitch response.

Adapting the BAPTA method to other tissues

As noted in the Introduction, the BAPTA method should be useful for determining the concentration of total Ca in samples from most tissues. To help with the introduction of the method in other laboratories and possibly with different tissue types, a condensed version of the protocol and practical suggestions are provided in “A model protocol with practical suggestions...” in Materials and methods.

APPENDIX

BAPTA should capture most of the Ca on intracellular Ca-binding proteins with the possible exception of Ca/Mg-binding sites on parvalbumin, troponin, and myosin

This section assesses BAPTA's ability to capture Ca from the main Ca-binding proteins in muscles. Toward this end, estimates are given for the amount of Ca expected to be bound to various Ca-binding proteins from the muscle compared with that bound to BAPTA, assuming that the Ca-binding properties of these proteins are not affected by the exposure to SDS, which denatures proteins. The Ca-binding sites considered here are troponin, SERCA, CSQ, myosin, and parvalbumin. The total amount of Ca bound to other muscle components is expected to be negligible because of their relatively low capacities compared with the proteins mentioned.

As a first approximation, the concentration of free Ca^{2+} ($[Ca^{2+}]$) is determined assuming a typical amount of Ca bound to BAPTA. This value of $[Ca^{2+}]$ is then used to estimate binding to the main Ca-binding protein components from muscle. As a typical example, we consider the case of a mouse whole muscle weighing 8 mg (8×10^{-6} kg; a typical weight for EDL and soleus muscles) that is homogenized and diluted into a final solution volume of 1.2 ml containing 120 μ M BAPTA resulting in a measured concentration of 2,400 μ moles of total Ca per kilogram of muscle weight. The concentration of total Ca bound to BAPTA in the final solution ($[CaB]$) would be 16 μ M ($16 \mu\text{M} = 2,400 \mu\text{moles/kg} \times 8 \times 10^{-6} \text{ kg} \div 1.2 \text{ ml} \times 10^3 \text{ ml/L}$). Assuming a K_d of 0.22 μ M for Ca binding to BAPTA (from Fig. 1) and a 1:1 binding reaction of Ca^{2+} with BAPTA, the concentration of free Ca^{2+} ($[Ca^{2+}]$) would be 0.034 μ M. These results for BAPTA and free Ca^{2+} are given in the first two rows with data in Table 2. As seen in the column 6 in Table 2, the free form of Ca^{2+} is only 0.0021 of the Ca bound to BAPTA ($0.0021 = 0.034 \mu\text{M} \div 16 \mu\text{M}$), so that neglecting this amount results in an underestimation of total Ca by only $\sim 0.2\%$. Table 2 also gives the amount of Ca bound to the main high capacity Ca-binding proteins in muscle—including the Ca-regulatory sites on troponin, SERCA, and CSQ, and the Ca/Mg-binding sites on parvalbumin, troponin, and myosin—assuming values from the literature for the concentration of total binding sites on the proteins referred to the myoplasm and the apparent K_d for Ca binding. Column 3 in Table 2 gives the concentration of Ca-binding sites after dilution in the final volume (after homogenization and the addition of more solution). Column 5 gives the calculated concentration of Ca bound to the BAPTA and to the muscle proteins, and the column 6 gives the fraction of this amount to that bound to BAPTA. The next four paragraphs, in order, give details for the calculation for Ca binding to: (1) the Ca-regulatory sites on troponin, (2) CSQ, (3) parvalbumin, and (4) the Ca/Mg-binding sites on troponin and myosin.

From Table 1 in Baylor et al. (1983), the concentration of Ca-binding sites on the Ca-regulatory sites on troponin is 240 μM (column 2 of Table 2; concentrations referred to the volume of myoplasmic water in frog skeletal muscle fibers), which corresponds to a concentration of 139 $\mu\text{moles/kg}$ of whole-muscle weight using the factor of 0.58 given in Baylor et al. (1983) for this conversion (they estimated 0.58 kg myoplasmic water per kilogram of whole muscle; $139 = 0.58 \times 240$; the myoplasmic water is also assumed to have a density of 1 g/ml). With the dilution of the homogenized muscle in the measurement solution, the concentration of Ca-binding sites on the Ca-regulatory sites on troponin in the final measurement solution would be 0.93 μM (Table 2, column 3; $0.93 \mu\text{M} = 139 \mu\text{moles/kg} \times 8 \times 10^{-6} \text{ kg} \div 1.2 \text{ ml} \times 10^3 \text{ ml/L}$). With the value of 0.2 μM for the K_d of Ca binding to these sites on troponin in a 1:1 reaction (Baylor et al., 1983), and the value from above of 0.034 μM for $[\text{Ca}^{2+}]$, 0.145 of the Ca-regulatory sites on troponin would be occupied by Ca ($0.145 = 0.034 \mu\text{M} \div (0.034 \mu\text{M} + 0.2 \mu\text{M})$), giving an estimated 0.135 μM of Ca bound to these sites (Table 2, column 5; $0.135 \mu\text{M} = 0.145 \times 0.93 \mu\text{M}$), which is only 0.0084 of the Ca captured by BAPTA (Table 2, column 6; $0.0084 = 0.135 \mu\text{M} \div 16 \mu\text{M}$).

