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Interaction and Quantification of Thymol in Active PLA-Based Materials Containing Natural Fibres

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ABSTRACT

The quantification of thymol, a commercial essential oil extract that is an antimicrobial (AM) agent, in poly(lactic acid) (PLA) and PLA/kenaf composites was investigated to explore the potential of these systems as AM food packaging materials. Neat PLA and PLA/kenaf composites containing thymol (5-10 wt%) were prepared *via* melt blending and compression molding. The quantification of the thymol in PLA and PLA/kenaf composites after processing as well as the interactions between the PLA matrix, kenaf fibres and the AM agent were investigated. The PLA/kenaf composites in the range of 10 to 40 wt% fibre content retained less thymol upon processing than PLA alone and the composites containing the highest fibre loadings demonstrated the lowest thymol retention. The observed losses were attributed to the higher mechanical shear that exists during the mixing process as well as the creation of voids in the composites that facilitate the release of thymol from the system.

INTRODUCTION

The utilization of biocomposites for active food packaging is currently under investigation with a major purpose being to reduce environmental pollution as well as recover biodegradable polymers.¹⁻³ Nowadays, renewable polymers such as PLA that are derived and synthesized from plant materials are widely used for films and coatings as well as

matrices for incorporating naturally sourced additives such as antimicrobial (AM) agents that prolong the shelf life of packaged food products.⁴⁻⁷ Several additives have been incorporated directly into polymers including organic acids, enzymes, bacteriocins, chelators and a range of plant extracts.⁸⁻¹¹

For food-packaging applications, the concept of AM agent migration is used to provide continuous AM activity to food products. This can be achieved by using volatile additives derived from plant extracts (e.g. essential oils) whereby these natural agents are considered to be much safer than synthetically derived chemical agents.¹² Thymol and carvacrol which are the major constituents of thyme essential oil can act as antioxidants and AM agents. These compounds are amongst the most currently studied natural additives that can be incorporated into packaging materials.¹³⁻¹⁷ However, essential oils have low thermal stability and high volatility and so their exposure to high temperature, shearing and pressure during processing (e.g. extrusion, injection and blown moulding) often results in their loss from the matrix and consequently a reduction in the AM activity of the system.¹⁵ For instance, extruded PLA demonstrated a loss of thymol with lower inhibition of *Listeria monocytogenes*.¹¹ Furthermore, PLA has a relatively lower melting temperature than many commercial food packaging materials such as poly(ethylene terephthalate) (PET). Nevertheless, the processing temperature of PLA using an extruder is normally greater than 150°C and this is crucial to ensure optimal melt viscosity as well as the complete melting of the crystalline phase in the matrix during extrusion.¹⁸ Such conditions enable the AM agent to be evenly distributed in the amorphous regions of the polymeric material and thus regulate a slow release of the agent from the film.¹⁹

Plasticizers and fillers such as polyethylene glycol (PEG), natural fibres and nanofillers are used to facilitate the controlled release and to increase the activity of AM agents. Recent studies by Ramos *et al.*²⁰ reported an increment in the thermal stability of the essential oil thymol upon processing due to the incorporation of nanoclay (e.g. montmorillonite) into active PLA films where the nanoclay was incorporated to control the thymol release in the active films. Liu *et al.*²¹ reported that the incorporation of plasticizers during the extrusion process lowered the temperature profile during the manufacturing of AM films. In that study, PLA/Nisaplin films showed no AM activity whereas PLA blended with the lactide dimer of PLA and PLA plasticized with glycerol triacetate (GTA) that were used to create membranes containing Nisaplin each prevented the growth of *L. monocytogenes* in brain-heart infusion (BHI) broth. Praprudivongs and Sombatsompop²² found that a higher loading of wood flour (10 wt%) resulted in facilitating the release of more triclosan onto a PLA composite surface due to the hydrophilic nature of the wood flour causing water molecules to be absorbed by the surface of the composite. Similar findings were reported by Woraprayote *et al.*²³ where sawdust particles helped to embed pediocin into coated PLA composites and significantly inhibited the growth of *L. monocytogenes* in agar media and sliced pork mince. Nevertheless, in these studies, there is little information on the quantification of AM agents after the processing of the active PLA-based materials containing natural fibres.

It is clear that more research needs to be conducted to investigate the quantification of AM agents when high temperature, shear and pressure are applied during processing. The release profile and AM inhibition activity in order to produce efficient active films or materials also requires further investigation along with the possible interaction of the AM

agent with other additives in the matrix. To date, it appears that no study in the literature has systematically addressed the quantification of thymol or other essential oil AM agents after plastic thermal processing for PLA-based materials containing natural fibres. In the current study a natural fibrous substance, namely kenaf fibre (*Hibiscus cannabinus L.*), is of interest as a reinforcing filler for PLA-based composites. The composite material is expected to be advantageous when compared to many synthetic composite materials due to the renewability of the raw materials from which it is comprised and its propensity to be environmentally friendly.^{2,24} Moreover, the incorporation of the natural fibres as a filler in the biopolymer can also improve its mechanical properties, reduce abrasion resistance during processing, promote good compatibility and enhance biodegradability.^{25,26} Tawakkal *et al.*⁷ reported that the PLA/kenaf composites demonstrated improved mechanical properties such as tensile strength and stiffness compared with commercial PLA due to the reinforcement of the kenaf fibres in the PLA matrix. However, the increased stiffness and hence less flexibility means that these materials are perhaps more suitable to be developed as rigid food packaging materials.

The objective of this study is to quantitatively investigate the retention of thymol incorporated into PLA and PLA/kenaf composite films during processing as well as the thermal release kinetics of thymol from these materials. These parameters are of importance in order to understand the interactions amongst the matrix, natural fibres and AM agents in the films and the ability of the films to initiate and maintain effective AM activity.

EXPERIMENTAL

Materials

Poly(lactic acid) (7001D Ingeo™; specific gravity 1.24; melt flow index (MFI) 6 g/10 min at 210°C and 2.16 kg; melting temperature range 145-160°C) was obtained from NatureWorks LLC, USA. Mechanically separated kenaf fibre (bast) was purchased from Ecofibre Industries, Australia. The thymol (T0501, purity of 99.5%) was purchased from Sigma Aldrich Pty. Ltd., Australia. Sodium hydroxide and acetic acid were purchased from Merck Chemicals, Australia. Un-denatured ethanol was purchased from Chem-Supply Pty Ltd., Australia. Isooctane (2,2,4-trimethylpentane, 36006) was purchased from Sigma Aldrich, Australia.

Preparation of Active PLA/Kenaf/Thymol Composites

The kenaf fibre surface treatment was performed by immersing fibres in 5% w/v sodium hydroxide (NaOH) for 2 h at room temperature. Acetic acid was used to adjust the pH (until neutralized) during the process of washing and rinsing the fibres with distilled water.

