



VICTORIA UNIVERSITY
MELBOURNE AUSTRALIA

Muscle oxygenation maintained during repeated-sprints despite inspiratory muscle loading

This is the Published version of the following publication

Rodriguez, Ramon F, Townsend, NE, Aughey, Robert and Billaut, Francois
(2019) Muscle oxygenation maintained during repeated-sprints despite
inspiratory muscle loading. PLoS ONE, 14 (9). ISSN 1932-6203

The publisher's official version can be found at
<https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0222487>
Note that access to this version may require subscription.

Downloaded from VU Research Repository <https://vuir.vu.edu.au/39773/>

RESEARCH ARTICLE

Muscle oxygenation maintained during repeated-sprints despite inspiratory muscle loading

Ramón F. Rodríguez¹, Nathan E. Townsend², Robert J. Aughey¹, François Billaut^{1,3*}

1 Institute for Health and Sport, Victoria University, Melbourne, Australia, **2** Aspetar, Doha, Qatar, **3** Department of kinesiology, University Laval, Quebec, Canada

☯ These authors contributed equally to this work.

* francois.billaut@kin.ulaval.ca



OPEN ACCESS

Citation: Rodríguez RF, Townsend NE, Aughey RJ, Billaut F (2019) Muscle oxygenation maintained during repeated-sprints despite inspiratory muscle loading. *PLoS ONE* 14(9): e0222487. <https://doi.org/10.1371/journal.pone.0222487>

Editor: Hugo A. Kerhervé, Univ Rennes, FRANCE

Received: August 9, 2018

Accepted: August 30, 2019

Published: September 19, 2019

Copyright: © 2019 Rodríguez et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: The data underlying this study have been uploaded to the Open Science Framework and are accessible using the following link: <https://osf.io/3ft2q/>.

Funding: Funding was provided by the Graduate Research Office (PhD Student Budget) at Victoria University. Funding was also provided in the form of a salary to NET by his employer, Aspetar Orthopaedic and Sports Medicine Hospital. The specific roles of this author are articulated in the 'author contributions' section. The funder had no role in study design, data collection and analysis,

Abstract

A high work of breathing can compromise limb oxygen delivery during sustained high-intensity exercise. However, it is unclear if the same is true for intermittent sprint exercise. This project examined the effect of adding an inspiratory load on locomotor muscle tissue reoxygenation during repeated-sprint exercise. Ten healthy males completed three experiment sessions of ten 10-s sprints, separated by 30-s of passive rest on a cycle ergometer. The first two sessions were 'all-out' efforts performed without (CTRL) or with inspiratory loading (INSP) in a randomised and counterbalanced order. The third experiment session (MATCH) consisted of ten 10-s work-matched intervals. Tissue saturation index (TSI) and deoxy-haemoglobin (HHb) of the vastus lateralis and sixth intercostal space was monitored with near-infrared spectroscopy. Vastus lateralis reoxygenation (ΔReoxy) was calculated as the difference from peak HHb (sprint) to nadir HHb (recovery). Total mechanical work completed was similar between INSP and CTRL (effect size: -0.18 , 90% confidence limit ± 0.43), and differences in vastus lateralis TSI during the sprint (-0.01 ± 0.33) and recovery (-0.08 ± 0.50) phases were unclear. There was also no meaningful difference in ΔReoxy (0.21 ± 0.37). Intercostal HHb was higher in the INSP session compared to CTRL (0.42 ± 0.34), whilst the difference was unclear for TSI (-0.01 ± 0.33). During MATCH exercise, differences in vastus lateralis TSI were unclear compared to INSP for both sprint (0.10 ± 0.30) and recovery (-0.09 ± 0.48) phases, and there was no meaningful difference in ΔReoxy (-0.25 ± 0.55). Intercostal TSI was higher during MATCH compared to INSP (0.95 ± 0.53), whereas HHb was lower (-1.09 ± 0.33). The lack of difference in ΔReoxy between INSP and CTRL suggests that for intermittent sprint exercise, the metabolic O_2 demands of both the respiratory and locomotor muscles can be met. Additionally, the similarity of the MATCH suggests that ΔReoxy was maximal in all exercise conditions.

decision to publish, or preparation of the manuscript.

Competing interests: The authors have read the journal's policy and have the following competing interests: NET is a paid employee of Aspetar Orthopaedic and Sports Medicine Hospital. This does not affect our adherence to PLOS ONE policies on sharing data and materials. There are no patents, products in development, or marketed products to declare. This does not alter our adherence to PLOS ONE policies on sharing data and materials.

Introduction

Repeated-sprint exercise is characterised by brief periods of “maximal” exertion, interspersed with incomplete recovery periods. Over the course of a repeated-sprint series, there is a progressive reduction in both peak and mean power output, with a plateau in the latter sprints [1–4]. While phosphocreatine (PCr) hydrolysis and anaerobic glycolysis are heavily relied on as a rapid source of adenosine triphosphate (ATP) replenishment in sprint exercise [2, 5], the aerobic system plays an increasingly significant role in maintaining performance when sprints are repeated. In fact, PCr resynthesis and removal of inorganic phosphate are exclusively performed through oxidative processes [6], and sensitive to oxygen (O_2) availability [7]. Maintaining O_2 delivery to locomotor muscles during repeated-sprint exercise is therefore an important mediating factor of performance.

Near-infrared spectroscopy (NIRS) offers the possibility to explore O_2 balance (delivery vs. consumption) in skeletal muscle during sprint activity in real-time. Deoxy-haemoglobin ([HHb]) and oxy-haemoglobin ($[O_2Hb]$) rise and fall, respectively, proportional to an increase in metabolic activity in the underlying tissue. Relative changes in [HHb] have been primarily examined during repeated-sprint exercise, because this variable is considered to be independent of blood volume [8, 9], and is assumed to provide a reliable estimate of muscle O_2 extraction [9, 10]. At sprint onset, there is a rapid increase in vastus lateralis [HHb], which recovers during rest periods [1, 11, 12]. Muscle reoxygenation between sprints may also describe the quality of metabolic recovery [13]. Improving this process typically increases repeated-sprint performance [14, 15], whereas a slower reoxygenation is associated with performance impairments [11, 13]. However, it is currently unclear if the O_2 cost of exercise hyperpnoea has any influence on locomotor muscle oxygenation trends during repeated-sprint exercise.

The respiratory muscles demand ≈ 10 –15% of total pulmonary O_2 uptake ($\dot{V}O_2$) during high-intensity exercise, as well as a considerable portion of cardiac output to maintain adequate O_2 delivery [16]. An elevated work of breathing during high-intensity exercise promotes competition between locomotor and respiratory muscles for available cardiac output [17]. In fact, the addition of an inspiratory load to artificially increase the work of breathing during severe exercise ($>95\% \dot{V}O_{2peak}$) limits endurance capacity via decreased limb perfusion and O_2 delivery that is mediated by a sympathetically activated vasoconstriction in the locomotor muscles [18, 19]. However, at moderate intensities (50 – $75\% \dot{V}O_{2peak}$) there is no change in vascular resistance or blood flow to the locomotor muscles [20], suggesting that exercise intensity is an important mediator of locomotor vasoconstriction when the work of breathing is high.

It is currently unclear if an elevated work of breathing influences reoxygenation capacity during repeated-sprint exercise. Therefore, we aimed to determine the impact of an elevated work of breathing on $\dot{V}O_2$, tissue oxygenation trends and mechanical output during repeated-sprint exercise. We believe that, by increasing the work of breathing, there will be no concurrent increase in $\dot{V}O_2$, but that vastus lateralis oxygenation will be compromised and repeated-sprint ability will be impaired.

Materials and methods

Subjects

Ten males from a variety of athletic backgrounds (team sports, road cycling, combat sports, CrossFit) were recruited for this study (age 25.5 ± 3.6 years; height 184.00 ± 7.69 cm; body mass 81.45 ± 8.29 Kg; $\dot{V}O_{2peak}$ 4.40 ± 0.36 L·min⁻¹, 54.4 ± 5.9 mL·min⁻¹·Kg⁻¹; \dot{V}_{Epeak} 173.6 ± 26.9 L·min⁻¹). Subject pulmonary function data is presented in Table 1. These subjects

Table 1. Pulmonary function data.

Measure	Raw	%Predicted
FVC (L)	6.0 ± 0.7	103 ± 8.8
FEV ₁ (L)	4.9 ± 0.5	99.7 ± 8.1
FVC/FEV ₁	79.1 ± 6.6	95.5 ± 7.5
MVV (L·min ⁻¹)	212.9 ± 24.7	110.9 ± 10.2
MIP (cmH ₂ O)	-146.6 ± 19.2	
MEP (cmH ₂ O)	151.8 ± 35.8	

Abbreviations are: forced vital capacity, FVC; forced expired volume in 1 s, FEV₁; maximum voluntary ventilation, MVV; maximum inspiratory pressure, MIP; maximum expiratory pressure, MEP. Values are reported as mean ± SD.

<https://doi.org/10.1371/journal.pone.0222487.t001>

were chosen because they were accustomed to producing “all-out” bouts of exercise. Subjects self-reported in a written questionnaire to be healthy non-smokers and with no known neurological, cardiovascular, respiratory diseases, or any other medical conditions. If subjects indicated “yes” to any of the contraindications, they were excluded from participation. After being fully informed of the requirements, benefits, and risks associated with participation, each subject gave written informed consent. Ethical approval for the study was obtained from the Victoria University Human Research Ethics Committee and the study conformed to the declaration of Helsinki. Prior to each experiment session, subjects were asked to replicate their preceding meal and to refrain from caffeinated beverages for 24 h. Subjects were also asked to refrain from any strenuous exercise for 48 h prior to every session.

Experiment design

Subjects visited the laboratory on six occasions. During visit one, subjects completed pulmonary function (spirometry and maximum voluntary ventilation) (Ultima CPX, MGC Diagnostics, Minnesota, United States of America) and respiratory muscle strength tests (MicroRPM, Micro Medical, Hoechst, Germany) followed by a maximal ramp exercise test familiarisation. In visit two, a ramp exercise test to volitional exhaustion was completed. During visit three, subjects completed a familiarisation session consisting of the same repeated-sprint protocol used in the experiment sessions. In visits four and five subjects completed the actual repeated-sprint test in a randomised, counterbalanced, cross-over design with either no restriction to their breathing (CTRL) or with inspiratory loading (INSP). During the sixth laboratory visit, ten work-matched intervals to that of the INSP experiment session were completed with normal breathing (MATCH). The MATCH condition was included in the experiment design to firstly minimise the physiological disturbances that are typically associated with “maximal” sprint performance [2, 3, 5], and to secondly allow for comparison of the data to an exercise trial (INSP) where the same amount of mechanical work was performed by the subjects. All exercise testing was performed on an electronically-braked cycle ergometer (Excalibur, Lode, Groningen, The Netherlands). Experiment sessions were conducted at the same time of day and separated by 3–7 days.