To obtain a rough estimate of possible Ca binding to SERCA, we assumed a value of 240 μM for the

concentration of Ca-binding sites on SERCA referred to myoplasmic water volume (from the estimates from three different methods of 190–490 μM , 220 μM , and 260 μM for frog muscle given in Baylor et al., 1983) and an apparent K_d of 0.43 μM (from the value for the apparent association constant with 1:1 binding of $2.3 \times 10^6 \text{ M}^{-1}$ given in Inesi et al., 1980; note that although Inesi et al. indicated that a cooperative reaction involving the binding of two Ca^{2+} ions per pump molecule better described their data, their K_d given here assuming 1:1 binding provided a rough match to their data and is adequate for the purpose here). As above, with the dilution of the homogenized muscle in the measurement solution, the concentration of Ca-binding sites on SERCA would be $\sim 0.93 \mu\text{M}$ ($240 \mu\text{M} \times 0.58 \times 0.008 \text{ ml} \div 1.2 \text{ ml}$), of which ~ 0.073 would be occupied by Ca ($0.073 = 0.034 \mu\text{M} \div (0.034 \mu\text{M} + 0.43 \mu\text{M})$), giving an estimated 0.068 μM of Ca bound to SERCA ($0.068 \mu\text{M} = 0.073 \times 0.93 \mu\text{M}$) or ~ 0.0043 of the amount of Ca bound to BAPTA ($0.0043 = 0.068 \mu\text{M} \div 16 \mu\text{M}$).

For CSQ, the value in Table 2 for the concentration of Ca-binding sites of 5,500 μM that referred to the myoplasm water volume was obtained from the value of 44,000 μM that referred to SR water volume for Case 1 in Table 1 of Fénelon et al. (2012) and the value of 8 for the ratio of myoplasmic water volume to SR water volume (Section 3

TABLE 2
Ca bound to BAPTA relative to other possible sites

1	2	3	4	5	6
Species	Concentration of Ca binding sites referred to Myoplasm μM	Final solution μM	K_d μM	[CaX] μM	Fraction relative to CaBAPTA
BAPTA		120	0.22	16	1.0
Free Ca^{2+}				0.034	0.0021
Troponin	240	0.93	0.2	0.135	0.0084
SERCA	240	0.93	0.43	0.068	0.0043
CSQ	5,500	21.3	780 ($n = 3$)	1.7×10^{-12}	10^{-13}
Ca/Mg-binding sites with $[\text{Mg}^{2+}] = 0$					
Parvalbumin	1,500	7.0	0.012	5.2	0.33
Troponin	240	0.93	0.002	0.88	0.055
Myosin	240	0.93	0.033	0.46	0.029
Ca/Mg-binding sites with $[\text{Mg}^{2+}] = 2 \text{ mM}$					
Parvalbumin	1,500	7.0	0.276	0.77	0.05
Troponin	240	0.93	0.202	0.134	0.0084
Myosin	240	0.93	18.4	0.002	0.0001

Ability of BAPTA to capture Ca from muscle components. Columns 2 and 3 give the concentrations of the Ca-binding sites associated with the substance in column 1. The concentrations in columns 2 and 3 refer to different volumes, the myoplasmic water volume of the muscle sample for column 2, and the volume of the final solution for the measurement (with the homogenized muscle) for column 3. Details for the conversion from column 2 to column 3 given in the text for troponin also apply to the other muscle components. Column 4 gives the K_d for the substance in column 1. Values in columns 2 and 4 were obtained from the literature as detailed in the text. Column 5 gives the concentration of Ca in the final solution (CaX) bound to BAPTA, present as the free form, or bound to muscle components. As indicated in the text, the assumed value of 16 μM for Ca bound to BAPTA is typical of the experimentally measured values. The concentration of free Ca^{2+} was determined from the relative concentrations of Ca-bound and Ca-free BAPTA and its K_d in column 4. Values for Ca bound to the muscle components were determined with this value for free Ca^{2+} using the values in columns 3 and 4. To assess how well BAPTA is expected to capture Ca from the different sites, column 6 gives the ratio of the values of [CaX] for the different components in column 5 to that for CaBAPTA. With the exception of the last three rows, it was assumed that no Mg was present. The last three rows were calculated with a free Mg^{2+} concentration of 2 mM. See text for additional details.

of the supplemental text for Fénelon et al., 2012; 5,500 $\mu\text{M} = 44,000 \mu\text{M} \div 8$). The value of the K_d and Hill coefficient for Ca binding to CSQ were also taken from Table 1 of Fénelon et al. (2012). The value of 10^{-13} for CSQ in column 6 of Table 2 indicates that BAPTA captures essentially all of the Ca from CSQ.

To summarize so far, the Ca in the free form and that possibly bound to troponin, SERCA, and CSQ amount to only 1.5% of the amount of Ca bound to BAPTA ($1.5\% = 100 \times (0.034 + 0.135 + 0.068 + 1.7 \times 10^{-12} \mu\text{M}) \div 16 \mu\text{M}$).

