



VICTORIA UNIVERSITY
MELBOURNE AUSTRALIA

APE1/Ref-1 as a Therapeutic Target for Inflammatory Bowel Disease

This is the Published version of the following publication

Sahakian, Lauren, Robinson, Ainsley, Sahakian, Linda, Stavely, Rhian, Kelley, Mark R and Nurgali, Kulmira (2023) APE1/Ref-1 as a Therapeutic Target for Inflammatory Bowel Disease. *Biomolecules*, 13 (11). ISSN 2218-273X

The publisher's official version can be found at
<https://www.mdpi.com/2218-273X/13/11/1569>

Note that access to this version may require subscription.

Downloaded from VU Research Repository <https://vuir.vu.edu.au/47592/>

Review

APE1/Ref-1 as a Therapeutic Target for Inflammatory Bowel Disease

Lauren Sahakian ¹, Ainsley M. Robinson ¹, Linda Sahakian ², Rhian Stavely ^{2,3} , Mark R. Kelley ^{4,5} 
and Kulmira Nurgali ^{1,2,6,*} 

¹ Institute for Health & Sport, Victoria University, Melbourne, VIC 3021, Australia; lauren.sahakian@live.vu.edu.au (L.S.); ainsley.robinson@vu.edu.au (A.M.R.)

² Department of Medicine Western Health, The University of Melbourne, Melbourne, VIC 3010, Australia; lindasahakian@hotmail.com (L.S.); rstavely@mgh.harvard.edu (R.S.)

³ Department of Pediatric Surgery, Massachusetts General Hospital, Harvard Medical School, Boston, MA 02114, USA

⁴ Department of Pediatrics, Herman B Wells Center for Pediatric Research, Indiana University School of Medicine, Indianapolis, IN 46202, USA; mkelley@iu.edu

⁵ Department of Pharmacology and Toxicology, Indiana University School of Medicine, Indianapolis, IN 46202, USA

⁶ Regenerative Medicine and Stem Cells Program, Australian Institute for Musculoskeletal Science (AIMSS), Melbourne, VIC 3021, Australia

* Correspondence: kulmira.nurgali@vu.edu.au; Tel.: +61-3-8395-8223

Abstract: Inflammatory bowel disease (IBD) is characterized by chronic relapsing inflammation of the gastrointestinal tract. The prevalence of IBD is increasing with approximately 4.9 million cases reported worldwide. Current therapies are limited due to the severity of side effects and long-term toxicity, therefore, the development of novel IBD treatments is necessitated. Recent findings support apurinic/apyrimidinic endonuclease 1/reduction-oxidation factor 1 (APE1/Ref-1) as a target in many pathological conditions, including inflammatory diseases, where APE1/Ref-1 regulation of crucial transcription factors impacts significant pathways. Thus, a potential target for a novel IBD therapy is the redox activity of the multifunctional protein APE1/Ref-1. This review elaborates on the status of conventional IBD treatments, the role of an APE1/Ref-1 in intestinal inflammation, and the potential of a small molecule inhibitor of APE1/Ref-1 redox activity to modulate inflammation, oxidative stress response, and enteric neuronal damage in IBD.

Keywords: apurinic/apyrimidinic endonuclease 1/reduction-oxidation factor 1 (APE1/Ref-1); redox signaling; inflammatory bowel disease (IBD); inflammation; oxidative stress



Citation: Sahakian, L.; Robinson, A.M.; Sahakian, L.; Stavely, R.; Kelley, M.R.; Nurgali, K. APE1/Ref-1 as a Therapeutic Target for Inflammatory Bowel Disease. *Biomolecules* **2023**, *13*, 1569. <https://doi.org/10.3390/biom13111569>

Academic Editor: Jaime Belkind-Gerson

Received: 30 September 2023

Revised: 18 October 2023

Accepted: 22 October 2023

Published: 24 October 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Inflammatory bowel disease (IBD) collectively describes two debilitating conditions, ulcerative colitis (UC) and Crohn's disease (CD), characterized by chronic, relapsing, and remitting inflammation within the gastrointestinal (GI) tract [1]. While CD and UC are distinguishable by disease location within the GI tract, the nature of histopathological alterations in the intestinal wall, and associated complications, there is some overlap in clinical and pathological manifestations, such as GI disruptions, damage to the enteric nervous system (ENS), and modifications in immune responses [2]. UC is distinguished by diffuse continuous inflammation confined to the rectal and colonic mucosa and submucosa. Typically, the inflammatory reaction initiates within the mucosal and submucosal lining of the rectum, impacting either the rectum exclusively or extending proximally to involve all or part of the colon [3]. In CD, inflammation is segmented, transmural, and focal, most commonly manifesting within the colon and terminal ileum, but may affect any part of the GI tract [4]. Clinical symptoms of IBD include diarrhea, weight loss, fatigue, abdominal pain, perianal fissures, bloody stool, and tenesmus [5,6]. Disease progression may lead to

complications including fistulas, strictures, abscesses (CD), rectal bleeding (UC), and an increased risk of colorectal cancer [4,7].

The prevalence and incidence of IBD are increasing with approximately 4.9 million cases worldwide in 2019 [8]. Although not commonly associated with mortality, IBD affects significant morbidity, severely impacting patient quality of life and placing a substantial burden on global economic and healthcare systems already fraught with delivering efficient care and access [8].

Considerable progress in understanding the pathogenesis of IBD has been made in recent years; however, the precise etiology remains unknown. Accumulating evidence suggests that IBD is a heterogeneous disorder of multifactorial etiology involving complex interactions between genetic predisposition, gut microbiota dysbiosis, immune response, and environmental factors, such as cigarette smoking, stress, diet, medications, appendectomy, or pancreatitis [9]. Advances toward understanding IBD etiology have enabled the development of diagnostic techniques and treatment strategies to improve disease severity and patient outcomes [10]. However, further investigation into the disease mechanisms and novel therapies are necessitated to achieve long-term therapeutic success.

1.1. ENS and IBD

Substantial bidirectional communication between the enteric neurons and immune cells has been established in both healthy and disease states, emphasizing the significance of ENS in GI immunity [11–14]. The ENS comprises an interconnected network of enteric neurons and glial cells within the GI tract projecting towards effector structures, such as smooth muscular layers, immune cells, and blood vessels [15]. Crosstalk between the ENS and immune cells results in the production and release of immune and neural mediators, such as cytokines, chemokines, and neurotransmitters [12,16,17].

Previous studies have demonstrated immune infiltration, gross morphological changes, and enteric neuronal loss in tissues from IBD patients and animal models of chronic colitis [18–24]. Inflammation-induced changes to enteric neuronal structure and function are associated with the development of IBD symptoms, such as diarrhea and/or constipation, heightened sensitivity, and pain, that persist beyond the resolution of active inflammation [25–27]. In addition, a compromised GI antioxidant capacity has been associated with inflammation-induced ENS damage [28]. The underlying mechanisms responsible for these effects primarily involve substantial alterations in neurologically regulated processes, encompassing intestinal motility and secretion. While it is widely regarded that ENS abnormalities emerge secondary to inflammation, the occurrence of anomalies in non-inflamed areas of the gut suggests that the ENS may also play a role in IBD pathogenesis [29].

It is evident that the role of neuroimmune interactions in inflammatory conditions is critical for prolonging remission, rendering the ENS an ideal target for the development of novel therapies to remedy the symptoms and underlying pathophysiology of IBD [30].

1.2. Current Treatments for IBD

Since the etiology of IBD is yet to be fully elucidated, available therapies are designed to alleviate symptoms, mitigate complications, and control acute mucosal inflammation to prolong remission, rather than target underlying pathological mechanisms [31]. Conventional medical treatments include 5-aminosalicylic acid (5-ASA)-based anti-inflammatories, corticosteroids, immunomodulators, antibiotics, biological therapies using antibodies, and targeting the gut microbiome [31,32].

For patients with mild to moderate UC, 5-ASA-based treatments, i.e., sulfasalazine and mesalazine, are the first line of treatment demonstrated as safe and effective to induce remission and prevent relapse [31]. However, 5-ASAs have been associated with exacerbation of IBD symptoms and serious side effects, such as pancreatitis, pleuritis, myocarditis, and nephritis [33,34].

Corticosteroids have been prescribed for IBD flare-ups for decades due to their broad-spectrum anti-inflammatory capability and are subsequently the recommended treatment

for induction of remission in UC patients who do not respond to 5-ASA, as well as in patients with mild to moderate CD [35]. While highly effective for inducing remission, corticosteroids are ineffective for the maintenance of remission due to toxicity effects, contribution to major infection, and a loss of response over time [31,36,37].

Immunomodulators, including thiopurines (azathioprine), are the mainstay treatment for the treatment of moderate to severe CD and active UC where 5-ASA therapy has failed [31]. The complex pathway by which the immunosuppressive anti-metabolite azathioprine exerts its anti-inflammatory effects results in the inhibition of protein synthesis in lymphocytes [38]. Dosage is dependent on the patient's levels of enzyme serum thiopurine S-methyltransferase (TPMT); when TPMT is low, the risk of myelosuppression, non-melanoma skin cancer, and non-Hodgkin's lymphoma is enhanced [37]. Thus, the toxicity of thiopurine therapy is highly variable and unpredictable among individuals. While the efficacy of immunomodulators to maintain remission has been established, long-term use is associated with an increased rate of infection and risk of cancer [37,39]. Additionally, the onset of action for anti-metabolites is slow, which is not ideal given these drugs are most effective when the IBD is moderate to severe [40].