Maximal ramp exercise testing

A maximal ramp cycling ergometer test was performed to determine $\dot{V}O_{2peak}$. The exercise test was initiated at a work rate of 0 W for 3 min. This was followed by an increase in work rate of 1 W every 2 s (30 W·min⁻¹) until volitional exhaustion or until cadence fell 10 rpm below self-selected rate. Expired gases were collected on a breath-by-breath basis (COSMED Quark

CPET; Cosmed, Rome, Italy), and peak $\dot{V}O_2$ was determined as the highest 30 s average prior to exercise termination ($\dot{V}O_{2\text{peak}}$). The corresponding \dot{V}_E at $\dot{V}O_{2\text{peak}}$ was deemed to be $\dot{V}_{E\text{peak}}$. Subjects were familiarised with this protocol during their first visit to the laboratory. Subjects completed the same protocol to volitional exhaustion on visit two.

Repeated-sprint exercise

After arriving at the laboratory, subjects were fitted with NIRS probes and a heart rate monitor. Testing was performed with the cycle ergometer set to isokinetic mode. In this mode, a variable resistance is applied to the flywheel proportional to the torque produced by the subjects to constrain their pedalling rate to 120 rpm. The handlebars and seat were individually adjusted to each subjects' characteristics and feet secured using toe cages and retention straps fitted to the ergometer. Crank arm length was standardised to 175 mm. After a 7-min warm-up consisting of 5 min of unloaded cycling at 60–70 rpm and two 4-s sprints (separated by 1 min), subjects rested for another 2.5 min before the repeated-sprint protocol was initiated. The repeated-sprint protocol consisted of ten consecutive 10-s sprints separated by 30 s of passive rest [4, 21–23]. Subjects were instructed to give an “all-out” effort for every sprint and verbally encouraged throughout to promote a maximal effort. Each sprint was performed in the seated position and initiated with the crank arm of the dominant leg at 45°. Before sprint one, subjects were instructed to accelerate the flywheel to 95 rpm over a 15-s period and assume the ready position 5 s before the commencement of the test. This ensured that each sprint was initiated with the flywheel rotating at ≈ 90 rpm so that subjects could quickly reach 120 rpm. Visual feedback of power output was not available to the subjects during any sprint. The cycle ergometer software provides power and cadence at 4 Hz. Data were exported to Microsoft Excel for analysis. Peak power output was determined by identifying the highest individual power value (watts), which happened to be during Sprint 1 in every case. Mean power output was calculated at the average power within each of the ten sprints. Mechanical work performed (J) was calculated by integrating the power curve over every 10-s sprint. Ratings of perceived exertion for exercise ($\text{RPE}_{\text{Exercise}}$) and breathing ($\text{RPE}_{\text{Breath}}$) were recorded following sprint one, five and ten, using a Borg 6–20 RPE scale. In every case, subjects were asked the questions “how difficult is the exercise?” and “how difficult is the breathing?”. The scale was anchored so that 6 represented no exertion, and 20 represented maximal exertion.

Inspiratory loading was achieved by placing a plastic disk with a 10-mm opening over the inspiratory side of a two-way non-rebreathing valve (Hans Rudolph inc., Kansas, United States of America) attached to the distal end of the breath-by-breath gas sampling line and turbine [24, 25]. The non-rebreathing valve was worn during all the repeated-sprint experiment sessions. The inspiratory load was applied after warm-up, 1 min before the commencement of the repeated-sprint protocol. Work-matched (MATCH) exercise was conducted by performing ten 10-s bouts of exercise separated by 30 s of passive rest matched for total work performed during the INSP experiment session. This was achieved by programming the cycle ergometer for ten bouts of fixed work rate intervals equivalent to each subject's calculated mean power output for every corresponding sprint repetition (1 to 10) in the INSP experiment session. Between each sprint interval, the cycle ergometer was programmed to 0 watts. The cycle ergometers isokinetic function cannot be used when controlling for power output, therefore, subjects were asked to maintain cadence at 120 rpm during each interval. Warm-up procedures were identical to the CTRL and INSP experiment sessions.

Participants were familiarised with the repeated-sprint protocol during their third visit to the laboratory. Subjects were only instrumented with a silicone facemask to which the gas sampling line and turbine would be attached in later trials, and non-rebreathing valve. To expose

subjects to the inspiratory loading, they performed the first two sprints of the series with restricted inhalation. The inspiratory loading was promptly removed, and the subjects continued with the remaining eight sprints. Subjects were also asked to rate their perceived exertion for exercise and breathing during this familiarisation trial.

Metabolic and ventilatory measurements

The same metabolic system that was used to determine $\dot{V}O_{2peak}$ was used during the repeated-sprint experiment sessions to collect breath-by-breath data. The analyser was calibrated before each test against known gas concentrations and the turbine volume transducer was calibrated using a 3 L syringe (Cosmed, Rome, Italy). Errant data points due to coughing or swallowing were initially removed and thereafter any breath further than 4 standard deviations from the local mean was removed [26, 27]. A 5-breath rolling average was applied for the calculation of peak and nadir for both $\dot{V}O_2$ and \dot{V}_E for every 40-s sprint/recovery period to give a single value for each sprint and recovery phase. Inspiratory volume (IV), respiratory frequency (f_R), end-tidal O_2 partial pressure ($P_{ET}O_2$), and end-tidal CO_2 partial pressure ($P_{ET}CO_2$) were averaged to give one value for each 40-s period. Because the facemask was removed immediately after the tenth sprint, only maximum values were calculated over the first 10-s. Mouth pressure (P_m) was recorded continuously at 50 Hz with a pressure transducer (Honeywell, New Jersey, United States of America) attached to the saliva port of the non-rebreathing valve via Tygon tubing. Representative data of the effects of inspiratory muscle loading on P_m from one subject is displayed in Fig 1. Mean inspiratory and expiratory P_m was then calculated as an index of respiratory muscle work. To account for any change in f_R , an index of inspiratory muscle force development was also calculated for each exercise experiment session ($\int P_m \times f_R$, expressed in arbitrary units) [28]. For statistical analysis, inspiratory P_m was converted to positive values and presented in the results as such. Heart rate (HR) (Cosmed, Rome, Italy) and arterial oxygen saturation (estimated by fingertip pulse oximetry; S_pO_2) (Nonin Medical, Minnesota, United States of America) was collected on a breath-by-breath basis integrated into the Cosmed system.

Near-infrared spectroscopy

Subjects were instrumented with two NIRS probes to assess muscle oxygenation (Oxymon MKIII, Artinis, the Netherland). The first probe was fixed over the distal part of the vastus

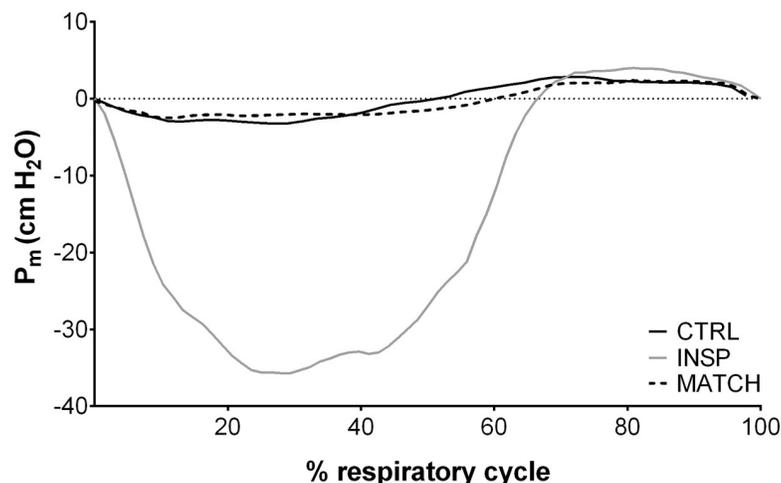


Fig 1. Representative data of the effects of inspiratory muscle loading (INSP) on mouth pressure (P_m) compared to control (CTRL) and work matched (MATCH) exercise conditions.

<https://doi.org/10.1371/journal.pone.0222487.g001>

lateralis muscle belly approximately 15 cm above the proximal border of the patella of the dominant leg. The second probe was fixed over the left 6th intercostal space at the anterior axillary line of the serratus anterior to assess changes in the accessory respiratory muscles. Probes were held in place with black plastic spacers secured to the skin using double-sided tape and shielded from light using a black self-adhesive elastic bandage. An indelible marker was used to trace the position of the probes to ensure placement was reproduced in subsequent visits. Optode spacing was set to 4.5 cm and 3.5 cm for vastus lateralis and respiratory muscles, respectively. Skinfold thickness was measured between the emitter and detector using a skinfold calliper (Harpenden Ltd.) to account for skin and adipose tissue thickness covering the muscle. The skinfold thickness for vastus lateralis (1.19 ± 0.69 cm) and respiratory muscles (1.12 ± 0.44 cm) was less than half the distance between the emitter and the detector in every case. A modified form of the Beer-Lambert law was used to calculate micromolar changes in tissue [HHb] and [O₂Hb] across time using the received optical density from continuous wavelengths of NIR light. A differential pathlength factor of 4.95 was used [12]. We chose to focus our analysis on Δ [HHb] to allow comparisons to previous research; because Δ [HHb] is independent of changes in total haemoglobin [8]; and taken to reflect venous [HHb] which provides an estimate of muscular oxygen extraction [9, 10]; and because Δ [O₂Hb] is influenced by rapid blood volume and perfusion variations caused by forceful muscle contractions [8, 29]. To assess muscle oxygenation status, we used spatially-resolved spectroscopy to calculate a Tissue Saturation Index (TSI). The Artinis system uses three light sources to calculate a correction factor which is used to determine [O₂Hb] and [HHb] [30–32]. Then, the ratio of [O₂Hb] to total haemoglobin is calculated ($TSI = [O_2Hb] / ([O_2Hb] + [HHb])$, expressed in %). This method robustly reflects the dynamic balance between O₂ supply and O₂ consumption in the tissue microcirculation beyond what [O₂Hb] alone can provide, and is independent of the coupling between the optodes and tissue [31]. In fact, spatially-resolved spectroscopy methods are less susceptible to be influenced by underlying adipose tissue thickness than modified Beer-Lambert law methods [33]. Moreover, skin blood flow does not appear to affect spatially-resolved spectroscopy derived variables (TSI), whereas modified Beer-Lambert law derived variables (O₂Hb and HHb) appear to be more vulnerable [34]. Our analysis of respiratory muscle TSI was limited to $n = 8$ due to partial data loss in one of the transmitter fibre optic cables for two subjects. An example of the raw vastus lateralis NIRS data from a single subject is presented in Fig 2, and an example of respiratory muscle responses can be found in our previous work [22].