Three proteins in fast-twitch skeletal muscle have binding sites that can bind either Ca or Mg to the same site (denoted “Ca/Mg-binding sites”) with enough capacity and/or high affinity for Ca to make nonnegligible contributions to total Ca in resting muscles and to compete for Ca with BAPTA. These three proteins are parvalbumin, myosin, and sites on troponin different than the Ca-regulatory sites (Robertson et al., 1981). For each of these proteins, the concentration of Ca bound to the Ca/Mg-binding sites (denoted “CMBS”) is given by the usual relationship for competing ligands, i.e.,

$$[CaCMBS] = [CaCMBS_T] \frac{[Ca^{2+}]}{[Ca^{2+}] + K_{Dapp,Ca}}, \quad (\text{A1})$$

where the subscript “T” is for the total concentration of the sites, and $K_{Dapp,Ca}$ is the apparent K_d for Ca given by the usual relationship,

$$K_{Dapp,Ca} = K_{D,Ca} \left(1 + \frac{[Mg^{2+}]}{K_{D,Mg}} \right), \quad (\text{A2})$$

where $K_{d,Ca}$ and $K_{d,Mg}$ are the K_d s for Ca and Mg binding, respectively, in the absence of the other divalent cation. Likewise for Mg binding,

$$[MgCMBS] = [MgCMBS_T] \frac{[Mg^{2+}]}{[Mg^{2+}] + K_{Dapp,Mg}}, \quad (\text{A3})$$

where

$$K_{Dapp,Mg} = K_{D,Mg} \left(1 + \frac{[Ca^{2+}]}{K_{D,Ca}} \right). \quad (\text{A4})$$

The parameters used for parvalbumin in Table 2 are the same as those given for mouse EDL muscle in Baylor and Hollingworth (2007). It is noted that the value of 1,500 μM for the concentration of Ca/Mg-binding sites on parvalbumin (from Baylor and Hollingworth, 2007) was obtained from a measured value in frog muscle. This value for frog should approximate that for fast-twitch mammalian muscle, as it is close to the estimate of 1,300 μM determined from the value of 4.4 g parvalbumin per kilogram of EDL muscle from mouse measured by Heizmann et al. (1982; the value of 1,300 μM assumed a molecular weight for parvalbumin of 12 kD and the factor of 0.58 above for converting from whole-muscle weight

to myoplasmic water volume, and noting that there are two metal-binding sites per parvalbumin molecule). The assumed values for $K_{d,Ca}$ and $K_{d,Mg}$ for parvalbumin were, respectively, 0.012 and 91.0 μM (Baylor and Hollingworth, 1998).

The concentration of the Ca/Mg-binding sites on troponin is the same as that for the Ca-regulatory sites on troponin (Robertson et al., 1981). As done in Robertson et al. (1981), this same concentration is assumed for the Ca/Mg sites on myosin. The values assumed for the K_d s for Ca and Mg binding to these sites were, respectively, 0.002 and 20 μM for troponin and 0.033 and 3.6 μM for myosin (from Table 1 of Robertson et al., 1981).

As seen in the last two sections in Table 2, Ca binding to the Ca/Mg sites on the three proteins were calculating with no Mg present and with $[Mg^{2+}] = 2 \text{ mM}$. From the results in column 6 of Table 2 with no Mg present (the situation with the usual measurement solution used with the BAPTA method), the estimated fraction of Ca bound to the three proteins relative to that captured by BAPTA was 0.414 ($0.33 + 0.055 + 0.029$), with 80% of this because of parvalbumin ($80\% = 100 \times 0.33 \div 0.414$). The reason for this is the much higher affinities (lower K_d s) of the three proteins compared with that of BAPTA in the absence of Mg (Table 2, column 4). The main reason BAPTA captures most ($\sim 70\%$) of the total Ca is that the total concentration of Ca/Mg-binding sites on these proteins is less than the concentration of total Ca in the muscle.

One way to minimize Ca bound to the Ca/Mg sites on parvalbumin, troponin, and myosin would be to add Mg to the measurement solution to reduce their apparent affinities for Ca. The last section of Table 2 (last three rows) indicates that the addition of 2 mM Mg to the measurement solution greatly reduces the affinities (increases the $K_{Dapp,Ca}$ values given in column 4) for the three proteins so that BAPTA would be able to capture much more of the Ca so that much less Ca would be undetected (or “lost”) by BAPTA. As indicated by the results shown in column 6, almost no Ca would be lost to troponin and myosin, and the predicted loss of Ca to parvalbumin would be reduced from 33 to 5% of that Ca captured by BAPTA. As indicated in the text associated with the experimental results with 2 mM Mg present, there is reason to suspect that the protein-denaturing effect of the SDS treatment of the muscle would have eliminated or greatly reduced parvalbumin’s ability to bind Ca, in which case “loss” of Ca to parvalbumin would not have occurred. To test this, experiments with the BAPTA method were performed with and without 2 mM Mg present. The section after the next section simulates possible outcomes of these experiments depending on whether or not the Ca/Mg sites on parvalbumin (and also troponin and myosin) were able to bind Ca after SDS treatment.