Treated kenaf (TK) fibres were filtered from the solution, washed, and later dried overnight in an air oven at 105°C. The dried fibres were then ground and sieved using a 300-500 µm aperture sieve. The aspect ratio (L/D) of the kenaf fibres was approximately 9 with an average length of 920 µm and an average diameter of 104 µm. The preparation of untreated kenaf (UK) and TK doped with thymol was performed by immersing 20 g of fibres in 800 mL of 10-25% v/w thymol/ethanol solution and stirring for 1-2 h. The doped fibres were then filtered from the solution and dried overnight in a laminar flow cabinet in order to evaporate the remaining ethanol. The micrograph images of TK and TK doped with 25 wt% thymol can be seen in Figure 1. These show that the doping of the TK was successful as a smooth fibre surface is observed on the doped TK sample. Prior to mixing, the PLA resin was

dried in an oven at 60°C overnight before blending with kenaf (undoped and doped with thymol) at various concentrations in the range zero to 40 wt%.

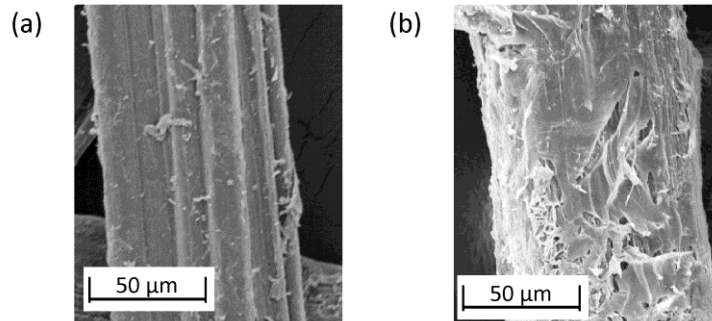


Figure 1 Scanning electron micrographs of: (a) TK fibre at 500× magnification and (b) TK fibre doped with 25 wt% thymol at 500× magnification.

To produce active PLA as well as PLA/kenaf films, the PLA, UK or TK fibres (zero to 40 wt%) or thymol were compounded using an internal mixer (Haake PolyLab OS, Germany) at 50 rpm and at a processing temperature of 155°C for 8-10 min. The samples were prepared by using a laboratory press (L0003, IDM Instrument Pty. Ltd., Australia). The PLA and composites were preheated at 150°C for 2 min and pressed at the same temperature for 3 min under a pressure of 50 kN before quench cooling to 30°C under pressure. The average thickness of the heat pressed PLA and PLA/kenaf films incorporated with thymol were 0.19 ± 0.03 and 0.25 ± 0.05 respectively. A hand-held micrometer (Hahn & Kolb, Stuttgart, Germany) was used for measuring the film thickness.

Infrared Analyses

The infrared spectral analysis of PLA and PLA/kenaf composite film samples was performed using a Shimadzu IR Prestige Fourier transform infrared (FTIR) spectrophotometer and utilizing the attenuated total reflectance (ATR) technique. For thymol and kenaf fibre

samples, a small portion of thymol or kenaf fibre powder was mixed in an agate mortar and pestle with a few drops of paraffin oil. The sample was then applied to a KBr disc and its FTIR spectrum recorded. All spectra were recorded in absorbance mode in the range of 550-4000 cm^{-1} with a resolution of 4 cm^{-1} and with 32 scans recorded at every point using Happ-Genzel apodization. Ten scans were performed for each acquisition.

Thermogravimetric Analysis

A Mettler Toledo (TGA/DSC 1 Star System) was used to undertake the thermogravimetric (TG) analyses. The weight percentage of thymol that was retained in the samples after processing was measured from the normalized weight loss curve and the derivative of the weight loss curve, the latter being used to identify the start and end points of the process. The PLA and PLA/kenaf composite samples containing thymol were heated from 30 to 500°C at a heating rate of 5°C min^{-1} and under a nitrogen atmosphere flow rate of 0.2 L min^{-1} .

Thymol Quantification in PLA and PLA/Kenaf Composites

Reflux extraction followed by gas chromatography (GC) was used to analytically determine the weight percentage of thymol that was retained in the samples after processing. One gram of compressed sample was extracted at 100°C for 2-5 h using 100 mL of isooctane or 95% ethanol. An aliquot of the solution was analyzed using GC. The conditions applied in the GC instrument were as follows: injected volume, 1.0 μL ; initial column temperature, 80°C; heating rate, 5°C min^{-1} up to 120°C, held at this temperature for an additional 5 min; injector temperature, 250°C; FID detector temperature, 300°C; flow rate, 2 mL min^{-1} ; splitting; carrier gas, nitrogen. A standard curve for thymol was also prepared and the thymol content of the samples was calculated using this curve.

Thymol Release using TG Kinetic Analyses

The application of non-isothermal techniques for the determination of kinetic parameters of reaction measured by loss in weight has been long established.²⁷ In the current study, non-isothermal TG kinetic analyses of the release of thymol from PLA and PLA/kenaf composite samples were performed by a computer-based iterative numerical method using original software. The software was developed to execute an integral solution of the general kinetic equation pertaining to TG analysis:

$$g(\alpha) = (AE_a/R\beta) \times p(x) \quad (1)$$

where α is the degree of conversion at time t in the process, A is the Arrhenius A-factor, E_a is the apparent activation energy for the process, R is the ideal gas constant and β is the heating rate. The function $p(x)$ represents the integral:

$$p(x) = \int_x^{\infty} [\exp(-x)/x^2] dx \quad (2)$$

where $x = E_a/RT$ and T is the absolute temperature. The data pertaining to the release of thymol were analyzed according to a 3D diffusional model²⁸ and an algorithm developed from the work of Dollimore *et al.*²⁹ was used to confirm that this model was the most appropriate one needed to fit the data. For a 3D diffusion model:

$$g(\alpha) = [1 - (1 - \alpha)^{1/3}]^2 \quad (3)$$

All TGA profiles were analyzed up to 85% conversion with respect to the corresponding first step in the TG analysis profile in order to extract the apparent activation energy and Arrhenius A-Factor data.

Morphology of Fibres and Composites

Scanning electron microscopy (SEM) was conducted to observe the morphology of TK, TK doped with thymol as well as that of the composites. The composite films were immersed in liquid nitrogen and then fractured in order to create a fracture surface of the films for observation. All micrographs were obtained using a JOEL NeoScope (JCM-5000) scanning electron microscope. Samples were coated with a thin layer of gold (6 nm) using a NeoCoater (MP19020NCTR) device under high vacuum and using an optimal accelerating voltage of 10 kV to avoid charging effects.

RESULTS AND DISCUSSION

Structural Analysis

Figure 2 shows the FTIR spectra of the neat PLA, thymol and TK fibres. The absence of the O–H stretching band in the spectrum of the neat PLA confirms that this particular grade of PLA (7001D) is hydrophobic. The carbonyl ($>C=O$) stretching peak at 1746 cm^{-1} is due to the carbonyl group in the lactic acid ester moiety of PLA.³⁰ The deformational vibrations of C–H in the methyl group of PLA are also observed in the range of $1300\text{ to }1500\text{ cm}^{-1}$.³⁰ The peak at 1180 cm^{-1} is due to the stretching vibration of C–O–C and another asymmetric stretching vibration of C–O–C is observed in the range of $1150\text{-}1060\text{ cm}^{-1}$.³¹