The use of biological therapies, such as anti-tumor necrosis factor (TNF) monoclonal antibodies, adalimumab, and infliximab, to induce and maintain remission in IBD patients has increased considerably in past years [31]. Although highly efficacious for moderate to severe UC and CD, most patients either do not respond to initial treatment or lose responsiveness over time [41]. Furthermore, biologic therapies have been associated with reactivating serious infections such as tuberculosis and hepatitis B, and an increased risk for lymphoma and non-melanoma skin cancer [31,37].

Dysbiosis of the gut microbiota has been implicated in the pathogenesis of IBD, leading to the development of strategies targeting microbial composition and modulation [42,43]. The gut microbiome consists of the community of microorganisms within the GI tract, including bacteria, archaea, viruses, and fungi [44]. Microbial composition continuously develops with influences from the environment, diet, age, hygiene, infections, and antibiotic usage [45]. This community of microbiota is the result of a mutually beneficial and harmonious relationship between the microorganisms, often referred to as symbiosis. In turn, the term dysbiosis is used to describe a shift in the balance of microorganisms, interrupting this mutually beneficial relationship, and has been linked to the pathophysiology of IBD [46]. In addition to the dysbiosis, damage to the intestinal epithelial barrier or mucus layer allows bacterial invasion into the lamina propria, promoting the inflammatory response observed in the tissues of IBD patients [44]. Methods for analyzing the gut microbiome in IBD include tissue biopsies, surgical sections, and fecal analysis [47–49]. From many of these studies, the changes in microbial composition conflict with one another due to several factors, such as antibiotic usage, age, gender, tobacco smoking, IBD type, analysis method, and GI tract location [49–51]. Current treatments that target microbial dysbiosis include probiotics, prebiotics, fecal microbiota transplantation (FMT), dietary interventions, antibiotics, and microbiome profiling. Treatments, such as FMT, constitute ethical and practical challenges, such as donor screening [52]. The response to microbiome-targeting treatments can vary due to the heterogeneity of IBD. Furthermore, the diversity of the microbiome composition among IBD patients adds a further level of complexity when treating the disease [53].

It is evident that the efficiency of current conventional IBD therapies is limited by the severity of side effects, loss of patient responsiveness, long-term toxicity, and/or failure to induce and maintain remission [54]. Therefore, the development of novel IBD therapies targeting pathophysiological mechanisms and pathways that are precise, efficacious, and reduce the occurrence of off-target effects is crucial.

1.3. Oxidative Stress and IBD

Oxidative stress has been implicated with the development and progression of IBD, therefore, redox-targeted therapy is a promising option for treatment [55]. Reactive oxy-

gen species (ROS) are mostly generated as by-products of oxidative metabolism during mitochondrial respiration, as well as in cellular response to xenobiotics, cytokines, and bacterial invasion [56]. Aerobic organisms develop a comprehensive endogenous antioxidant defense system to maintain cellular redox homeostasis [57]; oxidative stress refers to the state of imbalance between ROS production and the capacity of the antioxidant defense system to mount an effective response, in favor of oxidants [58]. Modulation of intracellular levels of ROS is vital for cellular homeostasis as different ROS levels produce contrasting biological responses. ROS functions as second messenger, signaling molecules to regulate cellular physiological and biological processes when produced at low to moderate concentrations. However, excessive amounts of ROS overwhelm the antioxidant defense system and result in redox imbalances disrupting cellular integrity and functions, including damage to cellular and mitochondrial lipids, proteins, and DNA [59,60]. Production of ROS by mitochondria is critical as it underlies oxidative damage in many pathologies, including inflammatory, cardiovascular, carcinogenic, autoimmune, and neurological degenerative diseases [60,61].

Previous studies have reported elevated levels of biomarkers for oxidative stress and ROS-mediated damage, together with reduced antioxidant levels in IBD patients and animal models of colitis [62–66]. It is indicated that an oxidative stress-induced exaggerated inflammatory response alters epithelial barrier permeability allowing luminal pathogen invasion and leukocyte infiltration [67]. Therefore, the cumulative damage results in intestinal mucosal necrosis and ulceration associated with IBD [68]. Increased expression and activation of nuclear factor kappa B (NF- κ B) has been detected in the colonic mucosa of patients with IBD [69,70]. Under physiological conditions, ROS-activated intracellular signaling pathways, such as NF- κ B, are recognized to play a role in the maintenance of the intestinal epithelial barrier function and coordination of the epithelial immune response in microorganisms [71,72]. However, oxidative activation of NF- κ B and activator protein-1 (AP-1) signaling stimulates expression of pro-inflammatory cytokines in intestinal epithelial cells, including tumor necrosis factor alpha (TNF- α), interleukin (IL)-1, IL-6, IL-8, and cyclooxygenases-2 (COX-2) [73]. These cytokines are associated with exacerbating existing inflammation and promoting carcinogenesis in IBD [74]. Therefore, given that persistent oxidative stress is considered to play a role in the pathogenesis and progression of IBD, it is indicative that sustained NF- κ B signaling stimulated by excessive ROS exaggerates the chronic intestinal inflammation in the mucosa of IBD patients.

In IBD, there is an abnormal immune response against the gut microbiome [75]. The mucosa provides a habitat for the microbiome, and in turn, the microbiome influences the health and function of the mucosa. Thus, there is a dynamic and bidirectional relationship between the mucosa and microbiome [76]. The composition of gut microbial communities is influenced by intestinal oxygenation. Healthy intestines are characterized by low oxygen levels and large bacterial communities of obligate anaerobes, whereas in IBD, chronic inflammation results in increased release of hemoglobin carrying oxygen and reactive oxygen species into the intestinal lumen, creating a microenvironment that favors facultative anaerobes [77–79]. The subsequent decrease in obligate anaerobes that release anti-inflammatory compounds initiates an increased inflammatory response, inaugurating a positive feedback loop that accelerates the disease process [79–81]. Therefore, oxidative stress and microbial dysbiosis are interconnected; the ROS generated during inflammation can directly compromise the integrity of the epithelial barrier, stimulating an immune response and triggering the production of microbial metabolites.

Oxidative stress and chronic inflammation intertwine as key pathologic factors contributing to enteric neuropathy [82,83]. The effects of oxidative stress on the ENS can be cumulative, affecting structural and functional changes [84]. Previous studies report a significant loss of enteric neurons, but an increased proportion of neuronal nitric oxide synthase (nNOS) neurons in the myenteric plexus in colonic tissues from IBD patients and experimental models of colitis [83,85–88]. Primarily inhibitory motor neurons, nNOS neurons are responsible for the relaxation of the intestinal smooth muscle cells, and altered

chemical coding of enteric neuronal subpopulations is associated with impaired smooth muscle contractility and intestinal dysmotility [89]. Changes in the size and proportion of nNOS neurons have been associated with oxidative stress indicating that sensitivity of colonic tissue to redox imbalance may arise from inadequacy of antioxidant defense systems under pathological conditions [84,90].

Colon biopsies from IBD patients and animal models of colitis have shown increased levels of nitric oxide (NO) [91,92] and 8-hydroxydeoxyguanosine (8-OHdG), a common index for examining oxidative damage to DNA [93,94]. Furthermore, 8-OHdG levels are reported to be permanently elevated in CD patients, independent of disease activity [94]. There is only one study that has evaluated the presence of 8-OHdG in myenteric neurons [19]. Results of this study demonstrated increased levels of 8-OHdG identifying oxidative stress-induced DNA damage within myenteric neurons in colons from *Winnie* mice, a spontaneous model of chronic intestinal inflammation. In *Winnie* mice, a decrease in DNA damage was demonstrated after treatment with APX3330, an apurinic/aprimidinic endonuclease/redox factor-1 (APE1/Ref-1) redox domain inhibitor [19]. Furthermore, APX3330 ameliorated colonic dysmotility, altered GI transit, and enteric neuropathy, including reducing superoxide production in myenteric neurons and preventing neuronal death, while providing anti-inflammatory and antioxidant effects in *Winnie* mice. Therefore, treatment with APX3330 had prominent therapeutic effects in a preclinical animal model of IBD, providing evidence of a potential novel therapy to protect the ENS from IBD-induced injury.

As there is limited pursuance in free radical antioxidants for the treatment of IBD, new therapeutic strategies should aim to obstruct the major sources of oxidative stress contributing to enteric neuropathy.

2. APE1/Ref-1

An essential regulator of cellular response to oxidative stress, APE1/Ref-1 is a dual-functioning protein with major roles in DNA repair and redox signaling [95,96]. The subcellular localization of APE1/Ref-1 is predominantly in the nucleus; however, it is also found in the mitochondria and other areas of the cytoplasm [97–99]. APE1/Ref-1 functions as the main apurinic/aprimidinic endonuclease in the base excision repair (BER) pathway, accounting for 95% of abasic site repairs [100]. The BER pathway repairs DNA damaged by oxidation, deamination, and alkylation to prevent mutagenesis and promote cellular survival. Given the vulnerability of neurons to ROS and altered BER function in neurological disorders; repair of oxidative DNA damage by efficient BER activity is considered protective against neurodegeneration [101].

In its role as a redox signaling protein, APE1/Ref-1 acts via redox-dependent mediation of DNA binding of transcription factors, involved in inflammatory regulation, immune response, angiogenesis, and cell survival [96,102]. Several studies have demonstrated enhanced DNA binding activity for AP-1, hypoxia-inducible factor-1 (HIF-1), NF- κ B, p53, and STAT3 to be associated with APE1/Ref-1 [103–108]. The capacity of APE1/Ref-1 to maintain transcription factors in an active reduced state subsequently affects the modulation of the expression of genes involved in oxidative stress and maintains genomic stability [96]. Silencing of APE1/Ref-1 via siRNA increases intracellular ROS production [109,110]. Correspondingly, blocking APE1/Ref-1 redox activity reduces the DNA binding activity of AP-1, HIF-1 α , and NF- κ B [111,112]. Cells are more susceptible to oxidative stress when expression of APE1/Ref-1 is compromised; reduced APE1/Ref-1 is associated with increased cytotoxicity of neurons, impacting cell survival and driving apoptosis [113].