Estimation of Ca bound in the resting state to parvalbumin and other intracellular, non-SR sites

It was of interest to estimate the concentration of Ca bound to parvalbumin and other myoplasmic components in the resting state in muscles from mice. The estimates here assume that resting myoplasmic concentrations of Ca^{2+} and Mg^{2+} in the myoplasm are 0.05 μM and 1 mM, respectively, the values assumed in Baylor and Hollingworth (2007). With this value of $[\text{Mg}^{2+}]$, the value for $K_{\text{Dapp,Ca}}$ for parvalbumin from Eq. A2 is 0.144 μM ($0.144 = 0.012 \times (1 + 1,000/91)$). With this value of $K_{\text{Dapp,Ca}}$ along with the above value of $[\text{Ca}^{2+}]$ and a value for $[\text{Parv}_T]$ of 1,500 μM from Table 2, $[\text{CaParv}]$ from Eq. A1 is estimated to be 387 μM (concentration referred to myoplasmic volume; $387 = 1,500 \times 0.05 / (0.05 + 0.144)$). With the factor of 0.58 kg myoplasm per kilogram of muscle (from Baylor et al., 1983), the concentration of CaParvalbumin referred to whole-muscle weight is 0.224 mmoles/kg of muscle ($0.224 = 387 / 1,000 \times 0.58$). Following the same procedure just given for parvalbumin, the estimated concentrations of Ca on the Ca/Mg sites on troponin and myosin at rest are, respectively, 79 and 1 mmoles/kg of muscle (calculated with their concentration values in Table 2 with the apparent K_{d} s in Eqs. A2 and A4 determined with their Ca and Mg K_{d} s given above). Ca bound to myosin at rest is therefore negligible. The estimated concentration of Ca bound to the Ca/Mg sites on troponin at rest is 0.046 mmoles/kg of muscle ($0.046 = 79 / 1,000 \times 0.58$). The resting values of Ca bound to parvalbumin and troponin are used with Eqs. 23c and 23d in the Discussion and in Section 7 of the supplemental text to provide a predicted value of total Ca in muscle minus the amount of Ca bound to CSQ.

Following the same procedure with values from Table 2 for Ca bound to the Ca-regulatory sites on troponin and SERCA in the resting state gave concentrations for these two components of 0.0089 and 0.025 mM, respectively (volumes referred to myoplasmic volume). The same procedure gave a resting concentration of 0.0002 mM for CaATP (using ATP's concentration and apparent K_{d} for Ca binding with $[\text{Mg}^{2+}] = 1$ mM from Table 1 of Baylor and Hollingworth, 1998). Ca binding to calmodulin should be much less than that to troponin, as the affinity for Ca binding to calmodulin is similar or lower than that to troponin, and calmodulin's concentration is much lower than that of troponin (Robertson et al., 1981; Baylor et al., 1983). Based on these results, Ca bound to parvalbumin and the Ca/Mg sites on troponin are expected to be the only nonnegligible myoplasmic Ca components of total Ca in resting muscle.

This paragraph assesses the possible contribution of Ca in mitochondria to the total Ca content of muscle. Scorzeto et al. (2013) reported values for the concentration of free Ca^{2+} in mitochondria ($[\text{Ca}^{2+}]_{\text{mito}}$) of fast-twitch muscles from mice (*flexor digitoralis brevis*) of 0.16 μM

with WT mice and 0.23 μM with both isoforms of CSQ knocked out. They also gave estimates of the volume fraction of fibers occupied by mitochondria just below the sarcolemma and the rest of the fiber or "intermyofibrillar" region, regions taking up ~ 10 and 90% of the fiber volume, respectively. The volume fraction of mitochondria (with respect to fiber volume) in the intermyofibrillar region averaged 4.3 and 7.2% in WT and the double-KO muscles, respectively. The corresponding averages in the subsarcolemmal regions averaged 5.8 and 4.7%, respectively. Therefore, the average volume fraction of mitochondria in the whole fibers increased from 4.5% in WT muscles to 7.0% in double-KO muscles (e.g., $4.5\% = 0.9 \times 4.3\% + 0.1 \times 5.8\%$). With double-KO muscles, which had the largest values of $[\text{Ca}^{2+}]_{\text{mito}}$ and mitochondrial volume fractions, the concentration of free Ca^{2+} in mitochondria with respect to fiber volume was 0.000016 mM, which is negligible ($0.000016 \text{ mM} = 0.00023 \text{ mM} \times 0.07$; concentration referred to fiber volume). The only way total Ca in mitochondria could have a nonnegligible resting Ca content is if mitochondria contained a high concentration of Ca-binding sites (a Ca-binding capacity comparable to that of CSQ) that had very high affinities for binding Ca^{2+} (like parvalbumin's). No such protein has been reported, so we conclude with near certainty that the contribution of mitochondria to non-SR Ca is negligible in both WT and double-KO muscles.

Simulations of BAPTA experiments and assessment of whether or not Ca was lost to parvalbumin

Table 3 simulates experimental results with the BAPTA method under three cases: Case 1: parvalbumin is unable to bind metals (Ca or Mg), and no Mg is present in the measurement solution; Case 2: parvalbumin can bind metals, and no Mg is present in the measurement solution; and Case 3: parvalbumin can bind metals and 2 mM of total Mg is present in the measurement solution. Based on the results from the section above with Table 2, Ca binding to SERCA, CSQ, and the Ca-regulatory sites on troponin should be negligible and is ignored. Although the simulations do not consider Ca binding to the Ca/Mg sites on troponin and myosin, similar results would be expected for these proteins, although significantly less than for parvalbumin (see Table 2). In brief, starting with assumed values for the total concentrations of the relevant ions and substances, the simulations calculate the concentrations of the Ca-bound, Mg-bound, and metal-free forms of the relevant substances before and after adding 1 mM EGTA. For each case, the predicted changes in concentration are then used to predict the associated absorbance changes, which are then summed to give the predicted, experimentally accessible value of $A_0 - A_M$ and its associated, predicted value of $[\text{Ca}_T]$.