Data were acquired at 10 Hz. A 10th order zero-lag low-pass Butterworth filter was applied to the data to remove artefacts and smooth pedalling induced fluctuations; the resulting output was used for analysis [21]. The application of the filter was conducted in the R environment (R: A language and environment for statistical computing, Vienna, Austria). Values were then normalised to femoral artery occlusion so that 0% represented a 5-s average immediately prior to the occlusion and 100% represented the maximum 5 s average. Occlusion was performed 3–5 min following the cessation of the sprints while the subjects were supine on an examination bed. Commencement of occlusion was largely influenced by the subject's wellness following the sprint protocol (e.g., exercise-related pre-syncope or nausea were present in two subjects after INSP). Subjects were asked to place their feet flat on the examination bed with $\approx 90^\circ$ of knee flexion. A pneumatic tourniquet (Rudolf Riester GmbH, Jungingen, Germany) was positioned as high as possible around the thigh and inflated to 350 mmHg. The tourniquet remained in place until there was a plateau of at least 10-s in vastus lateralis HHb, which took approximately 5–7 min [9]. Tourniquet pressure was monitored continuously to ensure it remained at 350 mmHg for the duration of the occlusion period. To obtain one value per sprint and recovery for vastus lateralis HHb (HHb_{VL}) and TSI (TSI_{VL}), peaks and nadirs were

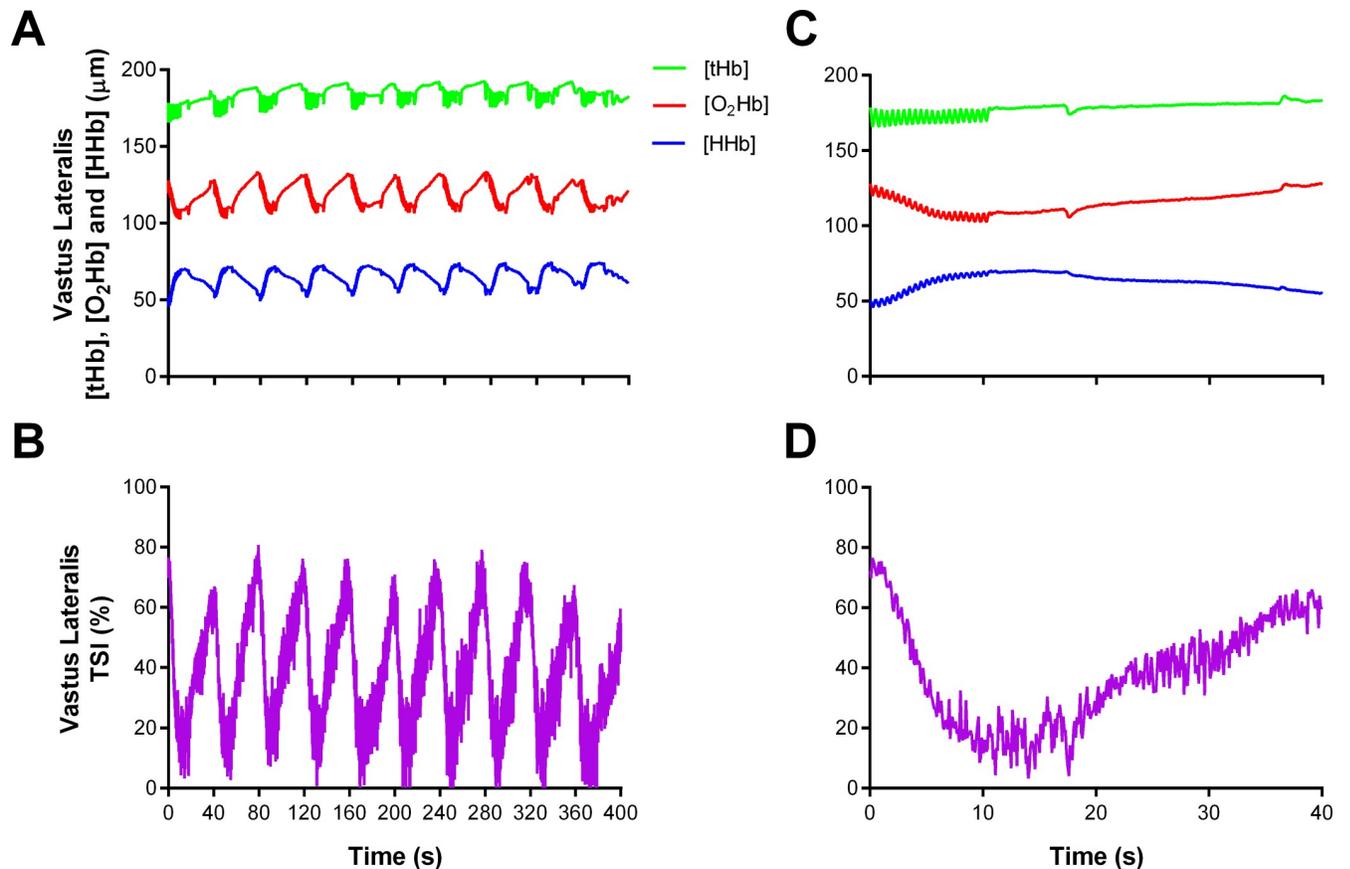


Fig 2. Representative data of the raw NIRS responses to repeated-sprint exercise. (A) Concentrations of vastus lateralis total haemoglobin ([tHb]), oxyhaemoglobin ([O₂Hb]), and deoxyhaemoglobin ([HHb]), and (B) vastus lateralis Tissue Saturation Index (TSI) for the entire repeated-sprint protocol. (C) [tHb], [O₂Hb], and [HHb], and (D) TSI for the first 10-s sprint and the following 30-s of passive rest.

<https://doi.org/10.1371/journal.pone.0222487.g002>

identified for each period using a rolling approach [21]. Time to peak HHb_{VL} (TTP_{HHb}) was also calculated as the time from sprint onset to peak HHb. Reoxygenation capacity (Δ Reoxy, %) and reoxygenation rate (Reoxy rate, %·s⁻¹) were also calculated as the change from sprint to recovery. The baseline for respiratory muscle HHb was established before warm-up while seated quietly on the cycle. Respiratory muscle HHb (HHb_{RM}) values are expressed as per cent change from baseline. Because there were no clear peaks and nadirs in the respiratory muscle variables (HHb_{RM} and TSI_{RM}), averages were calculated for each 40-s sprint/recovery period.

Statistical analysis

Data in text and figures are presented as mean \pm standard deviation. A custom made spreadsheet was used to analyse the effects of INSP and MATCH on laboratory measurements [35]. All measures, other than $\dot{V}O_2$, S_pO₂, RPE and NIRS responses (except for TTP_{HHb}), were log-transformed before analysis then back-transformed to express the changes in per cent units and standardised effects. Relative changes (%) and standardised effects are expressed with 90% confidence limits (90% CL). Practical significance was assessed by calculating Cohen's d effect size (ES) [36]. Standardised effect sizes of <0.2, 0.2–0.5, 0.5–0.8, >0.8 were considered as trivial, small, moderate and large respectively and presented with 90% CL. Probabilities were also calculated to establish if the chance the true (unknown) differences were lower, similar to or

higher than the smallest worthwhile change (ES = 0.2). Effects were not considered meaningful if there was <75% probability of being substantially positive/negative relative to the smallest worthwhile change. If the chance of having higher/lower values than the smallest worthwhile difference was both >5%, the true difference was assessed as *unclear*. For clear effects, the likelihood that the true effect was substantial were assessed qualitatively as follows: *likely* (75 to <95%), *very likely* (95 to <99%), *almost certainly* (>99%) [37].

Results

Mouth pressure

Mouth pressure responses to exercise and inspiratory muscle loading are presented in Table 2, and representative data from a single subject is presented in Fig 1. Mean inspiratory P_m was greater during INSP compared to CTRL (relative difference = 616.6%, 90% CL ±62.2%; ES 13.33, 90% CL ±0.59). Similarly, inspiratory muscle force generation ($\int P_m \times f_R$) was *almost certainly* higher during INSP compared to CTRL (753.6% ±91.1%; ES 13.04 ±0.65). But there was an *unclear* difference in mean expiratory P_m (-0.6% ±7.1%; ES -0.05 ±0.61).

Table 2. Physiological and perceptual responses to repeated-sprint exercise during control (CTRL), inspiratory loading (INSP), and work match (MATCH) conditions.

Variable	CTRL	INSP	MATCH
Inspiratory P _m (cm H ₂ O)	2.4 ± 0.3	17.6 ± 3.6***	1.6 ± 0.3###
$\int P_m \times f_R$ (AU)	76 ± 11	658 ± 143***	45 ± 9###
Expiratory P _m (cm H ₂ O)	2.1 ± 0.2	2.1 ± 0.2	1.2 ± 0.2###
Sprint $\dot{V}O_2$ (% $\dot{V}O_{2peak}$)	90.7 ± 6.7	95.4 ± 4.3**	73.7 ± 6.1###
Recovery $\dot{V}O_2$ (% $\dot{V}O_{2peak}$)	70.7 ± 5.2	74.9 ± 4.5*	58.6 ± 4.3###
Sprint \dot{V}_E (L·min ⁻¹)	161.5 ± 27.5	129.1 ± 16.8***	83.4 ± 15.2###
Recovery \dot{V}_E (L·min ⁻¹)	101.4 ± 13.9	89.7 ± 11.4**	60.2 ± 10.0###
IV (L)	3.0 ± 0.5	3.1 ± 0.5	2.6 ± 0.4###
f _R (b·min ⁻¹)	48.1 ± 7.8	37.8 ± 5.0***	30.2 ± 6.0##
P _{ET} O ₂ (mmHg)	117.7 ± 2.3	113.4 ± 2.8***	105 ± 5.1###
P _{ET} CO ₂ (mmHg)	35.5 ± 2.3	38.6 ± 2.6***	41.6 ± 3.2##
S _p O ₂ (%)	97 ± 1	97 ± 1	97 ± 1
HR (b·min ⁻¹)	153 ± 12	154 ± 9	131 ± 12###
RPE _{Exercise} (AU)			
Sprint 1	15 ± 3	14 ± 2	12 ± 2###
Sprint 5	17 ± 2	17 ± 2	13 ± 2###
Sprint 10	18 ± 2	18 ± 2	13 ± 2###
RPE _{Breath} (AU)			
Sprint 1	12 ± 2	15 ± 2***	11 ± 2###
Sprint 5	15 ± 1	18 ± 2***	12 ± 2###
Sprint 10	16 ± 2	19 ± 1***	12 ± 2###

Abbreviations are: mouth pressure, P_m; inspiratory muscle force development, $\int P_m \times f_R$; pulmonary oxygen uptake, $\dot{V}O_2$; pulmonary ventilation, \dot{V}_E ; inspiratory volume, \dot{V}_E ; inspiratory volume, IV; respiratory frequency, f_R; end-tidal oxygen partial pressure, P_{ET}O₂; end-tidal carbon dioxide partial pressure, P_{ET}CO₂; arterial oxygen saturation estimated by pulse oximetry, S_pO₂; heart rate, HR; rating of perceived exertion for exercise, RPE_{Exercise}; rating of perceived exertion for breathing, RPE_{Breath}. The symbols represent comparisons between INSP and CTRL (*), INSP and MATCH (#). The number of symbols; one, two and three denote likely, very likely and almost certainly, respectively, that the chance of the true effect exceeds a small (-0.2–0.2) effect size. Values are reported as mean ± SD.

<https://doi.org/10.1371/journal.pone.0222487.t002>

Mean inspiratory P_m was lower during MATCH compared to INSP with a large effect ($-91.0\% \pm 0.9$; ES -10.18 ± 0.42). Likewise, $\int P_m \times f_R$ was lower during the MATCH experiment sessions ($-93.1\% \pm 1.0\%$; ES -9.92 ± 0.52). A large effect was also present for expiratory P_m with MATCH being lower than INSP ($-40.3\% \pm 5.8\%$; ES -4.14 ± 0.78).