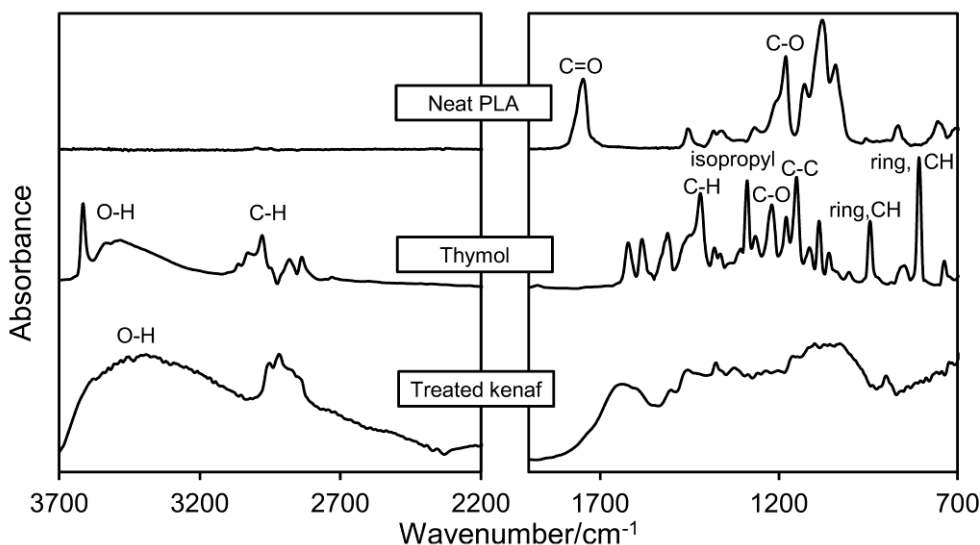


Figure 2 FTIR spectra of PLA, thymol and TK fibre.

The FTIR spectrum of thymol exhibits a number of major peaks as seen in Figure 2 and these are also tabulated in Table 1 along with the major peaks observed in the spectra of PLA and PLA/TK composites containing thymol. The following spectral features are apparent in Figure 2: a broad band due to the O–H hydroxyl group stretching vibration appears in the range of 3400-3500 cm^{-1} ; the C–H methyl group stretching at 2945 cm^{-1} ; a strong absorption due to the phenolic C–O stretching in the region at 1215 cm^{-1} ; C–C stretching at 1419 cm^{-1} ; and strong peaks due to isopropyl stretching and ring aromatic C–H bending at 1288 cm^{-1} and 810 cm^{-1} respectively.³² Moreover, the FTIR spectrum of the TK fibres exhibits a broad O–H band at 3500-3400 cm^{-1} with the expected absence of a sharp, carbonyl group absorption at approximately 1700 cm^{-1} . This is due to the removal of ester groups in the hemicellulose during the alkali treatment of the surface of the kenaf fibres.³³ A similar observation was reported by Himmelsbach *et al.*³⁴ who found that the ester groups of

hemicellulose or the ester linkage of the carboxylic group of ferulic and *p*-coumaric acids of lignin and/or hemicellulose disappeared in the spectrum of cellulose fibres.

Table 1. Major peaks of thymol, PLA and PLA containing thymol and TK fibres.

Functional group	Thymol	PLA	PLA/thymol ^a	PLA/TK ^b	PLA/thymol/TK ^c	Notes
OH	3483.6	-	3510.9	3505.8	3514.5	significant shift of thymol O-H band incorporated in composite; peak broadening with presence of TK
CH	2978.2	2951.2	2945.4	2951.2	2947.4	significant shift of thymol C-H stretch when incorporated in PLA and PLA/kenaf
C=O	-	1747.6	1750.5	1747.6	1755.4	small shift in C=O absorption across the active composites; peak broadening in active composites <i>cf.</i> neat PLA
ring CH	945.2	-	947.1	-	947.1	small shift of thymol ring C-H stretch when incorporated in PLA and PLA/kenaf
CH	2881.8	-	2875.9	-	2875.9	no significant shift
ring	1622.2	-	1620.3	-	1618.4	no significant shift; weak peak
ring	1581.7	-	1585.6	-	1585.6	no significant shift; weak peak
CC	1419.7	-	1423.5	-	1419.7	no significant shift; weak peak
CH or CC isopropyl	1288.5	-	1292.4	-	1292.4	no significant shift; weak peak
COC	-	1180.5	1180.5	1180.5	1182.5	no significant shift; peak broadening in composites <i>cf.</i> neat PLA
COC	-	1078.3	1082.1	1078.3	1084.1	no significant shift; peak broadening in composites <i>cf.</i> neat PLA
ring CH	808.2	-	810.1	-	810.1	no significant shift

^a thymol at 20 wt%; ^b TK fibre at 20 wt%; ^c PLA containing 20 wt% TK fibre and 20 wt% thymol

From Table 1, it can be noted that the spectra of the PLA and PLA/TK composites containing thymol are similar. This could be due to the high content of PLA present in the surface of the pressed films. Nevertheless, each of the PLA and PLA/TK composites containing 20 wt% thymol demonstrated a significant shift of O–H group absorptions that appear in the regions of approximately 3510 cm^{-1} respectively as compared with the thymol spectrum. This suggests that the thymol interacts with the PLA and/or the TK fibres. Intermolecular hydrogen bonding is presumed to exist between thymol and TK as well as thymol and PLA. Furthermore, hydrogen bonding is likely to be present in the composite between the hydroxyl groups in the TK fibres and the terminal hydroxyl groups of PLA,³⁵ the carbonyl groups of the ester linkages of PLA³⁶ as well as the thymol terminal hydroxyl group. The FTIR spectrum of the PLA/TK composite containing 20 wt% TK and 20 wt% thymol was similar to that of the PLA containing 20 wt% thymol (see Table 1). A small but noticeable peak broadening of the hydroxyl group absorption was observed in the active PLA/TK composite compared to PLA containing 20 wt% thymol. This is attributable to the presence of the hydrophilic TK fibres in the composite (see Figure 3(a)).

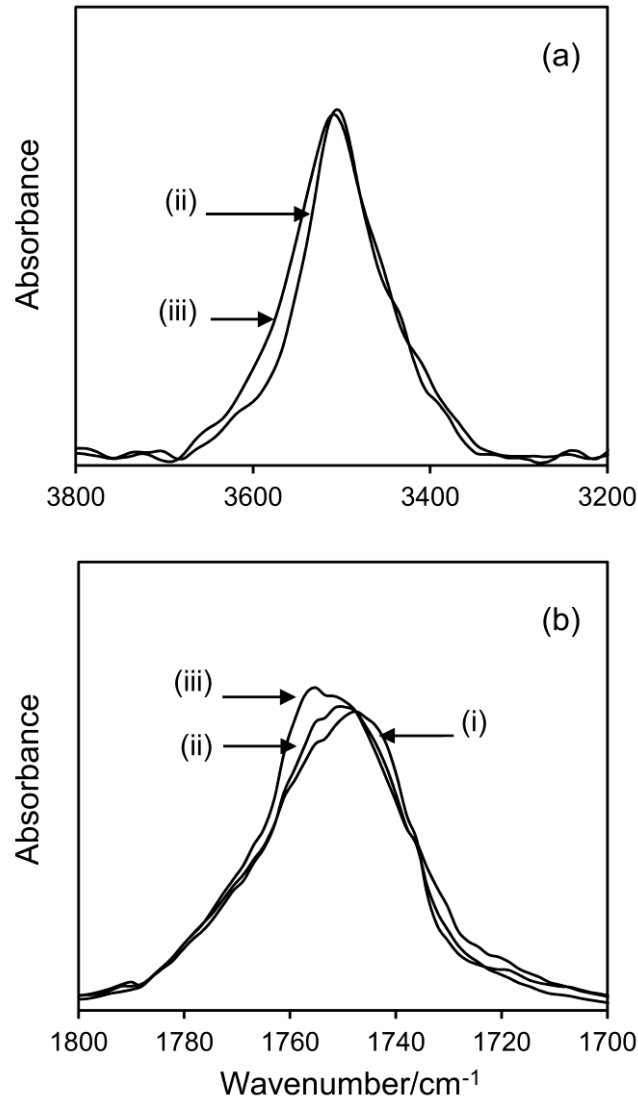


Figure 3 FTIR spectra showing: (a) hydroxyl group and (b) carbonyl group absorptions for: (i) neat PLA, (ii) PLA containing 20 wt% thymol, and (iii) PLA/TK fibre composite containing 20 wt% thymol.