Rows 1–5 of Table 3 give the assumed total concentrations of Ca, BAPTA, EGTA, parvalbumin, and Mg,

respectively, in terms of the concentrations in the final solution. Some of the starting assumptions are the same as those used and detailed above for Table 2. The concentration of 120 μM for total BAPTA was the same as that used for Table 2. The value of 21.234 μM for the concentration of total Ca was the sum of the concentrations of Ca bound to BAPTA (16 μM), free Ca (0.034 μM), and Ca bound to parvalbumin (5.2 μM) given in column 5 of Table 2. The value of 7.0 μM for the concentration of total parvalbumin (Table 3, row 4) is from column 3 of Table 2. Columns 2, 4, and 6 give the concentrations

for all of the components considered for the simulated A_M aliquot for cases 1–3, respectively. Columns 3, 5, and 7 give concentrations and related values with 1 mM EGTA added (row 3) for cases 1–3, respectively; these columns correspond to the A_0 aliquots associated with columns 2, 4, and 6, respectively. As seen in row 5, 2 mM of total Mg was present for case 3. The simulations involved adjusting the concentration of free Ca^{2+} to give the concentration of total Ca in row 1; the values of $[\text{Ca}^{2+}]$ are given in row 6. For case 3, the similarly adjusted values of $[\text{Mg}^{2+}]$ are given in row 7. The calculated

TABLE 3
Simulations of possible results of experiments to help assess if Ca was “lost” to parvalbumin

1	2	3	4	5	6	7
	Case 1: No Mg and no parvalbumin Muscle solution	Case 2: No Mg and with parvalbumin With EGTA	Case 2: No Mg and with parvalbumin Muscle solution	Case 2: No Mg and with parvalbumin With EGTA	Case 3: With both Mg and parvalbumin Muscle solution	Case 3: With both Mg and parvalbumin With EGTA
	(all concentrations in μM)					
	Rows 1–5 are assumed (typical or expected total values for sample)					
1	Ca_T	21.234	21.234	21.234	21.234	21.234
2	BAPTA_T	120.0	120.0	120.0	120.0	120.0
3	EGTA_T		1,000.0		1,000.0	1,000.0
4	Parv_T			7.0		7.0
5	Mg_T				2,000.0	2,000.0
	Ca^{2+} and Mg^{2+} values adjusted to give Ca_T and Mg_T values above					
6	Ca^{2+}	0.0472	8.9×10^{-5}	0.0339	8.9×10^{-5}	0.0495
7	Mg^{2+}				1,983.893	1590.948
	Rows 8–17 determined with above values for Ca^{2+} and Mg^{2+}					
8	BAPTA	98.8132	119.9513	103.9706	119.9514	89.449
10	CaBAPTA	21.1868	0.0487	16.0294	0.0486	20.113
11	MgBAPTA					10.439
12	EGTA		978.8149		978.8664	586.676
13	CaEGTA		21.1851		21.1336	21.152
14	MgEGTA					392.173
15	Parvalbumin			1.8293	6.9484	0.260
16	CaParvalbumin			5.1707	0.0516	1.072
17	MgParvalbumin					5.668
	Differences between values with and without EGTA					
18	ΔBAPTA		21.1381		15.9808	20.214
19	$\Delta\text{CaBAPTA}$		-21.1381		-15.9808	-20.039
20	$\Delta\text{MgBAPTA}$					-0.176
	Rows 21–24: Absorbance changes at 292 nm calculated from rows 18–20					
21	ΔA_{BAPTA}		0.1059		0.0800	0.1013
22	$\Delta A_{\text{CaBAPTA}}$		-0.0067		-0.0051	-0.0064
23	$\Delta A_{\text{MgBAPTA}}$					-0.0005
24	$\Delta A_{\text{total}} \text{ OR } A_0 - A_M$		0.0992		0.0749	0.0944
	Predicted $[\text{Ca}_T] = (A_0 - A_M) / (\Delta\epsilon \cdot l)$					
25	Predicted Ca_T		21.151		15.970	20.132
	Row 26 gives the value in row 25 divided by that in row 1					
26	Predicted/actual		0.996		0.752	0.948

This table provides simulations of experimentally predicted values of $[\text{Ca}_T]$ with and without Mg present and with and without metal binding to parvalbumin. The text provides most of the details for the assumed and predicted values. For the case of parvalbumin, Ca and Mg binding was calculated as described with Table 2. Relationships used to calculate Ca and Mg binding to BAPTA and EGTA are given in Section 1 of the supplemental text. For rows 21 and 22, values of 5,009 and 320 $\text{M}^{-1} \text{cm}^{-1}$, respectively, were used for the extinction coefficients at 292 nm of the metal-free and Ca-bound forms of BAPTA (ϵ_B and ϵ_{CaB} , respectively; see Materials and methods). The value of 2,665 $\text{M}^{-1} \text{cm}^{-1}$ assumed for the extinction coefficient for the Mg-bound form of BAPTA (ϵ_{MgB}) in row 23 was based on the observation of Tsien (1980) that the decrease in absorbance of BAPTA with Mg binding was about half of that produced by Ca binding ($2,665 = 320 + 0.5 \times (5,009 - 320)$). As used throughout, the value of $\Delta\epsilon$ used to calculate $[\text{Ca}_T]$ (row 25) from $A_0 - A_M$ (row 24) was $\epsilon_{\text{CaB}} - \epsilon_B$, i.e., $-4,689 \text{ M}^{-1} \text{cm}^{-1}$ ($-4,689 = 320 - 5,009$). Row 26 gives the ratios of the values in row 25 to those in row 1.