Mechanical measurements

Total work completed on the cycle ergometer per sprint and over the entire protocol for each condition is presented in Fig 3. There was no meaningful effect of INSP on total work completed over the entire repeated-sprint protocol compared to CTRL ($-2.7\% \pm 6.4\%$; ES -0.17 ± 0.42). Similarly and as expected, total work performed during MATCH and INSP trials ($-0.6\% \pm 0.1\%$; ES -0.04 ± 0.01) was not different. There was no meaningful effect of INSP on peak power output (1097 ± 148 W) compared to CTRL (1158 ± 172 W) ($-5.1\% \pm 6.1$; ES $-0.30, \pm 0.35$). However, there was an *almost certainly* large effect between MATCH (773 ± 122 W) and INSP ($-29.7\%, \pm 2.3\%$; ES -2.21 ± 0.20).

Physiological responses

Responses to exercise are presented in Table 2 and Fig 4. Over the entire protocol, $\dot{V}O_2$ was greater during both sprint ($4.7\% \pm 2.7\%$; ES 0.64 ± 0.37) and recovery ($4.2\% \pm 3.1$; ES 0.74 ± 0.55) for INSP compared to CON. Additionally, $\dot{V}O_2$ was lower during MATCH compared to INSP during both sprint ($-21.7\% \pm 5.0\%$; ES -4.59 ± 1.06) and recovery ($16.3\% \pm 2.9\%$; ES -3.30 ± 0.58). Likewise, \dot{V}_E during INSP was lower both during sprint ($-19.6\% \pm 3.5\%$; ES -1.13 ± 0.22) and recovery ($-11.5\% \pm 5.6$; ES -0.80 ± 0.41) compared to CTRL. Throughout MATCH, \dot{V}_E was lower during both sprint ($-35.8\% \pm 9.5\%$; ES -2.92 ± 0.98) and recovery ($-33.2\% \pm 6.2\%$; ES -2.81 ± 0.65). There was no meaningful difference of IV between INSP and CTRL ($2.8\% \pm 4.8\%$; ES 0.16 ± 0.27). On the other hand, IV was *almost certainly* lower during MATCH compared to INSP ($-15.6\% \pm 5.1\%$; ES -0.91 ± 0.32). There was an *almost certainly* large effect of INSP on f_R compared to CTRL ($-21.2\% \pm 4.7\%$; ES -1.41 ± 0.35). Additionally, f_R was *very likely* lower during MATCH compared to INSP ($-20.8\% \pm 9.6\%$; ES -1.53 ± 0.79).

During INSP, $P_{ET}O_2$ was lower than CTRL ($-3.7\% \pm 2.3\%$; ES -1.71 ± 0.65), and lower during MATCH compared to CTRL ($-7.4\% \pm 2.3$; ES -2.81 ± 0.91). Conversely, $P_{ET}CO_2$ was higher

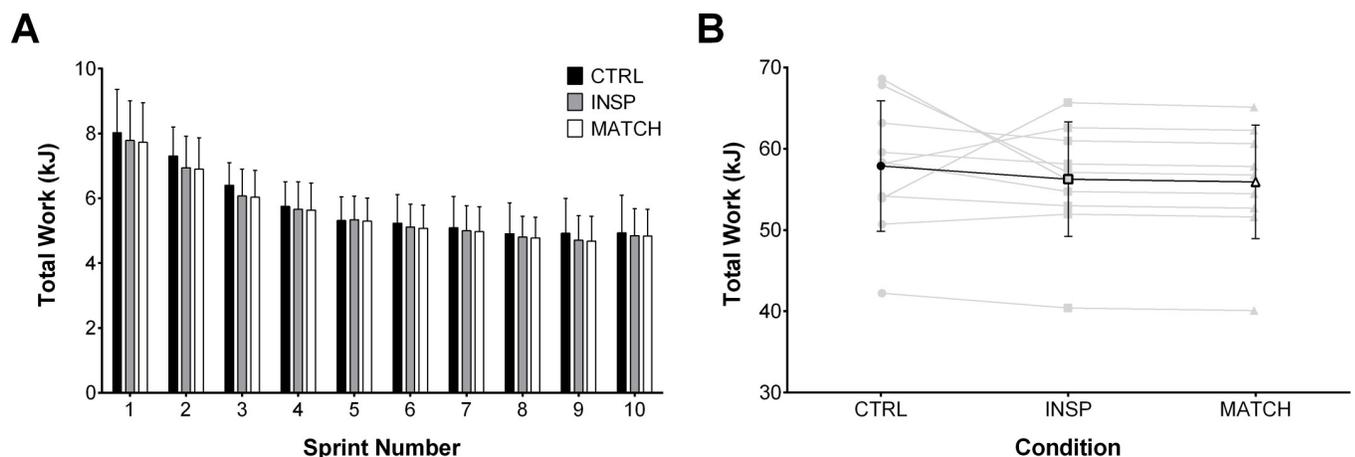


Fig 3. Total mechanical work performed during control (CTRL), inspiratory loading (INSP), and work match (MATCH) exercise conditions. (A) Total mechanical work performed during each 10-s sprint. (B) Individual (grey lines) and mean total work (black line) completed over the entire repeated-sprint protocol. Values are presented as mean \pm SD.

<https://doi.org/10.1371/journal.pone.0222487.g003>

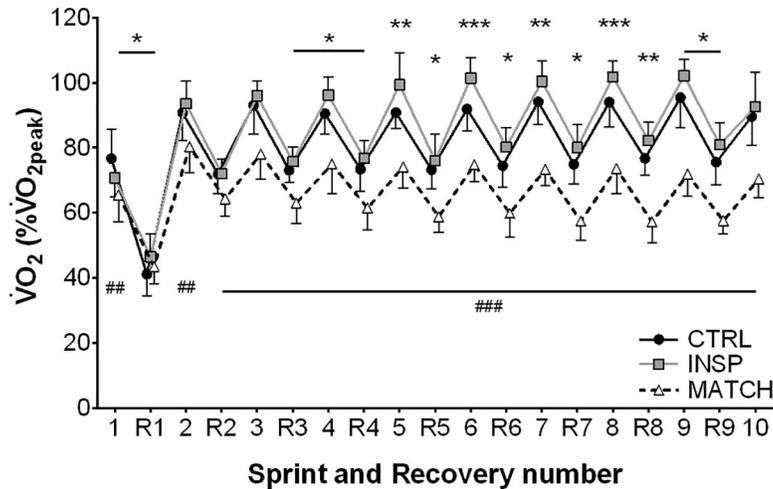


Fig 4. Sprint and recovery pulmonary oxygen uptake ($\dot{V}O_2$) expressed as a percentage of $\dot{V}O_{2peak}$ for control (CTRL), inspiratory muscle loading (INSP) and worked matched (MATCH) exercise. The symbols represent comparisons between INSP and CTRL (\square), INSP and MATCH ($\#$). The number of symbols; one, two and three denote likely, very likely and almost certainly respectively, that the chance of the true effect exceeds a small (-0.2–0.2) effect size. Values are presented as mean \pm SD.

<https://doi.org/10.1371/journal.pone.0222487.g004>

during INSP compared to CTRL (9.1% \pm 4.0%; ES 1.22 \pm 0.51), and higher during MATCH compared to INSP (7.7% \pm 4.2; ES 1.03 \pm 0.54). There were *unclear* differences for S_pO_2 between both INSP and CTRL (-0.1% \pm 0.5; ES -0.10 \pm 0.43), and, MATCH and INSP (0.2% \pm 0.4%; ES 0.16 \pm 0.34).

Differences for HR were *unclear* between INSP and CTRL (1.0% \pm 4.4%; ES 0.11 \pm 0.50). However, there was a clear *almost certainly* large effect between MATCH and INSP (-15.6% \pm 3.3; ES -2.66 \pm 0.61).

Subjects self-report of $RPE_{Exercise}$ during INSP was *likely* trivial after sprint 1 (-2% \pm 0.5%; ES -0.08 \pm 0.21) compared to CTRL, and *unclear* after sprint 5 (0.5% \pm 1.1%; ES 0.14 \pm 0.52) and sprint 10 (0.4% \pm 1.0%; ES 0.19 \pm 0.49). Whereas RPE_{Breath} was *almost certainly* higher after sprint 1 (2.8% \pm 0.7%; ES 1.46 \pm 0.37), sprint 5 (2.4% \pm 1.0; ES 1.79 \pm 0.77), and sprint 10 (2.7% \pm 1.2%; ES 1.32 \pm 0.57). Comparing MATCH to INSP, $RPE_{Exercise}$ was *almost certainly* lower after sprint 1 (-2.1% \pm 0.8%; ES -0.98 \pm 0.37), sprint 5 (-4.4% \pm 1.3%; ES -2.78 \pm 0.79), and sprint 10 (-5.6% \pm 1.2; ES -3.43 \pm 0.73). Similarly, RPE_{Breath} was *almost certainly* lower after sprint 1 (-4.0% \pm 1.1%; ES -1.69 \pm 0.46), sprint 5 (-6.1% \pm 2.0%; ES -2.36 \pm 0.78), and sprint 10 (-6.8% \pm 1.6%; ES -3.93 \pm 0.95)

Table 3. Near-infrared spectroscopy responses to repeated-sprint exercise during control (CTRL), inspiratory loading (INSP), and work match (MATCH) conditions.

Variable	CTRL	INSP	MATCH
TTP _{HHb} (s)	13.0 \pm 3.3	12.3 \pm 3.4	13.8 \pm 3.7 #
Δ Reoxy (%)	49.94 \pm 20.30	54.66 \pm 19.24	49.45 \pm 21.08
Reoxy rate (% \cdot s ⁻¹)	2.16 \pm 0.78	2.20 \pm 0.75	2.27 \pm 0.79

Values are reported as mean \pm SD. Abbreviations are: time to peak deoxy-haemoglobin, TTP_{HHb}; reoxygenation, Δ Reoxy; reoxygenation rate, Reoxy rate. The symbols represent comparisons between INSP and CTRL (\square), INSP and MATCH ($\#$). The number of symbols; one, two and three denote likely, very likely and almost certainly respectively, that the chance of the true effect exceeds a small (-0.2–0.2) effect size. Values are reported as mean \pm SD.