Figure 3(b) shows the normalized carbonyl absorptions of neat PLA and PLA/TK composites containing 20 wt% thymol, where a slight hypsochromic shift in the carbonyl peak (at 1755 cm⁻¹) of the PLA is observed upon the addition of thymol to the PLA. A small shoulder on the peak is observed in the carbonyl group absorption at 1750 cm⁻¹ for PLA containing 20

wt% thymol and this shoulder becomes more pronounced with the presence of TK fibres in the film. The peak shift and presence of the peak shoulder support the notion of an intermolecular interaction existing between the PLA and thymol. A similar observation was made by Praprudivongs and Sombatsompop²² who investigated the interaction between PLA, 10 wt% wood flour and 1.5 wt% triclosan by using FTIR. They reported that the incorporation of triclosan and wood flour into PLA broadened the carbonyl absorbance peak and caused carbonyl peak splitting at wavenumbers of 1753 and 1746 cm^{-1} . The data listed in Table 1 suggest that overall the PLA/TK composite demonstrates similar spectral features to those of neat PLA. The hydroxyl group absorption at a wavenumber of *ca.* 3515 cm^{-1} is possibly due to the low amount of fibres on the surface of the pressed film. These fibres are expected to create a surface roughness and may inhibit the resolution of the ATR technique compared to the case of PLA alone.

Thermal Analysis

Figure 4 shows the normalized weight loss as a function of temperature for PLA composites containing TK and 10 wt% thymol. The profiles typically show an initial step that occurs over the temperature range of *ca.* 90 to 300°C and corresponds to the degradation/release of thymol from the matrix.¹³ The second, more pronounced step at 300-370°C corresponds to the degradation of the PLA that presumably occurs by thermal depolymerization and decomposition.³⁷ As the TK loading in the formulation is increased the level of char remaining in the system at elevated temperatures (*ca.* 390°C and above) is observed to increase accordingly.

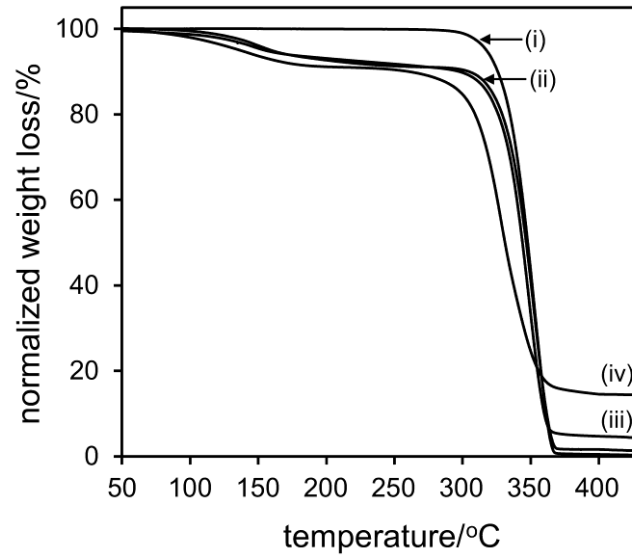


Figure 4 Normalized weight loss as a function of temperature for: (i) neat PLA and PLA containing: (ii) 10 wt% thymol, (iii) 10 wt% TK and 10 wt% thymol and (iv) 40 wt% TK and 10 wt% thymol. The thermograms were obtained using a heating rate of $5^{\circ}\text{C min}^{-1}$.

The quantification of thymol in the PLA and PLA/TK composites can be calculated from the TG profile as in the work of Ramos *et al.*²⁰ who investigated the retention of thymol in a PLA matrix and found that there was some loss of this volatile AM agent during processing. An example of a detailed analysis of the TG profiles obtained in the current study is shown in Figure 5 that shows the first derivative with respect to temperature, dw/dT , of the TG weight loss profiles. The value of the initial temperature at which thymol is released, T_{rel} , decreases by *ca.* 3.8°C upon the addition of 40 wt% TK to the formulation (see Figure 5) suggesting that the addition of TK to the polymer facilitates the loss of thymol from the PLA matrix. Similarly, a smaller decrease of *ca.* 1.7°C in T_{rel} is also observed in the case where 10 wt% of TK is present in the formulation although this decrease may not be significant. These findings are also supported by the observation that the temperature at which the

maximum rate of degradation of PLA occurs, T_{deg} , is lower in the case of the sample with the highest loading of fibre than in the case of PLA alone.

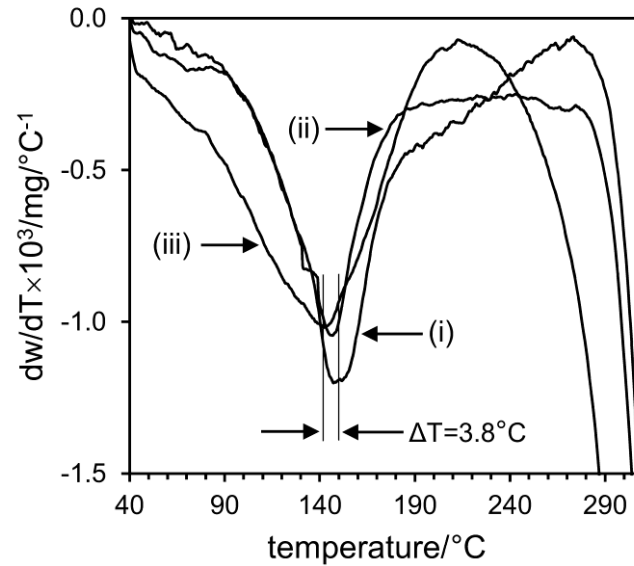


Figure 5 Plots of the first derivative with respect to temperature, dw/dT , of the respective TG weight loss profiles shown in Figure 4 for PLA containing: (i) 10 wt% thymol, (ii) 10 wt% TK and 10 wt% thymol and (iii) 40 wt% TK and 10 wt% thymol.