On the other hand, APE1/Ref-1 mediation of fundamental transcription factors that control cell cycle arrest and apoptotic programs can promote the growth, migration, and survival of cancer cells, as well as angiogenesis in the tumor microenvironment [95]. Overexpression in cancer cells and irregular cytoplasmic versus nuclear distribution of APE1/Ref-1 is associated with tumor aggressiveness and poorer patient prognosis in many tumor types, including colorectal cancer [114,115].

The dual functions of APE1/Ref-1 are molecularly distinct and completely independent in their function [116]. The apurinic/apyrimidinic endonuclease activity of APE1/Ref-1 is critical for maintaining genomic stability and cellular existence, evidenced by the embryonic lethality of murine APE1/Ref-1 knockout models and the inability to establish viable cell lines completely deficient for APE1 [96]. APE1/Ref-1 redox function is significant for cellular response to oxidative stress, however, the implications of cancer development and progression have motivated recent investigations into inhibition of the redox activity of APE/Ref-1 [19,96,112]. Compounds that specifically inhibit the redox function of APE1/Ref-1 do not obstruct DNA repair activity and previous studies suggest that APE1/Ref-1 redox inhibitors actually augment the endonuclease repair activity of APE1/Ref-1 in the hippocampus, sensory, and enteric neurons, thus providing a neuroprotective effect [19,117,118].

APE1/Ref-1 in Intestinal Inflammation

Oxidative stress is implicated in inflammatory conditions resulting in altered APE1/Ref-1 response; without intercept of redox activation, cell apoptosis and carcinogenesis are likely [119]. Elevated APE1/Ref-1 expression has been reported in tissues from IBD patients with active inflammation and in an animal model of colitis, signifying underlying oxidative stress within the gut [62,63,120]. Furthermore, increased expression of APE1/Ref-1 in colon sections from UC patients has been associated with an enhanced pro-inflammatory response, which is a precursor for colorectal cancer (CRC) susceptibility [63,120].

In intestinal epithelial cells, APE1/Ref-1 plays a major role in controlling the onset of oxidative stress-based inflammatory processes by modulating NF- κ B-mediated IL-8 gene expression [121]. Studies have shown enhanced binding and IL-8 promoter activity in colitis and cancer where APE1/Ref-1 is overexpressed [120,122]. Silencing APE1/Ref-1 in gastric epithelial cells infected with *Helicobacter Pylori* inhibited IL-8 expression [123]. These findings illustrate the importance of APE1/Ref-1 in IL-8 regulation.

In addition to IL-8, IL-6 also has a crucial role in the development and progression of uncontrolled intestinal inflammatory processes characteristic of IBD [124]. Secretory APE/Ref-1 induces IL-6 expression in response to inflammatory challenges, such as lipopolysaccharides (LPS) and TNF- α stimulation, and increased IL-6 expression enhances excessive APE1/Ref-1 secretion in a feedforward loop [125]. Furthermore, silencing of APE1 expression by siRNA decreases the release of IL-6 [126]. Expression and activation of STAT3 are increased in intestinal epithelial cells during active IBD [127]. Given that IL-6 activates STAT3 and constitutively activated STAT3 augments APE1/Ref-1 expression, an underlying role of APE1/Ref-1 in the pathogenesis of GI inflammation is indicated [128].

Increased expression of APE1 in IBD is positively correlated with microsatellite instability (MSI) [63]. In chronic inflammatory diseases such as IBD, MSI is affected by the overproduction of free radicals saturating the ability of the cell to repair DNA damage preceding replication [129,130]. Prolonged intracellular stress induced by chronic inflammation and associated increases in APE1 lead to cumulative genomic instability as the endonuclease activity of APE1 produces cytotoxic DNA repair intermediates [131]. It is, therefore, considered that chronic inflammation in IBD causes adaptive increases in APE1 levels, which paradoxically enhance MSI in affected tissues, and enzymes that normally facilitate DNA repair are instead causing mutations [132]. Chromosomal instabilities have been detected in dysplastic tissues from patients in the early stages of IBD [133]. These instabilities contribute to gene mutations leading to colon carcinogenesis and a 20–30-fold increased risk of developing colorectal cancer (CRC) [134].

Despite considerable evidence supporting enteric dysregulation in IBD, the effect of oxidative stress and the mechanistic role of APE1/Ref-1 within the ENS has not fully been elucidated [83,86–89]. Specific inhibition of APE1/Ref-1 redox function has shown promising effects in preventing enteric neuropathy and alleviating intestinal inflammation in a murine model of spontaneous chronic colitis [19]. Hence, the APE1/Ref-1 redox domain inhibitor APX3330 provides an opportunity to target specific redox mechanisms of

the oxidative stress response associated with intestinal inflammation and provide further understanding of its role in IBD pathogenesis.

3. Targeting APE1/Ref-1 as a Therapeutic Approach

Currently branded as APX3330, (E)-3-(5,6-dimethoxy-3-methyl-1,4-dioxocyclohexa-2,5-dienyl)-2-nonylpropenoic acid (E3330) has been synthesized as a known small-molecular APE1/Ref-1 redox function inhibitor [135]. APX3330 has shown efficacy and safety in clinical trials for oncology and diabetic retinopathy [136,137]. The use of APX3330 facilitates endeavors in further understanding of APE1/Ref-1 redox interactions and isolation, providing an opportunity for specific therapeutics targeting pathophysiological mechanisms of IBD. APX3330 is a highly selective inhibitor of Ref-1 redox activity specifically binding to sites located on the APE1/Ref-1 redox domain while leaving the DNA repair domain untouched [138–141]. The specific inhibition activity of APX3330 blocks Ref-1's ability to convert transcription factors from their oxidized, inactive state to an active, reduced state. APX3330 has been shown to suppress the production of pro-inflammatory cytokines and inflammatory mediators in murine macrophages resulting in the inability of NF- κ B and AP-1 to bind to their target DNA sequence [122]. The inhibition activity of APX3330 downregulates secretion of the inflammatory cytokines IL-6 and IL-12, and the inflammatory mediators PGE2 and NO, as well as expression of COX-2 and iNOS which regulate the production of PGE2 and NO [122]. Moreover, APX3330 has been demonstrated to inhibit HIF-1 DNA binding activity under the same mechanisms that HIF-1 regulates APE1/Ref-1 expression; although it is itself regulated by APE1/Ref-1 [140,142,143]. Inhibition of HIF-1 and NF κ B function in patients treated with APX3330 resulted in the suppression of genes downstream of these transcription factors in a cancer clinical trial [144]. In a murine model of spontaneous chronic colitis, APX3330 affected repair of inflammation-induced ENS damage, and ameliorated IBD symptoms, in addition to demonstrated immunomodulatory function [19]. Thus, targeting the specific inhibition of APE1/Ref-1 redox pathways while preserving the DNA repair pathway in intestinal inflammation is promising and a potential novel treatment for IBD and its associated enteric neuropathy (Figure 1).

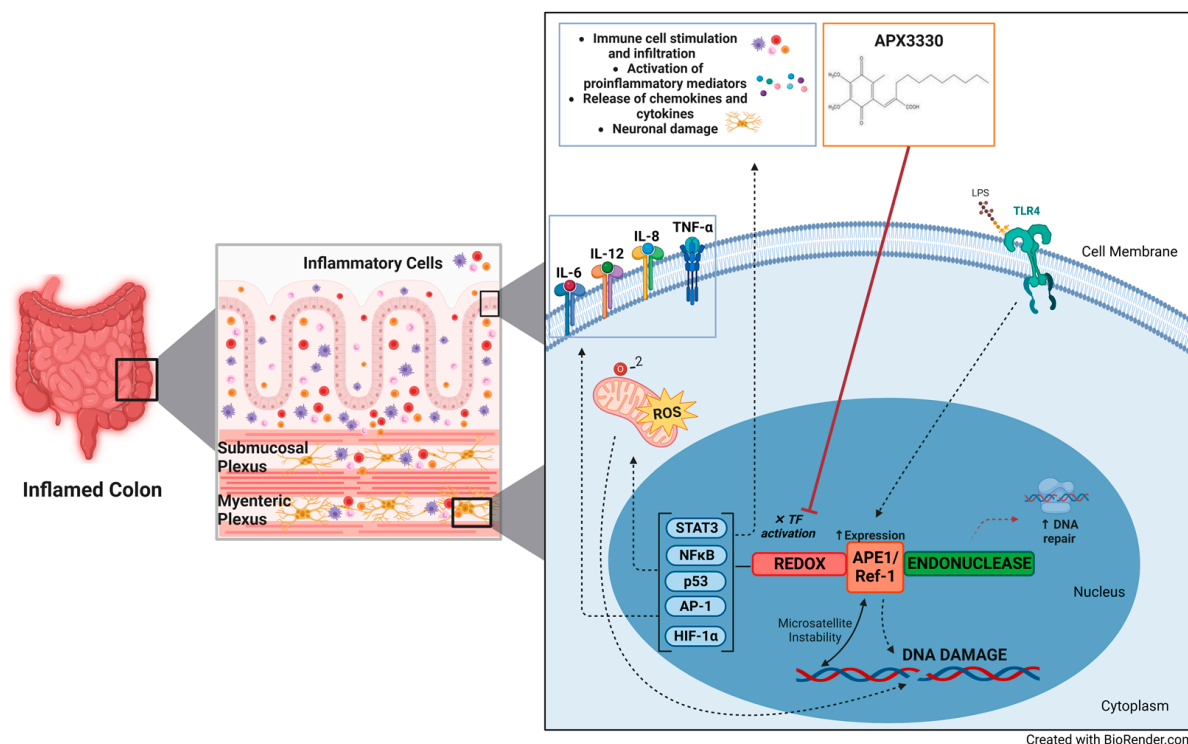


Figure 1. APE1/Ref-1 role in inflammatory bowel disease and the potential of redox signaling inhibition. The activation of transcription factors through the redox function of APE1 is implicated in

the promotion of inflammation in inflammatory bowel disease. Furthermore, the subsequent activation of downstream mediators directly affects the inflammatory responses. By inhibiting the redox signaling pathway using APX compounds, the excessive production of pro-inflammatory cytokines, chemokines, reactive oxygen species (ROS), and neuronal damage can be reduced, leading to the alleviation of inflammation and gastrointestinal functions. Additionally, the repair function of APE1 plays a crucial role in mending the DNA or RNA lesions caused by inflammation and oxidative stress. In summary, the utilization of APX compounds can effectively diminish inflammatory markers, ROS levels, and enteric neuropathy, while also enhancing the DNA repair function, thereby presenting a novel therapeutic opportunity for IBD (created with BioRender.com (accessed on 30 September 2023)).