<https://doi.org/10.1371/journal.pone.0222487.t003>

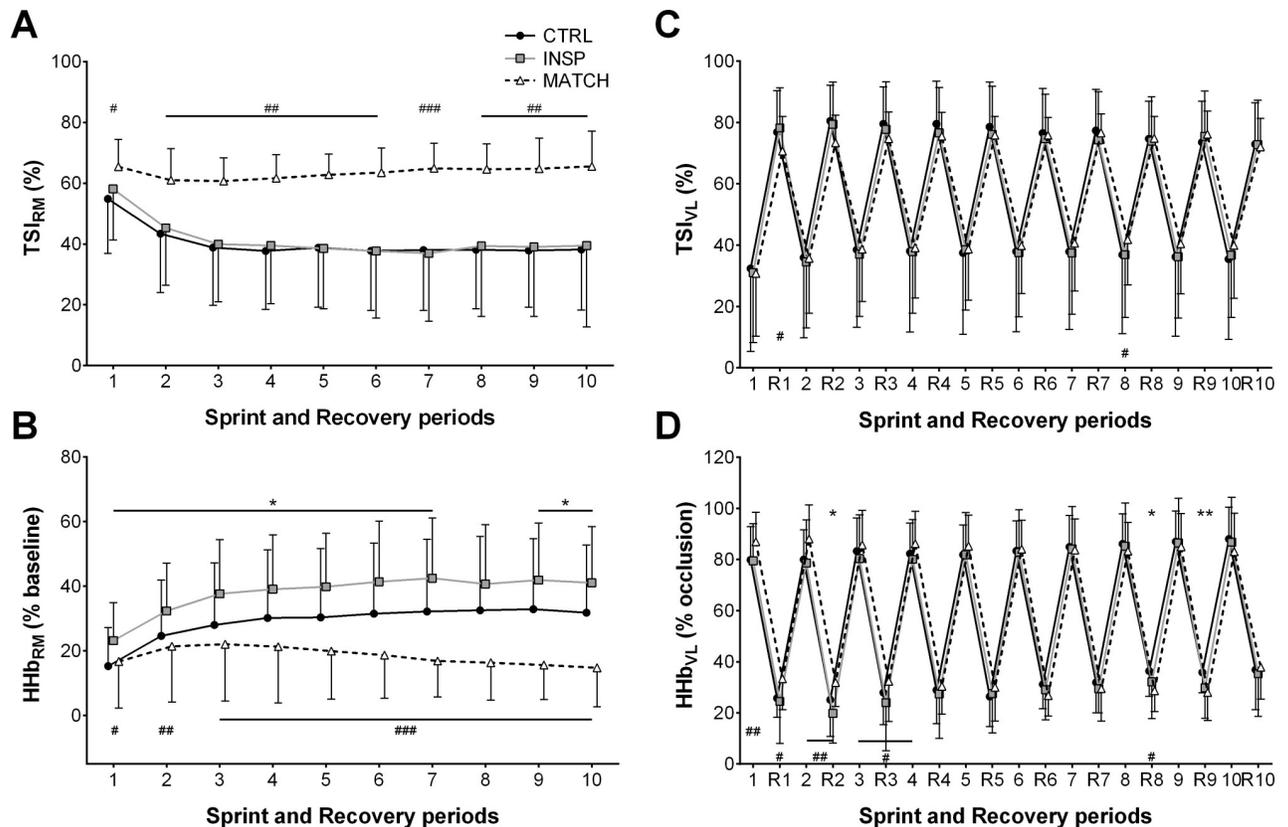


Fig 5. Respiratory and locomotor muscle NIRS responses to repeated-sprints during the control (CTRL) inspiratory loading (INSP) and work matched (MATCH) experiment sessions. (A) Respiratory muscle tissue saturation index (TSI_{RM}) ($n = 8$). (B) Respiratory muscle deoxy-haemoglobin (HHb_{RM}). (C) Vastus lateralis tissue saturation index (TSI_{VL}). (D) Vastus lateralis deoxy-haemoglobin (HHb_{VL}). Values are presented as mean \pm SD. The symbols represent comparisons between INSP and Control (*), INSP and MATCH (#). The number of symbols; one, two and three denote likely, very likely and almost certainly respectively, that the chance of the true effect exceeds a small (-0.2 – 0.2) effect size. Values are presented as mean \pm SD.

<https://doi.org/10.1371/journal.pone.0222487.g005>

Muscle oxygenation

Muscle oxygenation responses to exercise are presented in Table 3 and Figs 5, 6, 7 and 8. Differences were *unclear* between INSP and CTRL for TSI_{RM} ($1.0\% \pm 7.5\%$), but MATCH was *very likely* lower than INSP ($22.1\% \pm 12.5\%$). Average HHb_{RM} was *likely* greater during INSP compared to CTRL ($9.0\% \pm 7.5\%$). Conversely, HHb_{RM} was lower during MATCH compared to INSP ($-19.6\% \pm 6.0\%$).

Differences of sprint TSI_{VL} ($-0.3\% \pm 9.5\%$) and recovery TSI_{VL} ($-1.1\% \pm 7.0\%$) were *unclear* between INSP and CTRL. Similarly, the differences between INSP and CTRL for both sprint HHb_{VL} ($-1.1\% \pm 5.1\%$) and recovery HHb_{VL} ($-2.7\% \pm 5.4\%$) were *unclear*. There was no meaningful difference between INSP and CTRL for TTP_{HHb} ($-5.8\% \pm 6.0\%$) and $\Delta Reoxy$ ($4.7\% \pm 8.3$). Additionally, there was an *unclear* difference in Reoxy rate ($0.0\% \pm 0.3\%$).

In MATCH exercise, differences in TSI_{VL} were *unclear* compared to INSP for both sprint ($2.2\% \pm 6.7\%$), and recovery ($-1.4\% \pm 7.6$) phases. Additionally, there was no meaningful difference in sprint HHb_{VL} ($-1.1\% \pm 5.1\%$), and an *unclear* difference for recovery HHb_{VL} ($3.0\% \pm 5.7\%$). The TTP_{HHb} was greater during MATCH than INSP ($12.0\% \pm 14.4\%$). There were also *unclear* differences in $\Delta Reoxy$ ($-5.2\% \pm 11.5\%$), and Reoxy rate ($0.1\% \pm 0.4\%$).

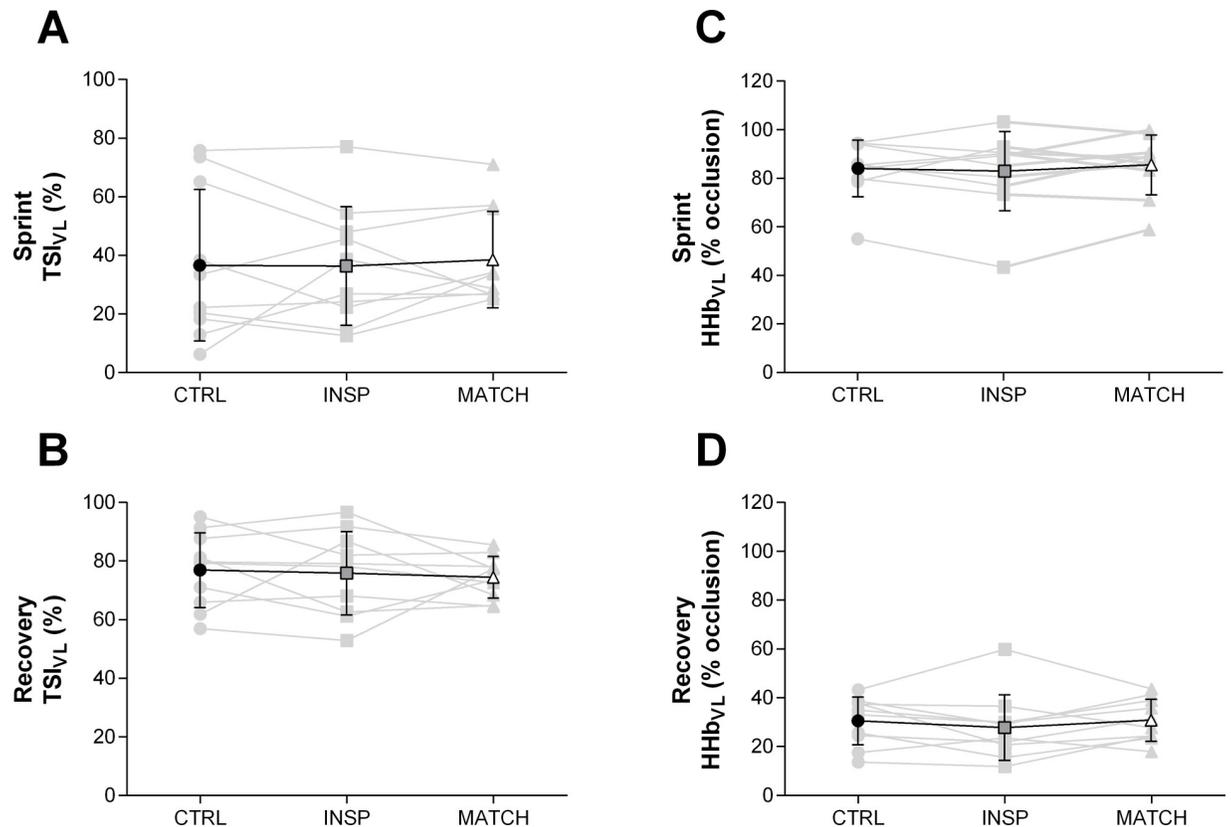


Fig 6. Mean locomotor muscle NIRS responses to repeated-sprints during the control (CTRL) inspiratory loading (INSP) and work matched (MATCH) experiment sessions. (A) Sprint vastus lateralis tissue saturation index (Sprint TSI_{VL}). (B) Recovery vastus lateralis tissue saturation index (Recovery TSI_{VL}). (C) Sprint vastus lateralis deoxy-haemoglobin (Sprint HHb_{VL}). (D) Recovery vastus lateralis deoxy-haemoglobin (Recovery HHb_{VL}). Individual responses are represented by the grey lines and mean responses are represented by black lines. The symbols represent comparisons between INSP and Control (\circ), INSP and MATCH ($\#$). The number of symbols; one, two and three denote likely, very likely and almost certainly respectively, that the chance of the true effect exceeds a small (-0.2–0.2) effect size. Vales are presented as mean \pm SD.

<https://doi.org/10.1371/journal.pone.0222487.g006>

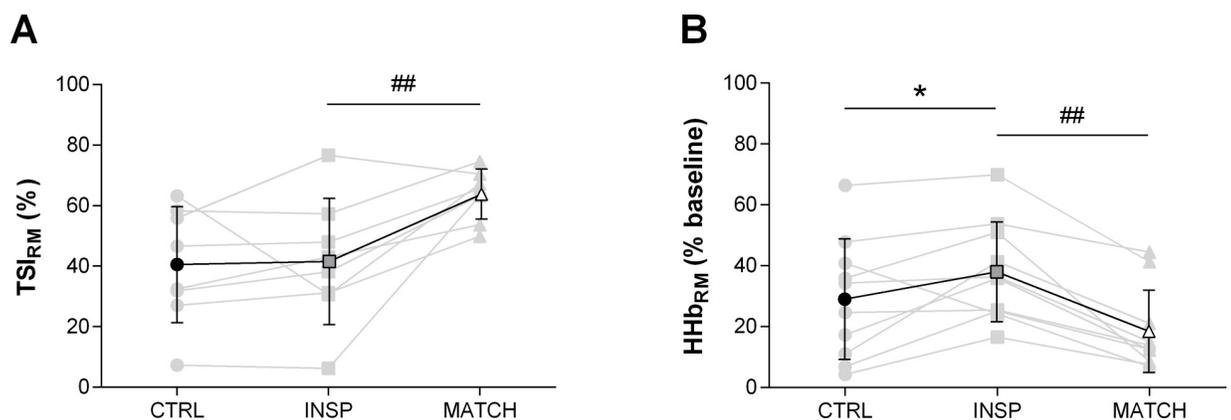


Fig 7. Mean respiratory muscle NIRS responses to repeated-sprints during the control (CTRL) inspiratory loading (INSP) and work matched (MATCH) experiment sessions. (A) Respiratory muscle tissue saturation index (TSI_{RM}) (n = 8). (B) Respiratory muscle deoxy-haemoglobin (HHb_{RM}). Individual responses are represented by the grey lines and mean responses are represented by black lines. The symbols represent comparisons between INSP and Control (\circ), INSP and MATCH ($\#$). The number of symbols; one, two and three denote likely, very likely and almost certainly respectively, that the chance of the true effect exceeds a small (-0.2–0.2) effect size. Vales are presented as mean \pm SD.