The values of T_{rel} , T_{deg} and the percentage of char residue of the various formulations were determined from the complete TG analyses and these are summarized in Table 2. Overall, it was found that the T_{rel} value of PLA/TK composites decreased with increasing TK fibre loading from 10% to 40 wt% over the temperature range of 149.9 to 144.0°C. Thus, it is clear from the results in Table 2 that the addition of TK to the formulation decreases the temperature at which thymol is released from the matrix at maximum rate and also decreases the temperature at which the maximum rate of degradation of the PLA occurs under the temperature ramp. The latter suggests that the addition of fibre destabilizes the polymer to some extent.³⁸ Moreover, the addition of 5-10 wt% thymol has no significant

effect on the value of the T_{deg} of PLA and this finding is in agreement with the work of Ramos *et al.*²⁰ who investigated the TG properties of PLA containing 8 wt% thymol. Thus, the addition of thymol to the formulation has little effect on the thermal stability of the material as a whole whereas the addition of TK affects the thermal stability.

Table 2. Thermal parameters obtained from the analyses of the TG profiles of PLA and PLA/ TK composites containing zero, 5% and 10 wt% thymol.

Thymol content / wt%	TK content / wt%	$T_{rel} / ^\circ\text{C}^*$	$T_{deg} / ^\circ\text{C}$	%Char residue at 400°C
0	0	-	352.9	0.5
	10	-	346.4	4.2
	40	-	332.5	11.2
5	0	149.9	354.8	1.4
	10	148.2	341.8	5.0
	40	146.8	328.7	12.1
10	0	147.8	352.3	1.7
	10	146.1	348.3	5.2
	40	144.0	328.1	13.2

* T_{rel} determinations were performed in triplicate

Quantification of Thymol

The effect of adding TK to the formulation on the quantification of thymol released from the matrix can be further explored by plotting the normalized T_{rel} values of PLA/kenaf composites containing 5 and 10 wt% thymol at various TK loadings in the range of zero to 40 wt% (see Figure 6). An almost linear reduction in the T_{rel} with the addition of TK fibre was observed at each of the concentrations of thymol for which the composite containing 40 wt% kenaf showed the lowest T_{rel} (see Figure 6 and Table 2). This suggests that the

presence of the less thermally stable fibres may also destabilize the PLA-thymol matrix. Moreover, no significant change was observed for the T_{rel} of PLA and PLA/TK composite formulations containing 5 and 10 wt% thymol.

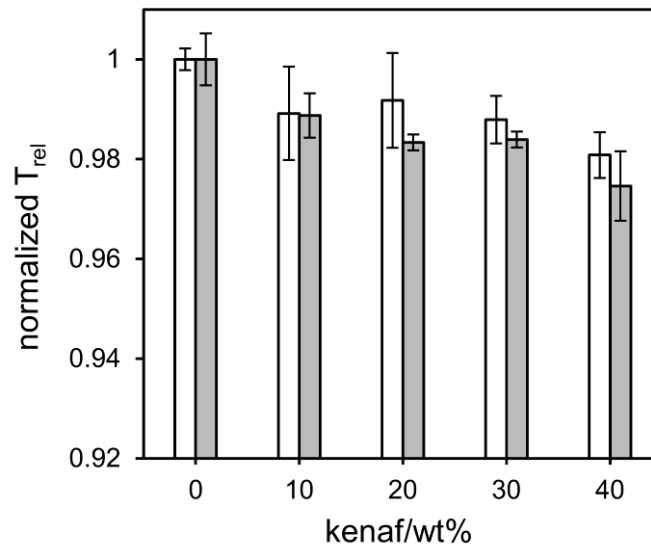


Figure 6 Normalized maximum release rate temperature, T_{rel} , of thymol from PLA and PLA/TK composites containing: 5 wt% (□) and 10 wt% (■) thymol. The determinations were performed in triplicate.

The effect of fillers such as TK on the release of thymol from the polymer matrix has important implications particularly with regard to the loss of the active agent during processing. The TG analysis technique can be used to provide indirect confirmation of the presence of AM agents in the polymer matrix after thermal processing³⁹ and so to this end it was decided to utilize the technique to investigate the effect of the TK filler on the quantification of thymol in the PLA system following thermal processing. The results from the TG experiments were corrected for the inherent water content of the TK fibres and the analytical results were also verified by solid-liquid extraction and GC analysis.¹⁹

Figure 7 shows the weight percentage of thymol that remains in the PLA formulation following thermal processing for neat PLA and PLA/TK composites containing different TK loadings where the analyses were conducted using the TG technique and independently confirmed by extraction followed by GC analysis. There is an acceptable level of consistency between the results obtained using the two techniques. The results suggest that the unfilled PLA formulation exhibits the highest level of thymol (*ca.* 8 wt%) following thermal processing and that the ability of the PLA to retain thymol as such decreases upon increased loadings of the TK filler. Generally, during the thermal mixing process, friction occurs between the barrel and screw that may lead to the degradation of the AM agents.^{40,41} In the current study, it was found that the degradation/release temperature of thymol T_{rel} in active PLA film (see Table 2) was lower (149°C) than the processing temperature (155°C) at which the melt was mixed for 8-10 min. The latter resulted in a considerable loss of thymol from the PLA formulation containing 10 wt% thymol. A similar finding was observed by Ramos *et al.*^{13,20} where *ca.* 75% of the initial thymol remained after processing. Active PLA-based formulations containing butylated hydroxytoluene (BHT) underwent similar losses due to factors such as poor mixing in the extruder, evaporation and thermal degradation of BHT.⁴²

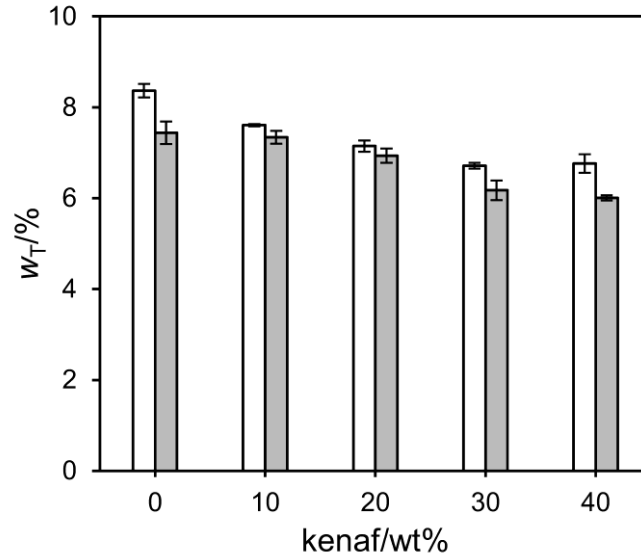


Figure 7 Weight percentage of thymol in the PLA formulation following thermal processing as determined using TGA, w_T , for neat PLA and PLA/TK composites containing different kenaf loadings. Analyses were conducted using: TG analysis (□) and extraction/GC (■). The determinations were performed in triplicate.

The effect of TK loading on the retention of thymol in the formulation can be measured by comparing the residual thymol concentration following processing with the nominal thymol concentration in the formulation. For example, Figure 8 shows plots of w_T , the weight percentage of thymol in the PLA formulation following thermal processing as determined using TG analysis versus w_F , the nominal weight percentage of thymol in the formulation for systems containing two different TK loadings as well as a plot for a control sample (zero TK loading). The PLA composite containing 10 wt% thymol and the higher TK loading (40 wt%) exhibited lower thymol retention compared to the PLA composite containing 10 wt% thymol and 10 wt% TK loading. However, this effect is not as pronounced at the lower thymol concentration of 5 wt%. Overall, it was found that the final weight percentage of thymol in the PLA/TK composites containing 40 wt% TK could be lower than the nominal weight

percentage by up to *ca.* 30%. Furthermore, the results confirm that as the TK loading is increased, the retention of thymol during processing is decreased.