values of the metal-free, Ca-bound, and Mg-bound forms are given in rows 8–17. For these rows, Section 1 of the supplemental text gives the relationships used for the BAPTA and EGTA components, and Eqs. A1–A4 above give the relationships used for the parvalbumin components. Rows 18–20 give the changes in concentration of the BAPTA components, the only components giving measurable absorbance differences. The corresponding absorbance changes are given in rows 21–23, and the sum of these absorbance differences, given in row 24, is the predicted value for the absorbance difference ($A_0 - A_M$) that would be measured for each case. Row 25 gives the predicted value of the concentration of total Ca that would be reported with the BAPTA method, and row 26 gives the ratio of this reported value to the actual concentration of total Ca in row 1.

The main conclusions from the results in row 26 (also given in the last section of Results) are that the BAPTA method should report almost all (>99%) of the total Ca if parvalbumin is unable to bind Ca (case 1). If the high affinity of parvalbumin for Ca was unaffected by the denaturing effect of the SDS treatment and Mg is not added, the BAPTA method is predicted to underestimate $[Ca_T]$ by ~25% because of “loss” of Ca to parvalbumin (case 2). Adding 2 mM Mg to the solution would greatly minimize this loss with a prediction of ~95% of the total Ca being reported (case 3). Another result from the simulations is that EGTA competes essentially all (>99.5%) of the Ca from BAPTA and parvalbumin, if the latter is able to bind Ca (compare values in row 13 with those in rows 10 and 16).

The contribution of Mg from muscle samples is negligible. This section justifies ignoring Mg from the muscle sample for the simulations in Table 3 and for the BAPTA method in general. Assuming that the high-end estimate of 15.5 mmoles/kg for total Mg in frog muscle (Godt and Maughan, 1988) applies to mouse muscle, the concentration of total Mg in the final volume for the example above (the value that would go in row 5 of Table 3) would be 0.10 mM ($0.10 \text{ mM} = 15.5 \text{ mmoles/kg} \times 8 \text{ mg} \times 10^{-6} \text{ kg/kg} \div 1.2 \text{ ml} \times 10^3 \text{ ml/L}$). As shown by comparing the value in column 7 for row 23 to that for row 24 in Table 3, the contribution of the added 2.0 mM of total Mg for Case 3 would lead to an underestimate of ΔA for BAPTA of only 0.5% ($-0.5\% = 100 \times -0.0005 \div 0.0944$). Because the 0.1 mM of Mg from the muscle is 1/20,000th of the added 2.0 mM for Case 3, the error would be ~2,000-fold less than 0.5% and, therefore, negligible.