Abbreviations: TFs: transcription factors; IL: interleukin; TNF- α : tumor necrosis factor-alpha; ROS: reactive oxygen species; STAT3: signal transducer and activator of transcription 3; NF κ B: nuclear factor kappa-light chain enhancer of activated B cells; AP-1: activator protein 1; HIF1 α : hypoxia-inducible factor 1 subunit alpha; redox: reduction-oxidation; LPS: lipopolysaccharide; TLR4: toll-like receptor 4; DNA: deoxyribonucleic acid.

4. Conclusions and Future Directions

Previous studies have demonstrated that APE1/Ref-1 simultaneously affects inflammation and oxidative stress. Although the effects of APE1/Ref-1 on diverse transcriptional targets have been extensively examined in various types of cancers and diseases, its impact on other transcriptional targets within the context of IBD remains unexplored. In addition, investigations elucidating the role of APE1/Ref-1 in inflammation-induced enteric neuropathy are warranted. Furthermore, the overall range of effects of APE1/Ref-1 redox inhibitors in IBD is currently unknown. Thus, further endeavors are imperative to comprehend the mechanism of APE1/Ref-1 and its redox inhibitors in chronic intestinal inflammation.

APE1/Ref-1 redox inhibitors provide a novel therapeutic strategy for managing colitis compared to conventional treatments. They present distinct advantages over current drugs for treating colitis by not only targeting inflammation and oxidative stress but also repairing DNA damage and are oral agents. The benefit of APE1/Ref-1 inhibitors as a prospective remedy for IBD lies in their minimal systemic adverse effects reported in clinical trials in cancer and diabetic retinopathy patients. Given the significance of the redox-regulated transcriptional control of APE1/Ref-1 in intestinal inflammation, targeting APE1/Ref-1 with APE1/Ref-1 redox inhibitors offers a novel treatment approach for IBD that has the potential to circumvent the challenges associated with current therapies and improve disease outcomes in many patients.

Author Contributions: Conceptualization, K.N. and M.R.K.; writing—original draft preparation, L.S. (Lauren Sahakian); writing—review and editing, A.M.R., L.S. (Linda Sahakian), R.S., M.R.K., and K.N.; supervision, K.N.; project administration, K.N. All authors have read and agreed to the published version of the manuscript.

Funding: This study is supported by the Crohn's & Colitis Foundation Senior Research Award (K.N., Award number: 903433).

Data Availability Statement: No new data were created.

Conflicts of Interest: M.R.K. has licensed APX3330 through Indiana University Research and Technology Corporation to Apexian Pharmaceuticals LLC. M.R.K. is CSO of Apexian Pharmaceuticals and a medical consultant for Ocuphire Pharma who licensed APX3330 from Apexian for eye diseases. Apexian Pharmaceutical nor Ocuphire Pharma had neither control nor oversight of the interpretation or presentation of anything in this manuscript. Other authors do not have any conflicts of interest.

References

1. Baumgart, D.C.; Sandborn, W.J. Inflammatory bowel disease: Clinical aspects and established and evolving therapies. *Lancet* **2007**, *369*, 1641–1657. [[CrossRef](#)]
2. Matricon, J.; Barnich, N.; Ardid, D. Immunopathogenesis of inflammatory bowel disease. *Self Nonself* **2010**, *1*, 299–309. [[CrossRef](#)] [[PubMed](#)]
3. Ungaro, R.; Mehandru, S.; Allen, P.B.; Peyrin-Biroulet, L.; Colombel, J.-F. Ulcerative colitis. *Lancet* **2017**, *389*, 1756–1770. [[CrossRef](#)] [[PubMed](#)]
4. Torres, J.; Mehandru, S.; Colombel, J.-F.; Peyrin-Biroulet, L. Crohn's disease. *Lancet* **2017**, *389*, 1741–1755. [[CrossRef](#)] [[PubMed](#)]
5. Strober, W.; Fuss, I.; Mannon, P. The fundamental basis of inflammatory bowel disease. *J. Clin. Investig.* **2007**, *117*, 514–521. [[CrossRef](#)]
6. Subasinghe, D.; Nawarathna, N.M.M.; Samarasekera, D.N. Disease characteristics of inflammatory bowel disease (IBD). *J. Gastrointest. Surg.* **2011**, *15*, 1562–1567. [[CrossRef](#)]
7. Tan, Z.; Zhu, S.; Liu, C.; Meng, Y.; Li, J.; Zhang, J.; Dong, W. Causal link between inflammatory bowel disease and fistula: Evidence from mendelian randomization study. *J. Clin. Med.* **2023**, *12*, 2482. [[CrossRef](#)]
8. Wang, R.; Li, Z.; Liu, S.; Zhang, D. Global, regional and national burden of inflammatory bowel disease in 204 countries and territories from 1990 to 2019: A systematic analysis based on the Global Burden of Disease Study 2019. *Br. Med. J.* **2023**, *13*, 65–138. [[CrossRef](#)]
9. Ananthakrishnan, A.N. Epidemiology and risk factors for ibd. *Nat. Rev. Gastroenterol. Hepatol.* **2015**, *12*, 205–217. [[CrossRef](#)] [[PubMed](#)]
10. Muzammil, M.A.; Fariha, F.; Patel, T.; Sohail, R.; Kumar, M.; Khan, E.; Khanam, B.; Kumar, S.; Khatri, M.; Varrassi, G.; et al. Advancements in inflammatory bowel disease: A narrative review of diagnostics, management, epidemiology, prevalence, patient outcomes, quality of life, and clinical presentation. *Cureus* **2023**, *15*, 411–420. [[CrossRef](#)]
11. Knowles, C.H.; Lindberg, G.; Panza, E.; Giorgio, R.D. New perspectives in the diagnosis and management of enteric neuropathies. *Nat. Rev. Gastroenterol. Hepatol.* **2013**, *10*, 206. [[CrossRef](#)] [[PubMed](#)]
12. Yoo, B.B.; Mazmanian, S.K. The enteric network: Interactions between the immune and nervous systems of the gut. *Immunity* **2017**, *46*, 910–926. [[CrossRef](#)]
13. Chesné, J.; Cardoso, V.; Veiga-Fernandes, H. Neuro-immune regulation of mucosal physiology. *Mucosal Immunol.* **2019**, *12*, 10–20. [[CrossRef](#)]
14. Verheijden, S.; Boeckstaens, G.E. Neuroimmune interaction and the regulation of intestinal immune homeostasis. *Am. J. Physiol.-Gastrointest. Liver Physiol.* **2018**, *314*, 75–80. [[CrossRef](#)] [[PubMed](#)]
15. Hansen, M.B. The enteric nervous system I: Organisation and classification. *Pharmacol. Toxicol.* **2003**, *92*, 105–113. [[CrossRef](#)]
16. Wang, H.; Foong, J.P.P.; Harris, N.L.; Bornstein, J.C. Enteric neuroimmune interactions coordinate intestinal responses in health and disease. *Mucosal Immunol.* **2022**, *15*, 27–39. [[CrossRef](#)] [[PubMed](#)]
17. Jacobson, A.; Yang, D.; Vella, M.; Chiu, I.M. The intestinal neuro-immune axis: Crosstalk between neurons, immune cells, and microbes. *Mucosal Immunol.* **2021**, *14*, 555–565. [[CrossRef](#)] [[PubMed](#)]
18. Rivera, L.R.; Poole, D.P.; Thacker, M.; Furness, J.B. The involvement of nitric oxide synthase neurons in enteric neuropathies. *Neurogastroenterol. Motil.* **2011**, *23*, 980–988. [[CrossRef](#)]
19. Sahakian, L.; Filippone, R.T.; Stavely, R.; Robinson, A.M.; Yan, X.S.; Abalo, R.; Eri, R.; Bornstein, J.C.; Kelley, M.R.; Nurgali, K. Inhibition of APE1/Ref-1 redox signaling alleviates intestinal dysfunction and damage to myenteric neurons in a mouse model of spontaneous chronic colitis. *Inflamm. Bowel Dis.* **2021**, *27*, 388–406. [[CrossRef](#)] [[PubMed](#)]
20. Robinson, A.M.; Rahman, A.A.; Carbone, S.E.; Randall-Demllo, S.; Filippone, R.; Bornstein, J.C.; Eri, R.; Nurgali, K. Alterations of colonic function in the Winnie mouse model of spontaneous chronic colitis. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2017**, *312*, 85–102. [[CrossRef](#)]
21. Robinson, A.M.; Stavely, R.; Miller, S.; Eri, R.; Nurgali, K. Mesenchymal stem cell treatment for enteric neuropathy in the Winnie mouse model of spontaneous chronic colitis. *Cell Tissue Res.* **2022**, *389*, 41–70. [[CrossRef](#)]
22. Oehmichen, M.; Reifferscheid, P. Intramural ganglion cell degeneration in inflammatory bowel disease. *Digestion* **1977**, *15*, 482–496. [[CrossRef](#)] [[PubMed](#)]
23. Bernardini, N.; Segnani, C.; Ippolito, C.; De Giorgio, R.; Colucci, R.; Fausone-Pellegrini, M.S.; Chiarugi, M.; Campani, D.; Castagna, M.; Mattii, L.; et al. Immunohistochemical analysis of myenteric ganglia and interstitial cells of Cajal in ulcerative colitis. *J. Cell. Mol. Med.* **2012**, *16*, 318–327. [[CrossRef](#)]
24. Riemann, J.F.; Schmidt, H. Ultrastructural changes in the gut autonomic nervous system following laxative abuse and in other conditions. *Scand. J. Gastroenterol.* **1982**, *71*, 111–124.
25. Krauter, E.M.; Strong, D.S.; Brooks, E.M.; Linden, D.R.; Sharkey, K.A.; Mawe, G.M. Changes in colonic motility and the electrophysiological properties of myenteric neurons persist following recovery from trinitrobenzene sulfonic acid colitis in the guinea pig. *Neurogastroenterol. Motil.* **2007**, *19*, 990–1000. [[CrossRef](#)] [[PubMed](#)]
26. Lakhan, S.E.; Kirchgessner, A. Neuroinflammation in inflammatory bowel disease. *J. Neuroinflammation* **2010**, *7*, 37. [[CrossRef](#)]
27. Lomax, A.E.; O'Hara, J.R.; Hyland, N.P.; Mawe, G.M.; Sharkey, K.A. Persistent alterations to enteric neural signaling in the guinea pig colon following the resolution of colitis. *Am. J. Physiol.-Gastrointest. Liver Physiol.* **2007**, *292*, 482–491. [[CrossRef](#)] [[PubMed](#)]