<https://doi.org/10.1371/journal.pone.0222487.g007>

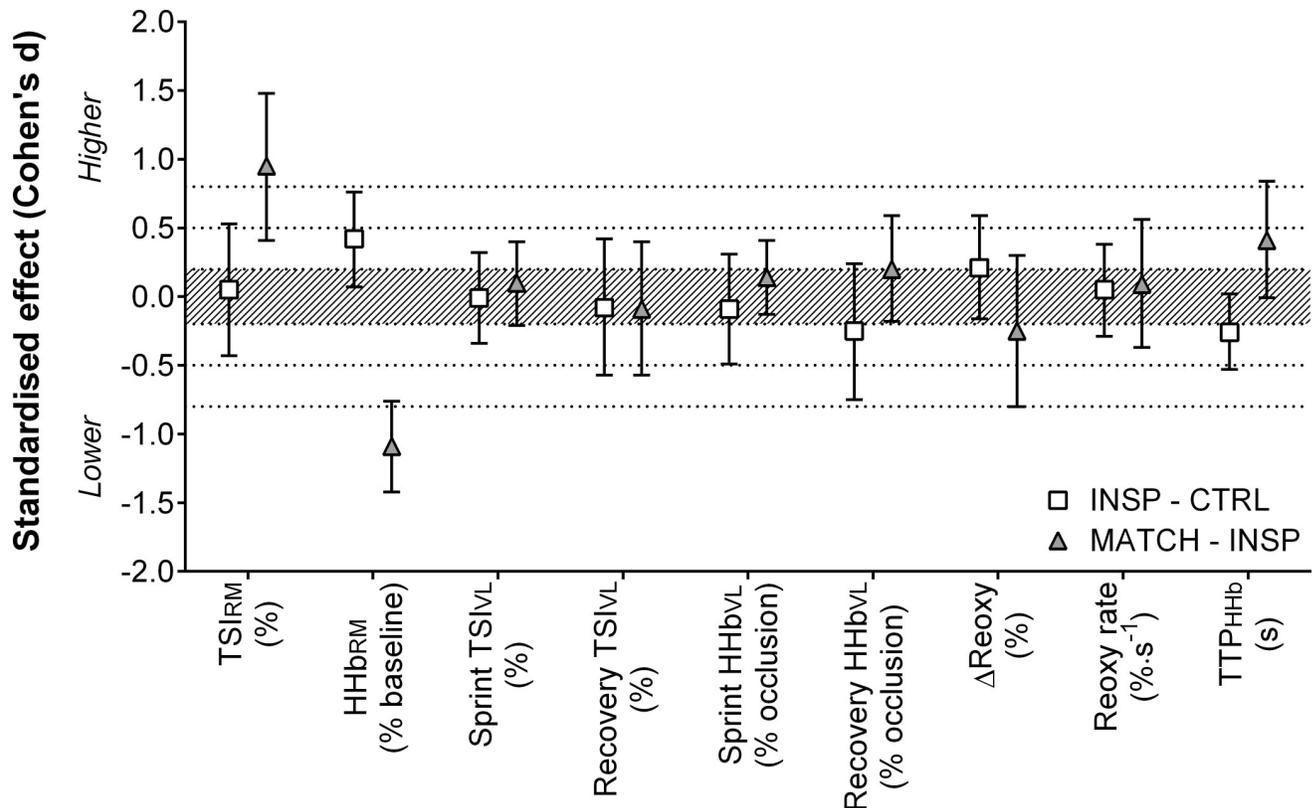


Fig 8. Standardised effects (Cohen's d) with 90% confidence intervals for NIRS variables comparing inspiratory loading (INSP) to control (CTRL), and work matched exercise (MATCH) to INSP. Shaded area indicates a trivial effect, and dotted lines thresholds for small, moderate and large effects. Abbreviations are: respiratory muscle tissue saturation index, TSI_{RM} ; respiratory muscle deoxy-haemoglobin, HHb_{RM} ; vastus lateralis tissue saturation index, TSI_{VL} ; vastus lateralis deoxy-haemoglobin, HHb_{VL} ; vastus lateralis reoxygenation, $\Delta Reoxy$; vastus lateralis reoxygenation rate, Reoxy rate; time to peak deoxy-haemoglobin, TTP_{HHb} .

<https://doi.org/10.1371/journal.pone.0222487.g008>

Discussion

This study examined intercostal and vastus lateralis muscle oxygenation trends during repeated-sprint exercise with heightened respiratory muscle work. The addition of inspiratory loading increased mouth pressure and respiratory muscle deoxygenation. However, these altered responses had no meaningful impact on blood arterial O_2 saturation and tissue oxygenation trends within the vastus lateralis locomotor muscle. We interpret these findings to suggest that during maximal intermittent work, O_2 delivery to the respiratory and locomotor muscles can be maintained. Unlike during continuous aerobic exercise [24, 25], the significant increase in $\dot{V}O_2$ appears to compensate for the elevated metabolic demands of the respiratory muscles when relatively short periods of rest separate bouts of all-out sprint.

Work of breathing and respiratory muscle oxygenation

Hyperpnoea during high-intensity exercise requires a considerable portion of whole-body $\dot{V}O_2$ to support the metabolic demands of the respiratory muscles [16], and is increased when an inspiratory load is added [19]. In the present study, pulmonary $\dot{V}O_2$ was elevated by 4–5% during both the sprint and recovery phases of the repeated-sprint protocol when an inspiratory load was added. This occurred even though there was no meaningful difference in total locomotor work completed during the INSP and CTRL experiment sessions.

When subjects exercised with the inspiratory load, \dot{V}_E was 19.6% lower compared to CTRL, achieved via a reduction in f_R . Regardless of these changes, S_{pO_2} was not different between exercise conditions. Either consciously or subconsciously choosing a lower \dot{V}_E , subjects may have counteracted, or at least attenuated, the expected work of breathing with INSP. When the change in f_R was accounted for, respiratory muscle force developments ($\int P_m \times f_R$) was substantially higher during INSP compared to CTRL (734%). Higher $\int P_m \times f_R$ in conjunction with the elevated HHb_{RM} in the current study, suggests that O_2 utilisation by the respiratory muscles was increased with inspiratory loading, as shown previously [20, 24]. Furthermore, there were *unclear* differences in TSI_{RM} , which when considered concomitantly with an elevated HHb_{RM} , suggests that $[O_2Hb]$ was higher to match the demands for O_2 delivery of the respiratory muscles. Previous studies have shown similar changes in HHb_{RM} in response to inspiratory loading [24] and resistive breathing [25]. The present data further suggests maintenance of respiratory muscle oxygenation with inspiratory loading which contrasts with others who reported similar values for $\int P_m \times f_R$ [38]. However, preserved respiratory muscle oxygenation may have negative consequences for exercise tolerance if blood flow is redistributed away from the active limbs to meet the metabolic demands of breathing.

During continuous high-intensity exercise when the work of breathing is high, there can be an increase in vascular resistance and a reduction in limb perfusion [18, 19, 39]. The accumulation of metabolites in the respiratory muscles stimulates group IV afferent discharge in these muscles [40], leading to sympathetically mediated efferent discharge and vasoconstriction in the locomotor muscles [18, 41]. Despite the clear increase in respiratory muscle deoxygenation with the addition of an inspiratory load, there was no clear negative effect on vastus lateralis oxygenation. The intermittent nature of repeated-sprint exercise may probably allow sufficient recovery time to prevent the accumulation of fatigue-inducing metabolites.

Locomotor muscle oxygenation

Inspiratory loading had no discernible effects on sprint HHb_{VL} or TSI_{VL} despite a considerable increase in the work of breathing and respiratory muscle O_2 utilisation (Figs 5, 6 and 8). At sprint onset there is a rapid decrease in muscle oxygenation as evidenced by a reduction in TSI_{VL} and an elevation in HHb_{VL} , and as sprints are repeated, there is an eventual plateau [1, 11, 12, 14]. This suggests that a maximal level of O_2 extraction in the locomotor muscles is achieved in “normal” exercise conditions [42]. However, a higher secondary ceiling point to vastus lateralis [HHb] has been observed when repeated-sprint exercise has been performed in simulated altitude (normobaric hypoxia) [13, 22]. Elevated muscle [HHb] during maximal exercise may compensate for reduced muscle O_2 availability [13, 43]. If vastus lateralis O_2 availability had been impacted in the present study by an elevated work of breathing, we would have expected sprint HHb_{VL} to be elevated during the INSP trial compared to CTRL. Nevertheless, vastus lateralis oxygenation during the sprint phase per se may play a limited role in prolonged repeated-sprint performance. Muscle O_2 availability during the recovery phase appears to be far more influential in maintaining performance as sprints are repeated [13]. The capacity to reoxygenate muscle tissues between sprints is highly sensitive to O_2 availability and underpins metabolic recovery between sprint bouts [11, 13–15]. Even with the addition of an inspiratory load in this study, which increased respiratory muscle O_2 utilisation, vastus lateralis tissue oxygenation was maintained, as evidenced by the similarity of TSI_{VL} between the conditions. Since TSI reflects the balance between O_2 availability and utilisation, if either $[O_2Hb]$ decreased or $[HHb]$ increased, we would have observed a reduction in TSI during INSP. It therefore appears that the cardiovascular system can support the metabolic O_2 demands of both the respiratory and locomotor muscles during repeated-sprint exercise.

It is likely that locomotor muscle oxygenation will only be compromised when cardiac output can no longer increase to meet the O_2 demands of both the respiratory and locomotor muscles simultaneously. It has been demonstrated that while exercising at near-maximal work rates, there is no accompanying increase in cardiac output and $\dot{V}O_2$ with inspiratory loading, and as a result limb blood flow is compromised [18, 19]. This presumably occurs when the prescribed exercise intensity is sufficient to elicit sustained $\dot{V}O_{2peak}$, and therefore, no further increase can occur. In the present study there was a 4–5% increase in $\dot{V}O_2$ throughout the repeated-sprint protocol with inspiratory loading, which is similar to others having shown an increase in $\dot{V}O_2$ during submaximal exercise [20]. These data demonstrate that during repeated-sprint exercise, there is additional capacity for $\dot{V}O_2$ increase to meet the heightened metabolic demands of inspiratory loading, which may be a crucial factor in maintaining locomotor muscle oxygenation. Secondly, the intermittent nature of repeated-sprint exercise will minimise the development of diaphragm fatigue and the activation of the respiratory muscle metaboreflex which is promoted during sustained high-intensity exercise [17]. As shown in Fig 4, pulmonary $\dot{V}O_2$ fluctuated between 90% and 70% of $\dot{V}O_{2peak}$ during sprint and recovery phases, respectively, during the control condition. Moreover, the addition of an inspiratory load increased $\dot{V}O_2$ by $\approx 5\%$ in both the sprint and recovery phases. The fluctuation in metabolic demands exemplified by the undulating $\dot{V}O_2$ likely minimises the opportunity for substantial competition of available cardiac output to develop, and the impairment of tissue reoxygenation.