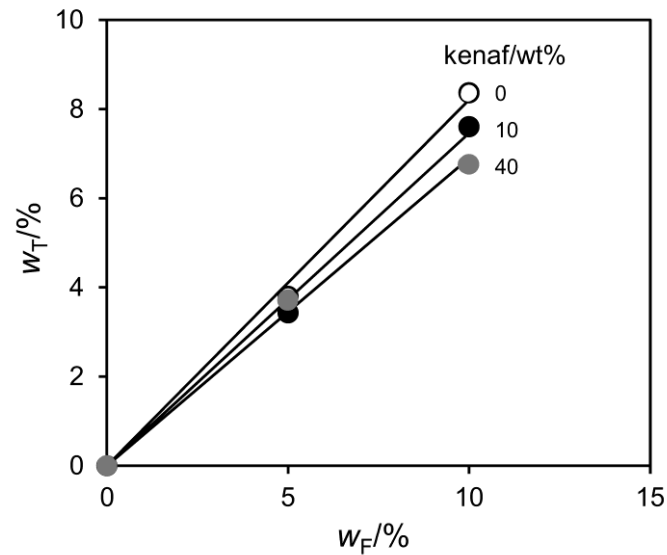


Figure 8 Plots of the weight percentage of thymol in the PLA formulation following thermal processing as determined using TGA, w_T , versus the nominal weight percentage of thymol in the formulation, w_F , for systems containing: zero (○), 10 wt% (●) and 40 wt% (●) TK loading.

Analyses similar to those depicted in Figure 8 were conducted at all loadings of TK used in this study and the gradients of the plots, dw_T/dw_F , along with the corresponding linear coefficient of determination (r^2) are presented in Table 3. In all cases the gradient of the neat PLA (control) is greater than the gradient obtained for the composites and there is a concomitant decrease in the gradient with an increase in the TK loading. This confirms that the presence of TK in these latter systems decreases the retention of thymol during processing with retentions ranging from *ca.* 85% (neat PLA) down to *ca.* 69% (40 wt% TK loading) for a thymol concentration of up to 10 wt%.

Table 3. Linear regression analyses of PLA and PLA/TK formulation containing zero, 5% and 10 wt% thymol.

Composition	dw_T/dw_F	r^2
Neat PLA	0.851	0.993
PLA+10 wt% TK	0.746	0.996
PLA+20 wt% TK	0.724	0.998
PLA+30 wt% TK	0.692	0.990
PLA+40 wt% TK	0.690	0.999

In the case of PLA/TK composites, the increased loss of thymol at higher TK loadings during processing may be attributable to the higher mechanical shear that exists during the mixing of the composites and that most likely contributes to the loss of thymol from the system through evaporation. The presence of a high content of fibre in the PLA-thymol matrix may also lead to the creation of voids that facilitate the release of thymol from the composite. This suggestion is consistent with the SEM images as seen in Figure 9 where voids and loose fibres are observed on the fracture surface of PLA/kenaf composites containing 10 and 30 wt% thymol. Furthermore, the heat evolved during the mixing process might liberate moisture in the TK fibre to produce steam and subsequently facilitate the evaporation of thymol. Indeed, the loss of thymol through a process akin to steam distillation is possible as the boiling point of thymol is lowered in the presence of steam. Mulvaney⁴³ reported that the boiling point of thymol is depressed in the presence of steam, allowing it to evaporate at a temperature below that at which the deterioration of the material becomes appreciable.

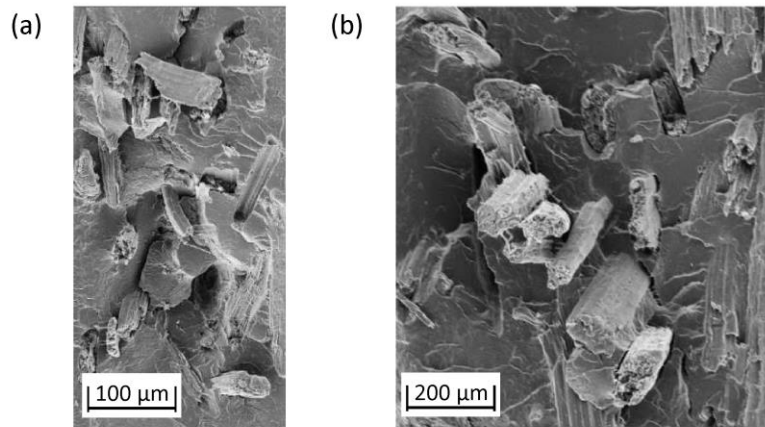


Figure 9 Scanning electron micrographs of: (a) PLA/TK composite containing 20 wt% TK and 10 wt% thymol at 100× magnification and (b) PLA/TK composites containing 20 wt% TK and 30 wt% thymol at 200× magnification.

TG Kinetics Analysis

The apparent activation energy (E_a) for the release of thymol from both active PLA and PLA/TK composites containing 10, 20 or 30 wt% thymol was calculated by applying a 3D-diffusion model (see equations (1) and (3)) based on the TG analysis results (release of thymol, T_{rel} curve). The results are given in Table 4 along with: (i) the corresponding Arrhenius A-factors, (ii) the goodness of fit to the 3D diffusion analysis model as determined on a scale of zero to unity from the computer algorithm, and (iii) the linear regression analyses of the plots of $g(\alpha)$ versus $p(x)$, including the corresponding coefficient of determination, r^2 of the latter. The linearity reflected in the regression analyses demonstrates that the TG fitting protocol is appropriate and provides some degree of confidence in the derived value of the apparent activation energy. Figure 10 shows a typical plot of $g(\alpha)$ versus $p(x)$. In this case the plot pertains to the analysis of the release of thymol from PLA containing 20 wt% TK and 30 wt% thymol. From the gradient of this plot and the

Arrhenius A-Factor, the value of E_a is calculated to be 68 kJ mol^{-1} (see equations (1) and (2)) and this value corresponds closely to the value of 65 kJ mol^{-1} that was delivered by the iterative computer analysis program.

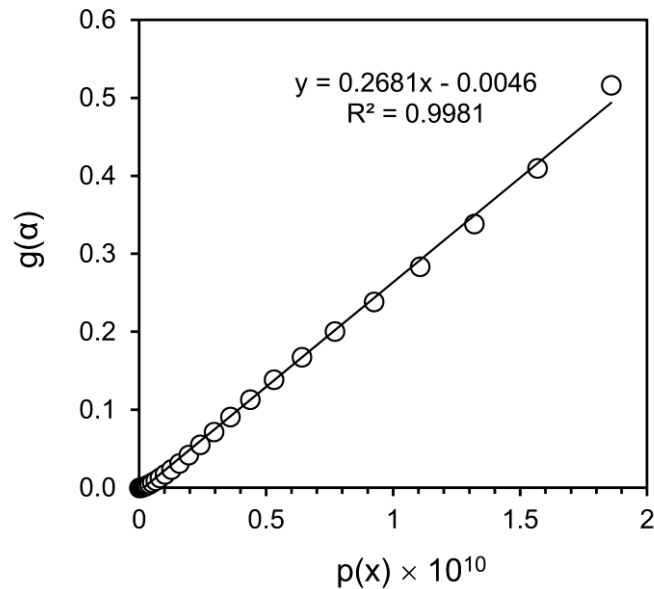


Figure 10 A typical plot of $g(\alpha)$ versus $p(x)$ for PLA/TK composite containing 20 wt% TK and 30 wt% thymol.