28. Brown, I.A.M.; Gulbransen, B.D. The antioxidant glutathione protects against enteric neuron death in situ, but its depletion is protective during colitis. *Am. J. Physiol.-Gastrointest. Liver Physiol.* **2017**, *314*, G39–G52. [[CrossRef](#)]
29. Villanacci, V.; Bassotti, G.; Nascimbeni, R.; Antonelli, E.; Cadei, M.; Fisogni, S. Enteric nervous system abnormalities in inflammatory bowel diseases. *Neurogastroenterol. Motil.* **2008**, *20*, 1009–1016. [[CrossRef](#)] [[PubMed](#)]
30. Nezami, B.G.; Srinivasan, S. Enteric nervous system in the small intestine: Pathophysiology and clinical implications. *Curr. Gastroenterol. Rep.* **2010**, *12*, 358–365. [[CrossRef](#)] [[PubMed](#)]
31. Wright, E.K.; Ding, N.S.; Niewiadomski, O. Management of Inflammatory Bowel Disease. *Med. J. Aust.* **2018**, *209*, 318–323. [[CrossRef](#)]
32. Mijan, M.A.; Lim, B.O. Diets, functional foods, and nutraceuticals as alternative therapies for inflammatory bowel disease: Present status and future trends. *World J. Gastroenterol.* **2018**, *24*, 2673–2685. [[CrossRef](#)] [[PubMed](#)]
33. Williams, C.; Panaccione, R.; Ghosh, S.; Rioux, K. Optimizing clinical use of mesalazine (5-aminosalicylic acid) in inflammatory bowel disease. *Ther. Adv. Gastroenterol.* **2011**, *4*, 237–248. [[CrossRef](#)] [[PubMed](#)]
34. Miyoshi, J.; Matsuoka, K.; Yoshida, A.; Naganuma, M.; Hisamatsu, T.; Yajima, T.; Inoue, N.; Okamoto, S.; Iwao, Y.; Ogata, H.; et al. 5-Aminosalicylic acid aggravates colitis mimicking exacerbation of ulcerative colitis. *Intest. Res.* **2018**, *16*, 635–640. [[CrossRef](#)]
35. Barrett, K.; Saxena, S.; Pollok, R. Using corticosteroids appropriately in inflammatory bowel disease: A guide for primary care. *Br. J. Gen. Pract.* **2018**, *68*, 497–498. [[CrossRef](#)] [[PubMed](#)]
36. Harlan, W.R.; Meyer, A.; Fisher, J. Inflammatory bowel disease: Epidemiology, evaluation, treatment, and health maintenance. *North. Carol. Med. J.* **2016**, *77*, 198–201. [[CrossRef](#)]
37. Bernstein, C.N. Treatment of IBD: Where we are and where we are going. *Am. J. Gastroenterol.* **2014**, *110*, 114. [[CrossRef](#)] [[PubMed](#)]
38. Cheah, M.; Khanna, R. Chapter 1—Current medical therapies for ulcerative colitis. In *Pouchitis and Ileal Pouch Disorders*; Shen, B., Ed.; Elsevier: Amsterdam, The Netherlands, 2019; pp. 1–15. [[CrossRef](#)]
39. Miyatani, Y.; Kobayashi, T. Evidence-based approach to the discontinuation of immunomodulators or biologics in inflammatory bowel disease. *Digestion* **2023**, *104*, 66–73. [[CrossRef](#)]
40. Hanauer, S.B.; Sandborn, W.J.; Lichtenstein, G.R. Evolving considerations for thiopurine therapy for inflammatory bowel diseases: A clinical practice update. *Gastroenterology* **2019**, *156*, 36–42. [[CrossRef](#)] [[PubMed](#)]
41. Ben-Horin, S.; Chowers, Y. Review article: Loss of response to anti-tnf treatments in Crohn’s disease. *Aliment. Pharmacol. Ther.* **2011**, *33*, 987–995. [[CrossRef](#)] [[PubMed](#)]
42. Nishida, A.; Inoue, R.; Inatomi, O.; Bamba, S.; Naito, Y.; Andoh, A. Gut microbiota in the pathogenesis of inflammatory bowel disease. *Clin. J. Gastroenterol.* **2018**, *11*, 1–10. [[CrossRef](#)]
43. Santana, P.T.; Rosas, S.L.B.; Ribeiro, B.E.; Marinho, Y.; de Souza, H.S.P. Dysbiosis in inflammatory bowel disease: Pathogenic role and potential therapeutic targets. *Int. J. Mol. Sci.* **2022**, *23*, 3464. [[CrossRef](#)]
44. Wu, Z.A.; Wang, H.X. A Systematic Review of the Interaction Between Gut Microbiota and Host Health from a Symbiotic Perspective. *SN Compr. Clin. Med.* **2018**, *1*, 224–235. [[CrossRef](#)]
45. Dominguez-Bello, M.G.; Blaser, M.J.; Ley, R.E.; Knight, R. Development of the Human Gastrointestinal Microbiota and Insights From High-Throughput Sequencing. *Gastroenterology* **2011**, *140*, 1713–1719. [[CrossRef](#)] [[PubMed](#)]
46. Matsuoka, K.; Kanai, T. The gut microbiota and inflammatory bowel disease. *Semin. Immunopathol.* **2015**, *37*, 47–55. [[CrossRef](#)] [[PubMed](#)]
47. Frank, D.N.; St. Amand, A.L.; Feldman, R.A.; Boedeker, E.C.; Harpaz, N.; Pace, N.R. Molecular-Phylogenetic Characterization of Microbial Community Imbalances in Human Inflammatory Bowel Diseases. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 13780–13785. [[CrossRef](#)] [[PubMed](#)]
48. Manichanh, C. Reduced diversity of faecal microbiota in Crohn’s disease revealed by a metagenomic approach. *Gut* **2006**, *55*, 205–211. [[CrossRef](#)]
49. Imhann, F.; Vich Vila, A.; Bonder, M.J.; Fu, J.; Gevers, D.; Visschedijk, M.C.; Spekhorst, L.M.; Alberts, R.; Franke, L.; Van Dullemen, H.M.; et al. Interplay of host genetics and gut microbiota underlying the onset and clinical presentation of inflammatory bowel disease. *Gut* **2018**, *67*, 108–119. [[CrossRef](#)] [[PubMed](#)]
50. Wu, S.; Li, W.; Smarr, L.; Nelson, K.; Yooseph, S.; Torralba, M. Large memory high performance computing enables comparison across human gut microbiome of patients with autoimmune diseases and healthy subjects. In Proceedings of the Conference on Extreme Science and Engineering Discovery Environment: Gateway to Discovery, San Diego, CA, USA, 22–25 July 2013. [[CrossRef](#)]
51. Morgan, X.C.; Tickle, T.L.; Sokol, H.; Gevers, D.; Devaney, K.L.; Ward, D.V.; Reyes, J.A.; Shah, S.A.; Leleiko, N.; Snapper, S.B.; et al. Dysfunction of the intestinal microbiome in inflammatory bowel disease and treatment. *Genome Biol.* **2012**, *13*, R79. [[CrossRef](#)]
52. Grigoryan, Z.; Shen, M.J.; Twardus, S.W.; Beuttler, M.M.; Chen, L.A.; Bateman-House, A. Fecal microbiota transplantation: Uses, questions and ethics. *Med. Microbiol.* **2020**, *6*, 100027. [[CrossRef](#)] [[PubMed](#)]
53. Lee, M.; Chang, E.B. Inflammatory bowel diseases (IBD) and the microbiome—searching the crime scene for clues. *Gastroenterology* **2021**, *160*, 524–537. [[CrossRef](#)] [[PubMed](#)]
54. Lee, H.S.; Park, S.-K.; Park, D.I. Novel treatments for inflammatory bowel disease. *Korean J. Intern. Med.* **2018**, *33*, 20–27. [[CrossRef](#)]
55. Tian, T.; Wang, Z.; Zhang, J. Pathomechanisms of oxidative stress in inflammatory bowel disease and potential antioxidant therapies. *Oxidative Med. Cell. Longev.* **2017**, *2017*, 4535194. [[CrossRef](#)] [[PubMed](#)]