Hyperventilation was present in both CTRL, and to a lesser degree, INSP experiment sessions, which may have had a protective effect on vastus lateralis oxygenation. Hyperventilation is associated with an increase in alveolar ventilation disproportionate to $\dot{V}O_2$ (pressure of alveolar O_2 increases), and P_ACO_2 (pressure of alveolar CO_2 decreases) [44]. This is a potential mechanism associated with high-intensity exercise which can constrain a fall in arterial O_2 and pH [44, 45]. Despite a reduction of \dot{V}_E in a state of heightened O_2 demand during the INSP session, S_pO_2 was maintained. In studies where vastus lateralis tissue oxygenation was impaired during exercise with resistive breathing and inspiratory loading, there was moderate exercise-induced arterial hypoxemia (88–93%) [24, 25, 46]. Exercise-induced arterial hypoxemia is a known limiting factor of exercise [46], and preventing it with supplemental O_2 can attenuate peripheral muscle fatigue [47]. Though exercise-induced arterial hypoxemia has been observed during similar repeated-sprint exercise protocols [12], there was no evidence of its occurrence in the present study. We acknowledge that interpretation of fingertip S_pO_2 data in cycling may be affected by some data loss caused by hand movement artefact and tight gripping of the handlebars [25, 48], and that monitoring at the earlobe could potentially attenuate this limitation [24, 49]. However, on close inspection of our S_pO_2 data, we do not believe that the minimal data loss we experienced affected our results and interpretation of the data. Nonetheless, we conclude that alterations in repeated-sprint oxygenation caused by an elevated work of breathing may only occur if a moderate level of exercise-induced arterial hypoxemia takes place.

The level of inspiratory loading may have also had an influence on the outcomes in this study. In previous work, inspiratory loading was achieved by reducing the inspiratory aperture to 10 mm and 8 mm [24]. Changes in [HHb] of the exercising limb were only detected with the smaller opening. Similarly, when resistive breathing has been used, the most noticeable changes in tissue oxygenation trends occurred when the aperture was reduced to 4.5 mm [25]. One may therefore argue that the inspiratory work in the present study was too low to induce a respiratory muscle metaboreflex, however, we reported peak P_m and $\int P_m \times f_R$ to be similar

to previous work using an 8-mm aperture [24, 38]. Additionally, \dot{V}_E was considerably higher than in previous work [24, 25], which would have contributed to a heightened work of breathing.

Work-matched exercise

To our knowledge, this is the first time repeated work matched bouts of exercise have been used to examine the demands of repeated-sprint exercise under altered metabolic conditions. Despite a similar degree of vastus lateralis tissue deoxygenation during the work-matched sprints (Figs 5 and 6), the physiological load placed on the cardiovascular system was considerably lower. This is evidenced by the consistently lower $\dot{V}O_2$, probably due to markedly lower respiratory muscle O_2 utilisation. But more importantly, $\dot{V}O_2$ was heavily influenced by how exercise was prescribed. Matching total work was achieved by replicating mean power output for each sprint, and therefore was lacking maximal acceleration and power production typically associated with sprint exercise. For example, there was a clear difference in peak power output between INSP (1097 ± 148 W) and MATCH (773 ± 122 W) exercise conditions. In a typical sprint, power output peaks within 1–2 s and progressively declines as the sprint continues [23, 50]. When sprints are repeated and separated by incomplete recovery periods, both peak and mean power output gradually decline as fatigue accumulates [1–4, 49]. By matching for mean power output and eliminating the opportunity for subjects to “maximally” exert themselves, the reliance on intramuscular ATP and PCr hydrolysis would have been reduced [2, 51], and metabolic perturbations associated with maximal exercise minimised [2, 5, 51]. Despite a substantial decrease in $\dot{V}O_2$ and the O_2 cost associated with the work of breathing, there was no substantial difference in $\Delta Reoxy$ or in $Reoxy$ rate. This implies that tissue reoxygenation was maximal in all exercise conditions since the now “available” cardiac output was not being utilised to reoxygenate the locomotor muscles [18]. It seems that there exists some degree of reserve in the cardiovascular system that is called upon to maintain respiratory and locomotor muscle oxygenation when the work of breathing is high. Therefore, the O_2 cost of breathing in repeated-sprint cycling is unlikely to have a meaningful negative impact on locomotor muscle oxygenation.

Conclusions

A crucial factor of repeated-sprint performance is the reoxygenation capacity between sprint bouts [11, 13, 14]. We further tested this mechanism by increasing the work of breathing, which is known to negatively influence limb blood flow and O_2 delivery at least in endurance exercise [16–18, 39]. The present data demonstrate that the addition of inspiratory loading did not impair oxygenation of the vastus lateralis. When maximal exercise is interspersed with short rest periods, the cardiovascular system appears to maintain O_2 delivery to both the locomotor and respiratory muscles in a state of heightened inspiratory muscle work.

Author Contributions

Conceptualization: Ramón F. Rodriguez, Nathan E. Townsend, Robert J. Aughey, François Billaut.

Data curation: Ramón F. Rodriguez.

Formal analysis: Ramón F. Rodriguez.

Funding acquisition: Ramón F. Rodriguez.

Investigation: Ramón F. Rodriguez.

Methodology: Ramón F. Rodríguez, Nathan E. Townsend, Robert J. Aughey, François Billaut.

Project administration: Ramón F. Rodríguez.

Resources: Ramón F. Rodríguez.

Software: Ramón F. Rodríguez.

Supervision: Nathan E. Townsend, Robert J. Aughey, François Billaut.

Validation: Ramón F. Rodríguez.

Visualization: Ramón F. Rodríguez.

Writing – original draft: Ramón F. Rodríguez.

Writing – review & editing: Ramón F. Rodríguez, Nathan E. Townsend, Robert J. Aughey, François Billaut.

References

1. Racinais S, Bishop DJ, Denis R, Lattier G, Mendez-Villanueva A, Perrey S. Muscle deoxygenation and neural drive to the muscle during repeated sprint cycling. *Medicine and Science in Sports and Exercise*. 2007; 39(2):268–74. Epub 2007/02/06. <https://doi.org/10.1249/01.mss.0000251775.46460.cb> PMID: 17277590.
2. Gaitanos GC, Williams C, Boobis LH, Brooks S. Human muscle metabolism during intermittent maximal exercise. *Journal of Applied Physiology*. 1993; 75(2):712–9. Epub 1993/08/01. <https://doi.org/10.1152/jappl.1993.75.2.712> PMID: 8226473.
3. Mendez-Villanueva A, Edge J, Suriano R, Hamer P, Bishop DJ. The recovery of repeated-sprint exercise is associated with PCr resynthesis, while muscle pH and EMG amplitude remain depressed. *PLoS ONE*. 2012; 7(12):e51977. Epub 2013/01/04. <https://doi.org/10.1371/journal.pone.0051977> PMID: 23284836; PubMed Central PMCID: PMC3524088.
4. Hureau TJ, Ducrocq GP, Blain GM. Peripheral and Central Fatigue Development during All-Out Repeated Cycling Sprints. *Medicine and Science in Sports and Exercise*. 2016; 48(3):391–401. Epub 2015/10/27. <https://doi.org/10.1249/MSS.0000000000000800> PMID: 26496420.
5. Parolin ML, Chesley A, Matsos MP, Spriet LL, Jones NL, Heigenhauser GJF. Regulation of skeletal muscle glycogen phosphorylase and PDH during maximal intermittent exercise. *Am J Physiol*. 1999; 277(5):E890–900. <https://doi.org/10.1152/ajpendo.1999.277.5.E890> WOS:000083598700016. PMID: 10567017
6. Harris RC, Edwards RH, Hultman E, Nordesjö LO, Ny Lind B, Sahlin K. The time course of phosphoryl-creatine resynthesis during recovery of the quadriceps muscle in man. *Pflügers Archiv: European journal of physiology*. 1976; 367(2):137–42. <https://doi.org/10.1007/bf00585149> PMID: 1034909
7. Sahlin K, Harris RC, Hultman E. Resynthesis of creatine phosphate in human muscle after exercise in relation to intramuscular pH and availability of oxygen. *Scandinavian Journal of Clinical and Laboratory Investigation*. 1979; 39(6):551–8. Epub 1979/10/01. <https://doi.org/10.3109/00365517909108833> PMID: 43580.
8. De Blasi RA, Cope M, Elwell C, Safoue F, Ferrari M. Noninvasive measurement of human forearm oxygen consumption by near infrared spectroscopy. *European Journal of Applied Physiology and Occupational Physiology*. 1993; 67(1):20–5. <https://doi.org/10.1007/bf00377698> PMID: 8375359.
9. Grassi B, Pogliaghi S, Rampichini S, Quaresima V, Ferrari M, Marconi C, et al. Muscle oxygenation and pulmonary gas exchange kinetics during cycling exercise on-transitions in humans. *Journal of Applied Physiology*. 2003; 95(1):149–58. <https://doi.org/10.1152/jappphysiol.00695.2002> PMID: 12611769
10. DeLorey DS, Kowalchuk JM, Paterson DH. Relationship between pulmonary O₂ uptake kinetics and muscle deoxygenation during moderate-intensity exercise. *Journal of Applied Physiology*. 2003; 95(1):113–20. Epub 2003/04/08. <https://doi.org/10.1152/jappphysiol.00956.2002> PMID: 12679363.
11. Buchheit M, Cormie P, Abbiss CR, Ahmaidi S, Nosaka KK, Laursen PB. Muscle deoxygenation during repeated sprint running: Effect of active vs. passive recovery. *International Journal of Sports Medicine*. 2009; 30(6):418–25. Epub 2009/05/14. <https://doi.org/10.1055/s-0028-1105933> PMID: 19437381.
12. Smith KJ, Billaut F. Influence of cerebral and muscle oxygenation on repeated-sprint ability. *European Journal of Applied Physiology*. 2010; 109(5):989–99. Epub 2010/04/01. <https://doi.org/10.1007/s00421-010-1444-4> PMID: 20354718.