The data listed in Table 4 suggest that the apparent activation energy for the release of thymol from PLA is *ca.* 46 kJ mol^{-1} . As expected, this value does not appear to depend on the level of thymol and repeated measurements of this parameter for six replicates at three different thymol concentrations in the range of 10 to 30 wt% thymol yielded an average result of $46 \pm 9 \text{ kJ mol}^{-1}$. Similarly, Soto-Valdez *et al.*⁴⁴ reported that the apparent activation energy of the diffusion of resveratrol (antioxidant) from PLA films immersed in different food simulants and at different resveratrol concentrations remained almost constant between 175 and 177 kJ mol^{-1} . The addition of TK to the formulation significantly increases

the apparent E_a for the release of thymol from the PLA composite matrix, presumably due to the interaction between TK and thymol that was established from the FTIR analysis (see Figure 3 and Table 1). Interestingly, it appears that the level of TK does not significantly affect the value of E_a for thymol release, as there seems only to be a small increase in the latter value when the TK loading is increased from 20 to 40 wt%. Assuming that the E_a for the release of thymol from these systems is independent of the level of thymol and loading of TK over the respective ranges that were tested in the current study, the E_a data for the PLA/TK composites containing thymol can be averaged to produce an overall result of 65 ± 4 kJ mol^{-1} . Clearly, by comparing this result with the average E_a value for the release of thymol from PLA alone demonstrates that the addition of TK to the formulation significantly increases the apparent activation energy for the release of thymol. This may be due to the interaction between the thymol and the kenaf as well as the presence of the filler producing a reduction in amorphous regions through which the additive molecules can be released.^{44,45}

Table 4. Kinetics analyses of TG data for PLA and PLA/ kenaf composites containing thymol.

Formulation	$g(\alpha)$ vs $p(x)$					
	E_a	E_a (ave)*	A/min^{-1}	3D Diff	linear regression analysis	
	$/\text{kJ mol}^{-1}$	$/\text{kJ mol}^{-1}$		Model Fit	regression equation	r^2
PLA+20Th	46	46 ± 9	$6.33\text{E}+03$	0.922	$y = 8.948\text{E}+06x - 2.136\text{E}-02$	0.981
PLA+30Th	44		$1.50\text{E}+03$	0.850	$y = 1.647\text{E}+06x - 3.666\text{E}-03$	0.995
PLA+10Th+30UTK [†]	52	53 ± 6	$3.44\text{E}+04$	0.953	$y = 4.551\text{E}+07x - 4.587\text{E}-03$	0.988
PLA+10Th+20TK	63		$6.26\text{E}+05$	0.919	$y = 6.202\text{E}+08x + 1.991\text{E}-02$	0.961
PLA+20Th+20TK	58	65 ± 4	$2.72\text{E}+05$	0.940	$y = 3.864\text{E}+08x - 6.240\text{E}-03$	0.998
PLA+30Th+20TK	65		$1.64\text{E}+06$	0.905	$y = 2.681\text{E}+09x - 4.579\text{E}-03$	0.998
PLA+10Th+40TK	69		$1.04\text{E}+07$	0.928	$y = 1.487\text{E}+10x + 5.603\text{E}-03$	0.997
UK+25Th [†]	96	98 ± 4	$1.80\text{E}+11$	0.960	$y = 4.577\text{E}+14x + 1.584\text{E}-03$	0.977
TK+25Th [†]	106	105 ± 1	$4.32\text{E}+12$	0.906	$y = 1.583\text{E}+16x - 6.636\text{E}-03$	0.991

* E_a averaged over: 6 different systems for PLA+thymol; 5 replicates for PLA+thymol+UK; 4 different systems for PLA+thymol+TK; 3 replicates for UK+thymol; 2 replicates for TK+thymol

† Fibres doped with thymol

The interaction between the TK and thymol is also confirmed in the apparent activation energy for the release of thymol from TK fibres doped with 25 wt% thymol. The results in Table 4 suggest that the latter E_a value (106 kJ mol^{-1}) is significantly greater than that which is associated with the release of thymol from either the PLA alone or the PLA/thymol/kenaf composites. Whence it can be proposed that the observed increased loss of thymol that

occurs on processing TK containing PLA composites (see Figures 6 to 9) may be due to the presence of the TK which increases friction during processing as well as creates a more open amorphous structure in the resulting material thereby facilitating the release of the thymol. The data in Table 4 are also consistent with the notion that the alkaline chemical pre-treatment of kenaf fibres enhances its compatibility and attractive interactions with substrates such as PLA.^{46,47} In the case of its interaction with thymol it can be seen that the apparent E_a for thymol release from TK (i.e. 106 kJ mol^{-1}) is greater than that for its release from untreated kenaf (UK) where the value of the latter is 96 kJ mol^{-1} . This difference in thymol release from the doped fibres containing 25 wt% thymol is also reflected in the release of thymol from the corresponding PLA composites. The data in Table 4 are consistent in this regard and show that the addition of UK doped with thymol to the PLA composite system lowers as expected the apparent E_a for the release of thymol from the system.

CONCLUSIONS

The FTIR analysis of the active PLA and PLA/TK composites containing thymol showed that the thymol interacts with PLA and kenaf as revealed by the observed significant shifts in the various FTIR absorption bands. Active PLA/KF composites retain less thymol upon processing than PLA alone and the PLA/KF composites containing the highest fibre loadings demonstrated the lowest retained thymol content. This is despite the fact that the apparent activation energy for thymol release from the PLA/TK composites containing thymol being greater than that found for the release of thymol from PLA alone. It would therefore appear that the disruption to the crystalline regions caused by the addition of kenaf, along with the

concomitant creation of voids and the resulting decrease in tortuosity, facilitate the release of the active agent thymol from the composite. These effects seem to overshadow the intermolecular attractions that occur as a result of hydrogen bonding between the components in the composite. Clearly, the interactions between PLA, thymol and kenaf when together in a polymer composite are complex and it is difficult to make any further generalizations based on the data obtained so far. Nonetheless, the exploration of the interactions that exist between the pairs of separate components in these systems can give valuable insight into the mechanism of AM loss during processing and assist in identifying measures that will minimize such losses in future commercial applications.