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REFERENCES

- Baylor, S.M., and S. Hollingworth. 1998. Model of sarcomeric Ca^{2+} movements, including ATP Ca^{2+} binding and diffusion, during activation of frog skeletal muscle. *J. Gen. Physiol.* 112:297–316. <http://dx.doi.org/10.1085/jgp.112.3.297>
- Baylor, S.M., and S. Hollingworth. 2007. Simulation of Ca^{2+} movements within the sarcomere of fast-twitch mouse fibers stimulated by action potentials. *J. Gen. Physiol.* 130:283–302. <http://dx.doi.org/10.1085/jgp.200709827>
- Baylor, S.M., W.K. Chandler, and M.W. Marshall. 1983. Sarcoplasmic reticulum calcium release in frog skeletal muscle fibres estimated from Arsenazo III calcium transients. *J. Physiol.* 344:625–666. <http://dx.doi.org/10.1113/jphysiol.1983.sp014959>
- Canato, M., M. Scorzeto, M. Giacomello, F. Protasi, C. Reggiani, and G.J.M. Stienen. 2010. Massive alterations of sarcoplasmic reticulum free calcium in skeletal muscle fibers lacking calsequestrin revealed by a genetically encoded probe. *Proc. Natl. Acad. Sci. USA.* 107:22326–22331. <http://dx.doi.org/10.1073/pnas.1009168108>
- Caputo, C., and P. Bolaños. 1994. Fluo-3 signals associated with potassium contractures in single amphibian muscle fibres. *J. Physiol.* 481:119–128. <http://dx.doi.org/10.1113/jphysiol.1994.sp020423>
- Celichowski, J., and K. Grottel. 1993. Twitch/tetanus ratio and its relation to other properties of motor units. *Neuroreport.* 5:201–204. <http://dx.doi.org/10.1097/00001756-199312000-00003>
- Cieslar, J., M.T. Huang, and G.P. Dobson. 1998. Tissue spaces in rat heart, liver, and skeletal muscle in vivo. *Am. J. Physiol.* 275:R1530–R1536.
- Cota, G., and C.M. Armstrong. 1988. Potassium channel “inactivation” induced by soft-glass patch pipettes. *Biophys. J.* 53:107–109. [http://dx.doi.org/10.1016/S0006-3495\(88\)83071-6](http://dx.doi.org/10.1016/S0006-3495(88)83071-6)
- Cox, J.A., D.R. Winge, and E.A. Stein. 1979. Calcium, magnesium and the conformation of parvalbumin during muscular activity. *Biochimie.* 61:601–605. [http://dx.doi.org/10.1016/S0300-9084\(79\)80157-1](http://dx.doi.org/10.1016/S0300-9084(79)80157-1)
- Everts, M.E., T. Lømo, and T. Clausen. 1993. Changes in K^+ , Na^+ and calcium contents during in vivo stimulation of rat skeletal muscle. *Acta Physiol. Scand.* 147:357–368. <http://dx.doi.org/10.1111/j.1748-1716.1993.tb09512.x>
- Fénelon, K., C.R.H. Lamboley, N. Carrier, and P.C. Pape. 2012. Calcium buffering properties of sarcoplasmic reticulum and calcium-induced Ca^{2+} release during the quasi-steady level of release in twitch fibers from frog skeletal muscle. *J. Gen. Physiol.* 140:403–419. <http://dx.doi.org/10.1085/jgp.201110730>
- Fenn, W.O. 1936. Electrolytes in muscle. *Physiol. Rev.* 16:450–486.
- Fryer, M.W., and D.G. Stephenson. 1996. Total and sarcoplasmic reticulum calcium contents of skinned fibres from rat skeletal muscle. *J. Physiol.* 493:357–370. <http://dx.doi.org/10.1113/jphysiol.1996.sp021388>
- Gissel, H., and T. Clausen. 1999. Excitation-induced Ca^{2+} uptake in rat skeletal muscle. *Am. J. Physiol.* 276:R331–R339.
- Glesby, M.J., E. Rosenmann, E.G. Nylén, and K. Wrogemann. 1988. Serum CK, calcium, magnesium, and oxidative phosphorylation in mdx mouse muscular dystrophy. *Muscle Nerve.* 11:852–856. <http://dx.doi.org/10.1002/mus.880110809>
- Godt, R.E., and D.W. Maughan. 1988. On the composition of the cytosol of relaxed skeletal muscle of the frog. *Am. J. Physiol.* 254:C591–C604.