56. Ray, P.D.; Huang, B.W.; Tsuji, Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell Signal* **2012**, *24*, 981–990. [[CrossRef](#)]
57. Birben, E.; Sahiner, U.M.; Sackesen, C.; Erzurum, S.; Kalayci, O. Oxidative stress and antioxidant defense. *World Allergy Organ. J.* **2012**, *5*, 9–19. [[CrossRef](#)]
58. Shan, J.-L.; He, H.-T.; Li, M.-X.; Zhu, J.-W.; Cheng, Y.; Hu, N.; Wang, G.; Wang, D.; Yang, X.-Q.; He, Y.; et al. APE1 promotes antioxidant capacity by regulating Nrf-2 function through a redox-dependent mechanism. *Free Radic. Biol. Med.* **2015**, *78*, 11–22. [[CrossRef](#)] [[PubMed](#)]
59. Schieber, M.; Chandel, N.S. ROS function in redox signaling and oxidative stress. *Curr. Biol.* **2014**, *24*, 453–462. [[CrossRef](#)]
60. Murphy, M.P. How mitochondria produce reactive oxygen species. *Biochem. J.* **2009**, *417*, 1–13. [[CrossRef](#)]
61. Li, X.; Fang, P.; Mai, J.; Choi, E.T.; Wang, H.; Yang, X.-f. Targeting mitochondrial reactive oxygen species as novel therapy for inflammatory diseases and cancers. *J. Hematol. Oncol.* **2013**, *6*, 19. [[CrossRef](#)]
62. Lih-Brody, L.; Powell, S.R.; Collier, K.P.; Reddy, G.M.; Cerchia, R.; Kahn, E.; Weissman, G.S.; Katz, S.; Floyd, R.A.; McKinley, M.J.; et al. Increased oxidative stress and decreased antioxidant defenses in mucosa of inflammatory bowel disease. *Dig. Dis. Sci.* **1996**, *41*, 2078–2086. [[CrossRef](#)] [[PubMed](#)]
63. Hofseth, L.J.; Khan, M.A.; Ambrose, M.; Nikolayeva, O.; Xu-Welliver, M.; Kartalou, M.; Hussain, S.P.; Roth, R.B.; Zhou, X.; Mechanic, L.E.; et al. The adaptive imbalance in base excision–repair enzymes generates microsatellite instability in chronic inflammation. *J. Clin. Investig.* **2003**, *112*, 1887–1894. [[CrossRef](#)] [[PubMed](#)]
64. Bardia, A.; Tiwari, S.K.; Gunisetty, S.; Anjum, F.; Nallari, P.; Habeeb, M.A.; Khan, A.A. Functional polymorphisms in XRCC-1 and APE-1 contribute to increased apoptosis and risk of ulcerative colitis. *Inflamm. Res.* **2012**, *61*, 359–365. [[CrossRef](#)] [[PubMed](#)]
65. Rezaie, A.; Parker, R.D.; Abdollahi, M. Oxidative stress and pathogenesis of inflammatory bowel disease: An epiphenomenon or the cause? *Dig. Dis. Sci.* **2007**, *52*, 2015–2021. [[CrossRef](#)] [[PubMed](#)]
66. Chami, B.; Martin, N.J.J.; Dennis, J.M.; Witting, P.K. Myeloperoxidase in the inflamed colon: A novel target for treating inflammatory bowel disease. *Arch. Biochem. Biophys.* **2018**, *645*, 61–71. [[CrossRef](#)] [[PubMed](#)]
67. Ordás, I.; Eckmann, L.; Talamini, M.; Baumgart, D.C.; Sandborn, W.J. Ulcerative colitis. *Lancet* **2012**, *380*, 1606–1619. [[CrossRef](#)] [[PubMed](#)]
68. Oshitani, N.; Sawa, Y.; Hara, J.; Adachi, K.; Nakamura, S.; Matsumoto, T.; Arakawa, T.; Kuroki, T. Functional and phenotypical activation of leucocytes in inflamed human colonic mucosa. *J. Gastroenterol. Hepatol.* **2008**, *12*, 809–814. [[CrossRef](#)]
69. Andresen, L.; Jørgensen, V.L.; Perner, A.; Hansen, A.; Eugen-Olsen, J.; Rask-Madsen, J. Activation of nuclear factor kappaB in colonic mucosa from patients with collagenous and ulcerative colitis. *Gut* **2005**, *54*, 503–509. [[CrossRef](#)] [[PubMed](#)]
70. Zhang, C.; Liu, L.-w.; Sun, W.-j.; Qin, S.-h.; Qin, L.-z.; Wang, X. Expressions of E-cadherin, p120ctn, β -catenin and NF- κ B in ulcerative colitis. *J. Huazhong Univ. Sci. Technol.* **2015**, *35*, 368–373. [[CrossRef](#)]
71. Sharma, R.; Young, C.; Neu, J. Molecular modulation of intestinal epithelial barrier: Contribution of microbiota. *J. Biomed. Biotechnol.* **2010**, *2010*, 305879. [[CrossRef](#)]
72. Kinnebrew, M.A.; Pamer, E.G. Innate immune signaling in defense against intestinal microbes. *Immunol. Rev.* **2012**, *245*, 113–131. [[CrossRef](#)] [[PubMed](#)]
73. Liu, T.; Zhang, L.; Joo, D.; Sun, S.-C. NF- κ B signaling in inflammation. *Signal Transduct. Target. Ther.* **2017**, *2*, 17023. [[CrossRef](#)] [[PubMed](#)]
74. Wang, Z.; Li, S.; Cao, Y.; Tian, X.; Zeng, R.; Liao, D.-F.; Cao, D. Oxidative stress and carbonyl lesions in ulcerative colitis and associated colorectal cancer. *Oxidative Med. Cell. Longev.* **2016**, *2016*, 9875298. [[CrossRef](#)] [[PubMed](#)]
75. Aldars-García, L.; Marin, A.C.; Chaparro, M.; Gisbert, J.P. The Interplay between immune system and microbiota in inflammatory bowel disease: A narrative review. *Int. J. Mol. Sci.* **2021**, *22*, 3076. [[CrossRef](#)]
76. Juge, N. Relationship between mucosa-associated gut microbiota and human diseases. *Biochem. Soc. Trans.* **2022**, *50*, 1225–1236. [[CrossRef](#)] [[PubMed](#)]
77. Zhu, H.; Li, Y.R. Oxidative stress and redox signaling mechanisms of inflammatory bowel disease: Updated experimental and clinical evidence. *Exp. Biol. Med.* **2012**, *237*, 474–480. [[CrossRef](#)] [[PubMed](#)]
78. Albenberg, L.; Esipova, T.V.; Judge, C.P.; Bittinger, K.; Chen, J.; Laughlin, A.; Grunberg, S.; Baldassano, R.N.; Lewis, J.D.; Li, H.; et al. Correlation between intraluminal oxygen gradient and radial partitioning of intestinal microbiota. *Gastroenterology* **2014**, *147*, 1055. [[CrossRef](#)]
79. Henson, M.A.; Phalak, P. Microbiota dysbiosis in inflammatory bowel diseases: In silico investigation of the oxygen hypothesis. *BMC Syst. Biol.* **2017**, *11*, 145. [[CrossRef](#)] [[PubMed](#)]
80. Miquel, S.; Leclerc, M.; Martin, R.; Chain, F.; Lenoir, M.; Raguideau, S.; Hudault, S.; Bridonneau, C.; Northen, T.; Bowen, B.; et al. Identification of metabolic signatures linked to anti-inflammatory effects of *Faecalibacterium prausnitzii*. *mBio* **2015**, *6*, e00300-15. [[CrossRef](#)] [[PubMed](#)]
81. Rigottier-Gois, L. Dysbiosis in inflammatory bowel diseases: The oxygen hypothesis. *ISME J.* **2013**, *7*, 1256–1261. [[CrossRef](#)]
82. McQuade, R.M.; Carbone, S.E.; Stojanovska, V.; Rahman, A.; Gwynne, R.M.; Robinson, A.M.; Goodman, C.A.; Bornstein, J.C.; Nurgali, K. Role of oxidative stress in oxaliplatin-induced enteric neuropathy and colonic dysmotility in mice. *Br. J. Pharmacol.* **2016**, *173*, 3502–3521. [[CrossRef](#)]
83. Nurgali, K.; Qu, Z.; Hunne, B.; Thacker, M.; Pontell, L.; Furness, J.B. Morphological and functional changes in guinea-pig neurons projecting to the ileal mucosa at early stages after inflammatory damage. *J. Physiol.* **2011**, *589*, 325–339. [[CrossRef](#)]