13. Billaut F, Buchheit M. Repeated-sprint performance and vastus lateralis oxygenation: Effect of limited O₂ availability. *Scandinavian Journal of Medicine and Science in Sports*. 2013; 23(3):185–93. Epub 2013/02/01. <https://doi.org/10.1111/sms.12052> PMID: 23362832.
14. Buchheit M, Ufland P. Effect of endurance training on performance and muscle reoxygenation rate during repeated-sprint running. *European Journal of Applied Physiology*. 2011; 111(2):293–301. Epub 2010/09/28. <https://doi.org/10.1007/s00421-010-1654-9> PMID: 20872150.
15. Jones B, Hamilton DK, Cooper CE. Muscle oxygen changes following sprint interval cycling training in elite field hockey players. *PLoS ONE*. 2015; 10(3):e0120338. <https://doi.org/10.1371/journal.pone.0120338> PMID: 25807517
16. Aaron EA, Seow KC, Johnson BD, Dempsey JA. Oxygen cost of exercise hyperpnea: implications for performance. *Journal of Applied Physiology*. 1992; 72(5):1818–25. Epub 1992/05/01. <https://doi.org/10.1152/jappl.1992.72.5.1818> PMID: 1601791.
17. Dempsey JA, Romer L, Rodman J, Miller J, Smith C. Consequences of exercise-induced respiratory muscle work. *Respiratory Physiology and Neurobiology*. 2006; 151(2–3):242–50. Epub 2006/04/18. <https://doi.org/10.1016/j.resp.2005.12.015> PMID: 16616716.
18. Harms CA, Babcock MA, McClaran SR, Pegelow DF, Nickele GA, Nelson WB, et al. Respiratory muscle work compromises leg blood flow during maximal exercise. *Journal of Applied Physiology*. 1997; 82(5):1573–83. Epub 1997/05/01. <https://doi.org/10.1152/jappl.1997.82.5.1573> PMID: 9134907.
19. Harms CA, Wetter TJ, McClaran SR, Pegelow DF, Nickele GA, Nelson WB, et al. Effects of respiratory muscle work on cardiac output and its distribution during maximal exercise. *Journal of Applied Physiology*. 1998; 85(2):609–18. <https://doi.org/10.1152/jappl.1998.85.2.609> PMID: 9688739
20. Wetter TJ, Harms CA, Nelson WB, Pegelow DF, Dempsey JA. Influence of respiratory muscle work on VO₂ and leg blood flow during submaximal exercise. *Journal of Applied Physiology*. 1999; 87(2):643–51. Epub 1999/08/13. <https://doi.org/10.1152/jappl.1999.87.2.643> PMID: 10444624.
21. Rodriguez RF, Townsend NE, Aughey RJ, Billaut F. Influence of averaging method on muscle deoxygenation interpretation during repeated-sprint exercise. *Scandinavian Journal of Medicine and Science in Sports*. 2018; 28(11):2263–71. Epub 2018/06/09. <https://doi.org/10.1111/sms.13238> PMID: 29883534.
22. Rodriguez RF, Townsend NE, Aughey RJ, Billaut F. Respiratory Muscle Oxygenation is not impacted by Hypoxia during Repeated-sprint Exercise. *Respiratory Physiology and Neurobiology*. 2019; 260:114–21. <https://doi.org/10.1016/j.resp.2018.11.006> PMID: 30453086.
23. Matsuura R, Arimitsu T, Yunoki T, Kimura T, Yamanaka R, Yano T. Effects of heat exposure in the absence of hyperthermia on power output during repeated cycling sprints. *Biology of Sport*. 2015; 32(1):15–20. <https://doi.org/10.5604/20831862.1125286> PMID: 25729145; PubMed Central PMCID: PMC4314599.
24. Turner LA, Tecklenburg-Lund S, Chapman RF, Stager JM, Duke JW, Mickleborough TD. Inspiratory loading and limb locomotor and respiratory muscle deoxygenation during cycling exercise. *Respiratory Physiology and Neurobiology*. 2013; 185(3):506–14. Epub 2012/12/12. <https://doi.org/10.1016/j.resp.2012.11.018> PMID: 23228896.
25. Nielsen HB, Boesen M, Secher NH. Near-infrared spectroscopy determined brain and muscle oxygenation during exercise with normal and resistive breathing. *Acta Physiologica Scandinavica*. 2001; 171(1):63–70. <https://doi.org/10.1046/j.1365-201X.2001.00782.x> PMID: 11350264.
26. Lamarra N, Whipp BJ, Ward SA, Wasserman K. Effect of interbreath fluctuations on characterizing exercise gas exchange kinetics. *Journal of Applied Physiology*. 1987; 62(5):2003–12. Epub 1987/05/01. <https://doi.org/10.1152/jappl.1987.62.5.2003> PMID: 3110126.
27. Rossiter HB, Howe FA, Ward SA, Kowalchuk JM, Griffiths JR, Whipp BJ. Intersample fluctuations in phosphocreatine concentration determined by ³¹P-magnetic resonance spectroscopy and parameter estimation of metabolic responses to exercise in humans. *The Journal of Physiology*. 2000; 528 Pt 2:359–69. Epub 2000/10/18. <https://doi.org/10.1111/j.1469-7793.2000.t01-1-00359.x> PMID: 11034625; PubMed Central PMCID: PMC2270138.
28. Witt JD, Guenette JA, Rupert JL, McKenzie DC, Sheel AW. Inspiratory muscle training attenuates the human respiratory muscle metaboreflex. *The Journal of Physiology*. 2007; 584(3):1019–28. <https://doi.org/10.1113/jphysiol.2007.140855> PMID: 17855758
29. Takaishi T, Sugiura T, Katayama K, Sato Y, Shima N, Yamamoto T, et al. Changes in blood volume and oxygenation level in a working muscle during a crank cycle. *Medicine and Science in Sports and Exercise*. 2002; 34(3):520–8. Epub 2002/03/07. <https://doi.org/10.1097/00005768-200203000-00020> PMID: 11880818.
30. Perrey S, Ferrari M. Muscle Oximetry in Sports Science: A Systematic Review. *Sports Medicine*. 2018; 48(3):597–616. Epub 2017/11/28. <https://doi.org/10.1007/s40279-017-0820-1> PMID: 29177977.

31. Wolf M, Ferrari M, Quaresima V. Progress of near-infrared spectroscopy and topography for brain and muscle clinical applications. *Journal of biomedical optics*. 2007; 12(6):062104. Epub 2008/01/01. <https://doi.org/10.1117/1.2804899> PMID: 18163807.
32. Kovacsova Z, Bale G, Mitra S, de Roever I, Meek J, Robertson N, et al. Investigation of Confounding Factors in Measuring Tissue Saturation with NIRS Spatially Resolved Spectroscopy. *Advances in Experimental Medicine and Biology*. 2018; 1072:307–12. Epub 2018/09/05. https://doi.org/10.1007/978-3-319-91287-5_49 PMID: 30178363; PubMed Central PMCID: PMC6142855.
33. Niemeijer VM, Jansen JP, van Dijk T, Spee RF, Meijer EJ, Kemps HM, et al. The influence of adipose tissue on spatially resolved near-infrared spectroscopy derived skeletal muscle oxygenation: the extent of the problem. *Physiological measurement*. 2017; 38(3):539–54. Epub 2017/02/06. <https://doi.org/10.1088/1361-6579/aa5dd5> PMID: 28151429.
34. Messere A, Roatta S. Influence of cutaneous and muscular circulation on spatially resolved versus standard Beer-Lambert near-infrared spectroscopy. *Physiological Reports*. 2013; 1(7):e00179. Epub 2014/04/20. <https://doi.org/10.1002/phy2.179> PMID: 24744858; PubMed Central PMCID: PMC3970749.
35. Hopkins WG. Spreadsheets for analysis of controlled trials, with adjustment for a subject characteristic. *Sportscience*. 2006; 10:46–50.
36. Cohen J. *Statistical power analysis for the behavioral sciences*: Routledge; 1988.
37. Hopkins WG, Marshall SW, Batterham AM, Hanin J. Progressive statistics for studies in sports medicine and exercise science. *Medicine and Science in Sports and Exercise*. 2009; 41(1):3–13. Epub 2008/12/19. <https://doi.org/10.1249/MSS.0b013e31818cb278> PMID: 19092709.
38. Turner LA, Tecklenburg-Lund SL, Chapman R, Shei RJ, Wilhite DP, Mickleborough T. The Effect of Inspiratory Muscle Training on Respiratory and Limb Locomotor Muscle Deoxygenation During Exercise with Resistive Inspiratory Loading. *International Journal of Sports Medicine*. 2016;(EFirst). Epub 18.05.2016. <https://doi.org/10.1055/s-0042-104198> PMID: 27191210
39. Sheel AW, Derchak PA, Morgan BJ, Pegelow DF, Jacques AJ, Dempsey JA. Fatiguing inspiratory muscle work causes reflex reduction in resting leg blood flow in humans. *The Journal of Physiology*. 2001; 537(Pt 1):277–89. Epub 2001/11/17. <https://doi.org/10.1111/j.1469-7793.2001.0277k.x> PMID: 11711580; PubMed Central PMCID: PMC2278925.
40. Hill JM. Discharge of group IV phrenic afferent fibers increases during diaphragmatic fatigue. *Brain Research*. 2000; 856(1–2):240–4. [https://doi.org/10.1016/S0006-8993\(99\)02366-5](https://doi.org/10.1016/S0006-8993(99)02366-5) PMID: 10677632.
41. St Croix CM, Morgan BJ, Wetter TJ, Dempsey JA. Fatiguing inspiratory muscle work causes reflex sympathetic activation in humans. *The Journal of Physiology*. 2000; 529(2):493–504. <https://doi.org/10.1111/j.1469-7793.2000.00493.x> PMID: 11101657
42. Esaki K, Hamaoka T, Radegran G, Boushel R, Hansen J, Katsumura T, et al. Association between regional quadriceps oxygenation and blood oxygen saturation during normoxic one-legged dynamic knee extension. *European Journal of Applied Physiology*. 2005; 95(4):361–70. Epub 2005/08/13. <https://doi.org/10.1007/s00421-005-0008-5> PMID: 16096839.
43. Legrand R, Ahmaidi S, Moalla W, Chocquet D, Marles A, Prieur F, et al. O₂ arterial desaturation in endurance athletes increases muscle deoxygenation. *Medicine and Science in Sports and Exercise*. 2005; 37(5):782–8. Epub 2005/05/05. <https://doi.org/10.1249/01.mss.0000161806.47058.40> PMID: 15870632.
44. Forster HV, Haouzi P, Dempsey JA. Control of breathing during exercise. *Comprehensive Physiology*. 2012; 2(1):743–77. Epub 2012/01/01. <https://doi.org/10.1002/cphy.c100045> PMID: 23728984.
45. Whipp BJ, Ward SA. Determinants and control of breathing during muscular exercise. *British Journal of Sports Medicine*. 1998; 32(3):199–211. <https://doi.org/10.1136/bjism.32.3.199> PMC1756098. PMID: 9773167
46. Dempsey JA, Wagner PD. Exercise-induced arterial hypoxemia. *Journal of Applied Physiology*. 1999; 87(6):1997–2006. <https://doi.org/10.1152/jappl.1999.87.6.1997> PMID: 10601141
47. Romer LM, Haverkamp HC, Lovering AT, Pegelow DF, Dempsey JA. Effect of exercise-induced arterial hypoxemia on quadriceps muscle fatigue in healthy humans. *American Journal of Physiology—Regulatory, Integrative and Comparative Physiology*. 2006; 290(2):R365–R75. <https://doi.org/10.1152/ajpregu.00332.2005> PMID: 16166208
48. Yamaya Y, Bogaard HJ, Wagner PD, Niizeki K, Hopkins SR. Validity of pulse oximetry during maximal exercise in normoxia, hypoxia, and hyperoxia. *Journal of Applied Physiology*. 2002; 92(1):162–8. <https://doi.org/10.1152/jappphysiol.00409.2001> PMID: 11744656.
49. Billaut F, Kerris JP, Rodriguez RF, Martin DT, Gore CJ, Bishop DJ. Interaction of central and peripheral factors during repeated sprints at different levels of arterial O₂ saturation. *PLoS ONE*. 2013; 8(10):e77297. <https://doi.org/10.1371/journal.pone.0077297> PMID: 24155938; PubMed Central PMCID: PMC3796493.

50. Bogdanis GC, Nevill ME, Boobis LH, Lakomy HK. Contribution of phosphocreatine and aerobic metabolism to energy supply during repeated sprint exercise. *Journal of Applied Physiology*. 1996; 80(3):876–84. Epub 1996/03/01. <https://doi.org/10.1152/jappl.1996.80.3.876> PMID: 8964751.
51. Glaister M. Multiple sprint work: physiological responses, mechanisms of fatigue and the influence of aerobic fitness. *Sports Medicine*. 2005; 35(9):757–77. Epub 2005/09/06. <https://doi.org/10.2165/00007256-200535090-00003> PMID: 16138786.