- Harrison, S.M., and D.M. Bers. 1987. The effect of temperature and ionic strength on the apparent Ca-affinity of EGTA and the analogous Ca-chelators BAPTA and dibromo-BAPTA. *Biochim. Biophys. Acta.* 925:133–143. [http://dx.doi.org/10.1016/0304-4165\(87\)90102-4](http://dx.doi.org/10.1016/0304-4165(87)90102-4)
- Heizmann, C.W., M.W. Berchtold, and A.M. Rowlerson. 1982. Correlation of parvalbumin concentration with relaxation speed in mammalian muscles. *Proc. Natl. Acad. Sci. USA.* 79:7243–7247. <http://dx.doi.org/10.1073/pnas.79.23.7243>
- Hollingworth, S., M.M. Kim, and S.M. Baylor. 2012. Measurement and simulation of myoplasmic calcium transients in mouse slow-twitch muscle fibres. *J. Physiol.* 590:575–594. <http://dx.doi.org/10.1113/jphysiol.2011.220780>
- Inesi, G., M. Kurzmack, C. Coan, and D.E. Lewis. 1980. Cooperative calcium binding and ATPase activation in sarcoplasmic reticulum vesicles. *J. Biol. Chem.* 255:3025–3031.
- Irving, M., J. Maylie, N.L. Sizto, and W.K. Chandler. 1987. Intrinsic optical and passive electrical properties of cut frog twitch fibers. *J. Gen. Physiol.* 89:1–40. <http://dx.doi.org/10.1085/jgp.89.1.1>
- Kake-Guena, S.A., C.R.H. Lamboley, P. Bouchard, J. Frenette, E. Rousseau, R.T. Dirksen, F. Protasi, and P.C. Pape. 2014. New method for determining the total calcium content of tissue applied to whole skeletal muscles from mice with and without calsequestrin knocked out. *Biophys. J.* 102:731a.
- Kirby, A.C., B.D. Lindley, and J.R. Picken. 1975. Calcium content and exchange in frog skeletal muscle. *J. Physiol.* 253:37–52. <http://dx.doi.org/10.1113/jphysiol.1975.sp011178>
- Lamboley, C.R., R.M. Murphy, M.J. McKenna, and G.D. Lamb. 2013b. Endogenous and maximal sarcoplasmic reticulum calcium content and calsequestrin expression in type I and type II human skeletal muscle fibres. *J. Physiol.* 591:6053–6068. <http://dx.doi.org/10.1113/jphysiol.2013.265900>
- MacLennan, D.H., and P.T.S. Wong. 1971. Isolation of a calcium-sequestering protein from sarcoplasmic reticulum. *Proc. Natl. Acad. Sci. USA.* 68:1231–1235. <http://dx.doi.org/10.1073/pnas.68.6.1231>
- Mendez, J., and A. Keys. 1960. Density and composition of mammalian muscle. *Metabolism.* 9:184–188.
- Murphy, R.M., N.T. Larkins, J.P. Mollica, N.A. Beard, and G.D. Lamb. 2009. Calsequestrin content and SERCA determine normal and maximal Ca²⁺ storage levels in sarcoplasmic reticulum of fast- and slow-twitch fibres of rat. *J. Physiol.* 587:443–460. <http://dx.doi.org/10.1113/jphysiol.2008.163162>
- Paolini, C., M. Quarta, A. Nori, S. Boncompagni, M. Canato, P. Volpe, P.D. Allen, C. Reggiani, and F. Protasi. 2007. Reorganized stores and impaired calcium handling in skeletal muscle of mice lacking calsequestrin-1. *J. Physiol.* 583:767–784. <http://dx.doi.org/10.1113/jphysiol.2007.138024>
- Paolini, C., M. Quarta, L. D'Onofrio, C. Reggiani, and F. Protasi. 2011. Differential effect of calsequestrin ablation on structure and function of fast and slow skeletal muscle fibers. *J. Biomed. Biotechnol.* 2011:1–10. <http://dx.doi.org/10.1155/2011/634075>
- Pape, P.C., D.-S. Jong, and W.K. Chandler. 1995. Calcium release and its voltage dependence in frog cut muscle fibers equilibrated with 20 mM EGTA. *J. Gen. Physiol.* 106:259–336. <http://dx.doi.org/10.1085/jgp.106.2.259>
- Pape, P.C., K. Fénelon, C.R.H. Lamboley, and D. Stachura. 2007. Role of calsequestrin evaluated from changes in free and total calcium concentrations in the sarcoplasmic reticulum of frog cut skeletal muscle fibres. *J. Physiol.* 581:319–367. <http://dx.doi.org/10.1113/jphysiol.2006.126474>
- Peachey, L.D., and A.F. Huxley. 1962. Structural identification of twitch and slow striated muscle fibers of the frog. *J. Cell Biol.* 13:177–180. <http://dx.doi.org/10.1083/jcb.13.1.177>
- Posterino, G.S., and G.D. Lamb. 2003. Effect of sarcoplasmic reticulum Ca²⁺ content on action potential-induced Ca²⁺ release in rat skeletal muscle fibres. *J. Physiol.* 551:219–237. <http://dx.doi.org/10.1113/jphysiol.2003.040022>
- Protasi, F., C. Paolini, and M. Dainese. 2009. Calsequestrin-1: a new candidate gene for malignant hyperthermia and exertional/environmental heat stroke. *J. Physiol.* 587:3095–3100. <http://dx.doi.org/10.1113/jphysiol.2009.171967>
- Ringer, S. 1883. A further contribution regarding the influence of the different constituents of the blood on the contraction of the heart. *J. Physiol.* 4:29–42. <http://dx.doi.org/10.1113/jphysiol.1883.sp000120>
- Robertson, S.P., J.D. Johnson, and J.D. Potter. 1981. The time-course of Ca²⁺ exchange with calmodulin, troponin, parvalbumin, and myosin in response to transient increases in Ca²⁺. *Biophys. J.* 34:559–569. [http://dx.doi.org/10.1016/S0006-3495\(81\)84868-0](http://dx.doi.org/10.1016/S0006-3495(81)84868-0)
- Rojas, L., and C. Zuazaga. 1988. Influence of the patch pipette glass on single acetylcholine channels recorded from *Xenopus* myocytes. *Neurosci. Lett.* 88:39–44. [http://dx.doi.org/10.1016/0304-3940\(88\)90312-6](http://dx.doi.org/10.1016/0304-3940(88)90312-6)
- Scorzeto, M., M. Giacomello, L. Toniolo, M. Canato, B. Blaauw, C. Paolini, F. Protasi, C. Reggiani, and G.J.M. Stienen. 2013. Mitochondrial Ca²⁺-handling in fast skeletal muscle fibers from wild type and calsequestrin-null mice. *PLoS ONE.* 8:e74919. <http://dx.doi.org/10.1371/journal.pone.0074919>
- Sztretze, M., J. Yi, L. Figueroa, J. Zhou, L. Royer, P. Allen, G. Brum, and E. Ríos. 2011. Measurement of RyR permeability reveals a role of calsequestrin in termination of SR Ca²⁺ release in skeletal muscle. *J. Gen. Physiol.* 138:231–247. <http://dx.doi.org/10.1085/jgp.201010592>
- Tsien, R.Y. 1980. New calcium indicators and buffers with high selectivity against magnesium and protons: design, synthesis, and properties of prototype structures. *Biochemistry.* 19:2396–2404. <http://dx.doi.org/10.1021/bi00552a018>
- Yoshida, M., A. Yonetani, T. Shirasaki, and K. Wada. 2006. Dietary NaCl supplementation prevents muscle necrosis in a mouse model of Duchenne muscular dystrophy. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 290:R449–R455. <http://dx.doi.org/10.1152/ajpregu.00684.2004>
- Zhang, L., J. Kelley, G. Schmeisser, Y.M. Kobayashi, and L.R. Jones. 1997. Complex formation between junctin, triadin, calsequestrin, and the ryanodine receptor. Proteins of the cardiac junctional sarcoplasmic reticulum membrane. *J. Biol. Chem.* 272:23389–23397. <http://dx.doi.org/10.1074/jbc.272.37.23389>