84. Chandrasekharan, B.; Anitha, M.; Blatt, R.; Shahnavaz, N.; Kooby, D.; Staley, C.; Mwangi, S.; Jones, D.P.; Sitaraman, S.V.; Srinivasan, S. Colonic motor dysfunction in human diabetes is associated with enteric neuronal loss and increased oxidative stress. *Neurogastroenterol. Motil.* **2011**, *23*, 131–138. [[CrossRef](#)]
85. Qu, Z.-D.; Thacker, M.; Castelucci, P.; Bagyánszki, M.; Epstein, M.L.; Furness, J.B. Immunohistochemical analysis of neuron types in the mouse small intestine. *Cell Tissue Res.* **2008**, *334*, 147–161. [[CrossRef](#)] [[PubMed](#)]
86. Robinson, A.M.; Miller, S.; Payne, N.; Boyd, R.; Sakkal, S.; Nurgali, K. Neuroprotective potential of mesenchymal stem cell-based therapy in acute stages of TNBS-induced colitis in guinea-pigs. *PLoS ONE* **2015**, *10*, e0139023. [[CrossRef](#)] [[PubMed](#)]
87. Robinson, A.M.; Sakkal, S.; Park, A.; Jovanovska, V.; Payne, N.; Carbone, S.E. Mesenchymal stem cells and conditioned medium avert enteric neuropathy and colon dysfunction in guinea pig TNBS-induced colitis. *Am. J. Physiol.-Gastrointest. Liver Physiol.* **2014**, *307*, G1115–G1129. [[CrossRef](#)] [[PubMed](#)]
88. Boyer, L.; Sidpra, D.; Jevon, G.; Buchan, A.M.; Jacobson, K. Differential responses of VIPergic and nitrergic neurons in paediatric patients with Crohn's disease. *Auton. Neurosci.* **2007**, *134*, 106–114. [[CrossRef](#)] [[PubMed](#)]
89. Winston, J.H.; Li, Q.; Sarna, S.K. Paradoxical regulation of ChAT and nNOS expression in animal models of Crohn's colitis and ulcerative colitis. *Am. J. Physiol.-Gastrointest. Liver Physiol.* **2013**, *305*, 295–302. [[CrossRef](#)]
90. Bagyanszki, M.; Bodi, N. Diabetes-related alterations in the enteric nervous system and its microenvironment. *World J. Diabetes* **2012**, *3*, 80–93. [[CrossRef](#)] [[PubMed](#)]
91. Boughton-Smith, N.K.; Evans, S.M.; Whittle, B.J.R.; Moncada, S.; Hawkey, C.J.; Cole, A.T.; Balsitis, M. Nitric oxide synthase activity in ulcerative colitis and Crohn's disease. *Lancet* **1993**, *342*, 2–340. [[CrossRef](#)]
92. Furness, J.B.; Li, Z.S.; Young, H.M.; Förstermann, U. Nitric oxide synthase in the enteric nervous system of the guinea-pig: A quantitative description. *Cell Tissue Res.* **1994**, *277*, 139–149. [[CrossRef](#)] [[PubMed](#)]
93. D'Inca, R.; Cardin, R.; Benazzato, L.; Angriman, I.; Martines, D.; Sturniolo Giacomo, C. Oxidative DNA damage in the mucosa of ulcerative colitis increases with disease duration and dysplasia. *Inflamm. Bowel Dis.* **2006**, *10*, 23–27. [[CrossRef](#)]
94. Fang, J.; Seki, T.; Tsukamoto, T.; Qin, H.; Yin, H.; Liao, L.; Nakamura, H.; Maeda, H. Protection from inflammatory bowel disease and colitis-associated carcinogenesis with 4-vinyl-2,6-dimethoxyphenol (canolol) involves suppression of oxidative stress and inflammatory cytokines. *Carcinogenesis* **2013**, *34*, 2833–2841. [[CrossRef](#)]
95. Tell, G.; Quadrioglio, F.; Tiribelli, C.; Kelley, M.R. The many functions of APE1/Ref-1: Not only a DNA repair enzyme. *Antioxid. Redox Signal.* **2009**, *11*, 601–619. [[CrossRef](#)]
96. Kelley, M.R.; Georgiadis, M.M.; Fishel, M.L. APE1/Ref-1 Role in redox signaling: Translational applications of targeting the redox function of the DNA repair/redox protein APE1/Ref-1. *Curr. Mol. Pharmacol.* **2012**, *5*, 36–53. [[CrossRef](#)] [[PubMed](#)]
97. Walker, L.J.; Robson, C.N.; Black, E.; Gillespie, D.; Hickson, I.D. Identification of residues in the human DNA repair enzyme HAP1 (Ref-1) that are essential for redox regulation of Jun DNA binding. *Mol. Cell. Biol.* **1993**, *13*, 5370–5376.
98. Xanthoudakis, S.; Miao, G.; Wang, F.; Pan, Y.C.; Curran, T. Redox activation of Fos-Jun DNA binding activity is mediated by a DNA repair enzyme. *EMBO J.* **1992**, *11*, 3323–3335. [[CrossRef](#)]
99. Georgiadis, M.M.; Luo, M.; Gaur, R.K.; Delaplane, S.; Li, X.; Kelley, M.R. Evolution of the redox function in mammalian Apurinic/aprimidinic endonuclease. *Mutat. Res.* **2008**, *643*, 54–63. [[CrossRef](#)] [[PubMed](#)]
100. Wilson, D.M.; Barsky, D. The major human abasic endonuclease: Formation, consequences and repair of abasic lesions in DNA. *Mutat. Res./DNA Repair* **2001**, *485*, 283–307. [[CrossRef](#)] [[PubMed](#)]
101. Krokan, H.E.; Bjørås, M. Base Excision Repair. *Cold Spring Harb. Perspect. Biol.* **2013**, *5*, 125. [[CrossRef](#)]
102. Franchi, L.P.; de Freitas Lima, J.E.B.; Piva, H.L.; Tedesco, A.C. The redox function of apurinic/aprimidinic endonuclease 1 as key modulator in photodynamic therapy. *J. Photochem. Photobiol. B Biol.* **2020**, *211*, 111–992. [[CrossRef](#)] [[PubMed](#)]
103. Mantha, A.K.; Sarkar, B.; Tell, G. A short review on the implications of base excision repair pathway for neurons: Relevance to neurodegenerative diseases. *Mitochondrion* **2014**, *16*, 38–49. [[CrossRef](#)] [[PubMed](#)]
104. Evans, A.R.; Limp-Foster, M.; Kelley, M.R. Going APE over ref-1. *Mutat. Res./DNA Repair* **2000**, *461*, 83–108. [[CrossRef](#)]
105. Mol, C.D.; Izumi, T.; Mitra, S.; Tainer, J.A. DNA-bound structures and mutants reveal abasic DNA binding by APE1 DNA repair and coordination. *Nature* **2000**, *403*, 451. [[CrossRef](#)]
106. Krokan, H.E.; Standal, R.; Slupphaug, G. DNA glycosylases in the base excision repair of DNA. *Biochem. J.* **1997**, *325*, 1–16. [[CrossRef](#)] [[PubMed](#)]
107. Flaherty, D.M.; Monick, M.M.; Carter, A.B.; Peterson, M.W.; Hunninghake, G.W. Oxidant-mediated increases in redox factor-1 nuclear protein and activator protein-1 DNA binding in asbestos-treated macrophages. *J. Immunol.* **2002**, *168*, 5675. [[CrossRef](#)] [[PubMed](#)]
108. Cardoso, A.A.; Jiang, Y.; Luo, M.; Reed, A.M.; Shahda, S.; He, Y.; Maitra, A.; Kelley, M.R.; Fishel, M.L. APE1/Ref-1 regulates STAT3 transcriptional activity and APE1/Ref-1-STAT3 dual-targeting effectively inhibits pancreatic cancer cell survival. *PLoS ONE* **2012**, *7*, e47462. [[CrossRef](#)]
109. Yang, S.; Misner, B.J.; Chiu, R.J.; Meyskens, J.F.L. Redox effector factor-1, combined with reactive oxygen species, plays an important role in the transformation of JB6 cells. *Carcinogenesis* **2007**, *28*, 2382–2390. [[CrossRef](#)] [[PubMed](#)]
110. Wang, D.; Luo, M.; Kelley, M.R. Human apurinic endonuclease 1 (APE1) expression and prognostic significance in osteosarcoma: Enhanced sensitivity of osteosarcoma to DNA damaging agents using silencing RNA APE1 expression inhibition. *Mol. Cancer Ther.* **2004**, *3*, 679–686. [[CrossRef](#)]