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REFERENCES

1. Lo Re, G.; Morreale, M.; Scaffaro, R.; La Mantia, F. P. *Polym Int* **2012**, *61*, 1542-1548.
2. Valdés, A.; Mellinas, A. C.; Ramos, M.; Garrigós, M. C.; Jiménez, A. *Polym Chem* **2014**, *2*, 1-10.
3. Li, W. L.; Coffin, D. R.; Jin, T. Z.; Latona, N.; Liu, C. K.; Liu, B.; Zhang, J.; Liu, L. S. *J Appl Polym Sci* **2012**, *126*, 361-373.
4. Kijchavengkul, T.; Auras, R. *Polym Int* **2008**, *57*, 793-804.
5. González, A.; Igarzabal, C. I. A. *Food Hydrocoll* **2013**, *33*, 289-296.
6. Jin, T.; Zhang, H. *J Food Sci* **2008**, *73*, 127-134.
7. Tawakkal, I. S. M. A.; Cran, M. J.; Bigger, S. W. *Ind Crop Prod* **2014**, *61*, 74-83.
8. Kumar, R.; Münstedt, H. *Polym Int* **2005**, *54*, 1180-1186.
9. Martínez-Camacho, A. P.; Cortez-Rocha, M. O.; Castillo-Ortega, M. M.; Burgos-Hernández, A.; Ezquerro-Brauer, J. M.; Plascencia-Jatomea, M. *Polym Int* **2011**, *60*, 1663-1669.
10. Bonilla, J.; Fortunati, E.; Vargas, M.; Chiralt, A.; Kenny, J. M. *J Food Eng* **2013**, *119*, 236-243.
11. Del Nobile, M. A.; Conte, A.; Buonocore, G. G.; Incoronato, A. L.; Massaro, A.; Panza, O. *J Food Eng* **2009**, *93*, 1-6.
12. Han, J. H. In *Novel Food Packaging Techniques*; R. Ahvenainen, Ed.; Woodhead Publishing Ltd., Cambridge, **2003**, Chapter pp 50-65.
13. Ramos, M.; Jiménez, A.; Peltzer, M.; Garrigós, M. C. *J Food Eng* **2012**, *109*, 513-519.
14. Wu, Y.; Yuan, M. W.; Yang, J. Y.; Qin, Y. Y.; Yuan, M. L.; Cao, J. X. *Adv Mater Res* **2014**, *884-885*, 481-484.
15. Guarda, A.; Rubilar, J. F.; Galotto, M. J.; Universidad de Santiago de Chile, Chile . 2012, p 43pp.
16. Tunç, S.; Duman, O. *LWT-Food Sci Technol* **2011**, *44*, 465-472.
17. Tao, F.; Hill, L. E.; Peng, Y.; Gomes, C. L. *LWT-Food Sci Technol* **2014**, *59*, 247-255.
18. Nyambo, C.; Mohanty, A. K.; Misra, M. *Macromol Mater Eng* **2011**, *296*, 710-718.
19. Suppakul, P.; Victoria University of Technology, Melbourne, 2004, p 259.
20. Ramos, M.; Jiménez, A.; Peltzer, M.; Garrigós, M. C. *Food Chem* **2014**, *162*, 149-155.
21. Liu, L. S.; Jin, T. Z.; Coffin, D. R.; Hicks, K. B. *J Agric Food Chem* **2009**, *57*, 8392-8398.
22. Praprudivongs, C.; Sombatsompop, N. *Composites: Part B* **2012**, *43*, 2730-2737.
23. Woraprayote, W.; Kingcha, Y.; Amonphanpokin, P.; Krueate, J.; Zendo, T.; Sonomoto, K.; Benjakul, S.; Visessanguan, W. *Int J Food Microbiol* **2013**, *167*, 229-235.
24. Akil, H. M.; Omar, M. F.; Mazuki, A. A. M.; Safiee, S.; Ishak, Z. A. M.; Abu Bakar, A. *Mater Des* **2011**, *32*, 4107-4121.
25. Saba, N.; Paridah, M. T.; Jawaid, M. *Construction and Building Materials* **2015**, *76*, 87-96.
26. Shukor, F.; Hassan, A.; Saiful Islam, M.; Mokhtar, M.; Hasan, M. *Mater Des* **2014**, *54*, 425-429.
27. Flynn, J. H. In *Thermal Analysis Volume 2: Inorganic Materials and Physical Chemistry*; R. F. Schwenkerand P. D. Garn, Eds.; Elsevier Science, Burlington, **1969**, Chapter pp 1111-1126.
28. Brown, M. E., *Introduction to Thermal Analysis: Vol. 1 Techniques and Applications*, Springer New York, 2001.
29. Dollimore, D.; Evans, T. A.; Lee, Y. F.; Pee, G. P.; Wilburn, F. W. *Thermochim Acta* **1992**, *196*, 255-265.
30. Wang, D. K.; Varanasi, S.; Fredericks, P. M.; Hill, D. J. T.; Symons, A. L.; Whittaker, A. K.; Rasoul, F. *J Polym Sci Part A* **2013**, *51*, 5163-5176.
31. Goncalves, C. M. B.; Coutinho, J. A. P.; Marrucho, I. M. In *Poly(lactic acid) : Synthesis, Structures, Properties, Processing and Applications*; R. A. Auras, L. T. Lim, S. E. M. Selkeand H. Tsuji, Eds.; Wiley, Chichester, **2011**, Chapter pp 97-112.
32. Al-Sheibany, I. S.; Kadhim, K. H.; Abdullah, A. S. *National J Chem* **2005**, *19*, 366-379.

33. Sun, Y.; Cheng, J. *Bioresour Technol* **2002**, *83*, 1-11.
34. Himmelsbach, D. S.; Khalili, S.; Akin, D. E. *J Sci Food Agric* **2002**, *82*, 685-696.
35. Bax, B.; Müssig, J. *Compos Sci Technol* **2008**, *68*, 1601-1607.
36. Garlotta, D. *J Polym Environ* **2001**, *9*, 63-84.
37. Ramos, M.; Fortunati, E.; Peltzer, M.; Dominici, F.; Jiménez, A.; Garrigós, M. D. C.; Kenny, J. M. *Polym Degrad Stab* **2014**,
38. Yussuf, A. A.; Massoumi, I.; Hassan, A. *J Polym Environ* **2010**, *18*, 422-429.
39. Persico, P.; Ambrogi, V.; Carfagna, C.; Cerruti, P.; Ferrocino, I.; Mauriello, G. *Polym Eng Sci* **2009**, *49*, 1447-1455.
40. Soto-Cantú, C. D.; Graciano-Verdugo, A. Z.; Peralta, E.; Islas-Rubio, A. R.; González-Córdova, A.; González-León, A.; Soto-Valdez, H. *J Dairy Sci* **2008**, *91*, 11-19.
41. Graciano-Verdugo, A. Z.; Soto-Valdez, H.; Peralta, E.; Cruz-Zárate, P.; Islas-Rubio, A. R.; Sánchez-Valdes, S.; Sánchez-Escalante, A.; González-Méndez, N.; González-Ríos, H. *Food Res Int* **2010**, *43*, 1073-1078.
42. Ortiz-Vazquez, H.; Shin, J.; Soto-Valdez, H.; Auras, R. *Polym Test* **2011**, *30*, 463-471.
43. Mulvaney, J. *Aus J Herbal Med* **2012**, *24*, 140-142.
44. Soto - Valdez, H.; Auras, R.; Peralta, E. *J Appl Polym Sci* **2011**, *121*, 970-978.
45. Limm, W.; Hollifield, H. C. *Food Add Contam* **1996**, *13*, 949-967.
46. Huda, M. S.; Drzal, L. T.; Mohanty, A. K.; Misra, M. *Compos Sci Technol* **2008**, *68*, 424-432.
47. Ibrahim, N. A.; Yunus, W. M. Z. W.; Othman, M.; Abdan, K. *J Reinf Plast Compos* **2011**, *30*, 381-388.