111. Luo, M.; Delaplane, S.; Jiang, A.; Reed, A.; He, Y.; Fishel, M.; Nyland, R.L.; Borch, R.F.; Qiao, X.; Georgiadis, M.M.; et al. Role of the multifunctional DNA repair and redox signaling protein APE1/Ref-1 in cancer and endothelial cells: Small-molecule inhibition of the redox function of APE1. *Antioxid. Redox Signal.* **2008**, *10*, 1853–1867. [[CrossRef](#)]
112. Sardar Pasha, S.P.B.; Sishitla, K.; Sulaiman, R.S.; Park, B.; Shetty, T.; Shah, F.; Fishel, M.L.; Wikel, J.H.; Kelley, M.R.; Corson, T.W. Ref-1/APE1 inhibition with novel small molecules blocks ocular neovascularization. *J. Pharmacol. Exp. Ther.* **2018**, *367*, 108–118. [[CrossRef](#)]
113. Mijit, M.; Caston, R.; Gampala, S.; Fishel, M.L.; Fehrenbacher, J.; Kelley, M.R. APE1/Ref-1—One target with multiple indications: Emerging aspects and new directions. *J. Cell. Signal.* **2021**, *2*, 151–161.
114. Fung, H.; Demple, B. A vital role for APE1/Ref1 protein in repairing spontaneous DNA damage in human cells. *Mol. Cell* **2005**, *17*, 463–470. [[CrossRef](#)] [[PubMed](#)]
115. Shah, F.; Logsdon, D.; Messmann, R.A.; Fehrenbacher, J.C.; Fishel, M.L.; Kelley, M.R. Exploiting the Ref-1-APE1 node in cancer signaling and other diseases: From bench to clinic. *NPJ Precis. Oncol.* **2017**, *1*, 19. [[CrossRef](#)] [[PubMed](#)]
116. McNeill, D.R.; Wilson, D.M., III. A dominant-negative form of the major human abasic endonuclease enhances cellular sensitivity to laboratory and clinical DNA-damaging agents. *Mol. Cancer Res.* **2007**, *5*, 61–70. [[CrossRef](#)] [[PubMed](#)]
117. Vasko, M.R.; Guo, C.; Thompson, E.L.; Kelley, M.R. The repair function of the multifunctional DNA repair/redox protein APE1 is neuroprotective after ionizing radiation. *DNA Repair* **2011**, *10*, 942–952. [[CrossRef](#)] [[PubMed](#)]
118. Kelley, M.R.; Jiang, Y.; Guo, C.; Reed, A.; Meng, H.; Vasko, M.R. Role of the DNA base excision repair protein, APE1 in cisplatin, oxaliplatin, or carboplatin induced sensory neuropathy. *PLoS ONE* **2014**, *9*, 106. [[CrossRef](#)]
119. Liu, H.; Liu, X.; Zhang, C.; Zhu, H.; Xu, Q.; Bu, Y.; Lei, Y. Redox imbalance in the development of colorectal cancer. *J. Cancer* **2017**, *8*, 1586–1597. [[CrossRef](#)]
120. Chang, I.Y.; Kim, J.N.; Maeng, Y.H.; Yoon, S.P. Apurinic/aprimidinic endonuclease 1, the sensitive marker for DNA deterioration in dextran sulfate sodium-induced acute colitis. *Redox Rep.* **2013**, *18*, 165–173. [[CrossRef](#)] [[PubMed](#)]
121. Thakur, S.; Sarkar, B.; Cholia, R.P.; Gautam, N.; Dhiman, M.; Mantha, A.K. APE1/Ref-1 as an emerging therapeutic target for various human diseases: Phytochemical modulation of its functions. *Exp. Mol. Med.* **2014**, *46*, 106. [[CrossRef](#)] [[PubMed](#)]
122. Jedinak, A.; Dudhgaonkar, S.; Kelley, M.R.; Sliva, D. Apurinic/Apyrimidinic endonuclease 1 regulates inflammatory response in macrophages. *Anticancer. Res.* **2011**, *31*, 379–385.
123. O'Hara, A.M.; Bhattacharyya, A.; Mifflin, R.C.; Smith, M.F.; Ryan, K.A.; Scott, K.G.E.; Naganuma, M.; Casola, A.; Izumi, T.; Mitra, S.; et al. Interleukin-8 induction by helicobacter pylori; in gastric epithelial cells is dependent on apurinic/aprimidinic endonuclease-1/redox factor-1. *J. Immunol.* **2006**, *177*, 7990. [[CrossRef](#)] [[PubMed](#)]
124. Lee, S.H.; Kwon, J.E.; Cho, M.L. Immunological pathogenesis of inflammatory bowel disease. *Intest. Res.* **2018**, *16*, 26–42. [[CrossRef](#)] [[PubMed](#)]
125. Nath, S.; Roychoudhury, S.; Kling, M.J.; Song, H.; Biswas, P.; Shukla, A.; Band, H.; Joshi, S.; Bhakat, K.K. The extracellular role of DNA damage repair protein APE1 in regulation of IL-6 expression. *Cell Signal* **2017**, *39*, 18–31. [[CrossRef](#)]
126. Xie, J.Y.; Li, M.X.; Xiang, D.B.; Mou, J.H.; Qing, Y.; Zeng, L.L.; Yang, Z.Z.; Guan, W.; Wang, D. Elevated expression of APE1/Ref-1 and its regulation on IL-6 and IL-8 in bone marrow stromal cells of multiple myeloma. *Clin. Lymphoma Myeloma Leuk.* **2010**, *10*, 385–393. [[CrossRef](#)]
127. Han, J.; Theiss, A.L. STAT3: Friend or foe in colitis and colitis-associated cancer? *Inflamm. Bowel Dis.* **2014**, *20*, 2405–2411. [[CrossRef](#)] [[PubMed](#)]
128. Haga, S.; Terui, K.; Zhang, H.Q.; Enosawa, S.; Ogawa, W.; Inoue, H.; Okuyama, T.; Takeda, K.; Akira, S.; Ogino, T.; et al. STAT3 protects against Fas-induced liver injury by redox-dependent and -independent mechanisms. *J. Clin. Investig.* **2003**, *112*, 989–998. [[CrossRef](#)]
129. Loeb, K.R.; Loeb, L.A. Genetic instability and the mutator phenotype : Studies in ulcerative colitis. *Am. J. Pathol.* **1999**, *154*, 1621–1626. [[CrossRef](#)] [[PubMed](#)]
130. Brentnall, T.A.; Crispin, D.A.; Bronner, M.P.; Cherian, S.P.; Hueffed, M.; Rabinovitch, P.S.; Rubin, C.E.; Haggitt, R.C.; Boland, C.R. Microsatellite instability in nonneoplastic mucosa from patients with chronic ulcerative colitis. *Cancer Res.* **1996**, *56*, 1237.
131. Kidane, D.; Chae, W.J.; Czochor, J.; Eckert, K.A.; Glazer, P.M.; Bothwell, A.L.M.; Sweasy, J.B. Interplay between DNA repair and inflammation, and the link to cancer. *Crit. Rev. Biochem. Mol. Biol.* **2014**, *49*, 116–139. [[CrossRef](#)] [[PubMed](#)]
132. Guo, H.H.; Loeb, L.A. Tumbling down a different pathway to genetic instability. *J. Clin. Investig.* **2003**, *112*, 1793–1795. [[CrossRef](#)] [[PubMed](#)]
133. Seril, D.N.; Liao, J.; Yang, G.-Y.; Yang, C.S. Oxidative stress and ulcerative colitis-associated carcinogenesis: Studies in humans and animal models. *Carcinogenesis* **2003**, *24*, 353–362. [[CrossRef](#)]
134. Kim, E.R.; Chang, D.K. Colorectal cancer in inflammatory bowel disease: The risk, pathogenesis, prevention and diagnosis. *World J. Gastroenterol.* **2014**, *20*, 9872–9881. [[CrossRef](#)] [[PubMed](#)]
135. Nyland, R.L.; Luo, M.; Kelley, M.R.; Borch, R.F. Design and synthesis of novel quinone inhibitors targeted to the redox function of Apurinic/aprimidinic endonuclease 1/redox enhancing factor-1 (APE1/Ref-1). *J. Med. Chem.* **2010**, *53*, 1200–1210. [[CrossRef](#)] [[PubMed](#)]
136. Caston, R.A.; Gampala, S.; Armstrong, L.; Messmann, R.A.; Fishel, M.L.; Kelley, M.R. The multifunctional APE1 DNA repair-redox signaling protein as a drug target in human disease. *Drug Discov. Today* **2021**, *26*, 218–228. [[CrossRef](#)]

137. Lally, D.; Brigell, M.; Withers, B.; Kolli, A.; Rahmani, K.; Sooch, M.; Lazar, A.; Patel, R.; Kelley, M.R.; Boyer, D.; et al. Masked safety data from ZETA-1, an ongoing 24-week Phase 2 clinical trial of APX3330, an oral therapeutic being developed for the treatment of diabetic retinopathy. In Proceedings of the Retina World Congress 2022, Fort Lauderdale, FL, USA, 12–15 May 2022.
138. Zhong, C.; Xu, M.; Wang, Y.; Xu, J.; Yuan, Y. An APE1 inhibitor reveals critical roles of the redox function of APE1 in KSHV replication and pathogenic phenotypes. *PLoS Pathog.* **2017**, *13*, e1006289. [[CrossRef](#)]
139. Shimizu, N.; Sugimoto, K.; Tang, J.; Nishi, T.; Sato, I.; Hiramoto, M.; Aizawa, S.; Hatakeyama, M.; Ohba, R.; Hatori, H.; et al. High-performance affinity beads for identifying drug receptors. *Nat. Biotechnol.* **2000**, *18*, 877. [[CrossRef](#)]
140. Zou, G.-M.; Maitra, A. Small-molecule inhibitor of the AP endonuclease 1/REF-1 E3330 inhibits pancreatic cancer cell growth and migration. *Mol. Cancer Ther.* **2008**, *7*, 2012–2021. [[CrossRef](#)] [[PubMed](#)]
141. Rai, R.; Dawodu, O.I.; Johnson, S.M.; Vilseck, J.Z.; Kelley, M.R.; Ziarek, J.J.; Georgiadis, M.M. Chemically induced partial unfolding of the multifunctional Apurinic/aprimidinic endonuclease 1. *bioRxiv* **2023**. [[CrossRef](#)]
142. Shah, Y.M. The role of hypoxia in intestinal inflammation. *Mol. Cell. Pediatr.* **2016**, *3*, 1. [[CrossRef](#)]
143. Xue, X.; Ramakrishnan, S.; Anderson, E.; Taylor, M.; Zimmermann, E.M.; Spence, J.R.; Huang, S.; Greenson, J.K.; Shah, Y.M. Endothelial PAS domain protein 1 activates the inflammatory response in the intestinal epithelium to promote colitis in mice. *Gastroenterology* **2013**, *145*, 831–841. [[CrossRef](#)] [[PubMed](#)]
144. Shahda, S.; Lakhani, N.J.; O'Neil, B.; Rasco, D.W.; Wan, J.; Mosley, A.L.; Liu, H.; Kelley, M.R.; Messmann, R.A. A phase I study of the APE1 protein inhibitor APX3330 in patients with advanced solid tumors. *J. Clin. Oncol.* **2019**, *37*, 3097. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.